

Northern Michigan FruitNet 2015 Northwest Michigan Horticultural Research Center

Weekly Update

April 17, 2015

CALENDAR OF EVENTS

2015

- 4/22** **2015 Tree Fruit IPM Kickoff**
- 4/30** **Sweet Cherry Pruning Demonstration**
Various locations
- 5/5-7/14** **Leelanau County IPM Updates**
Bardenhagen Farm
- 5/5-7/14** **Grand Traverse County IPM Updates**
Wunsch Farm
- 5/6-7/15** **Antrim County IPM Updates**
Jack White Farms
- 5/6-7/15** **Benzie County IPM Updates**
Blaine Christian Church

2015 Tree Fruit IPM Kick-off

Northwest Michigan Horticultural Research Center

Please join Michigan State University Extension and AgBioResearch for the annual Tree Fruit IPM Kickoff at the Northwest Michigan Horticultural Research Center on Wednesday, **April 22 from 4:00 – 7:00 PM**. This year, we are pleased to host Drs. George Sundin and Larry Gut from the MSU Fruit Team to provide the latest information in cherry and apple disease and insect pest management. Pesticide label changes and updates will also be discussed. This meeting is free of charge and no registration is necessary. Refreshments will be provided, and three pesticide recertification credits and three certified crop advisor credits are available. Please bring your tough insect and disease questions to this meeting as the experts will be here to answer them!

Following the Kick-off presentations, the Leelanau Horticultural Society and Grand Traverse Fruit Growers' Council will host a short roundtable discussion on recent changes within the CherrCo, Inc. Jim Jensen, CherrCo, Inc President, will be on hand to answer growers' questions about the organization heading into 2015.

- 3:45** **Welcome and refreshments**
- 4:00 – 4:30** **Cherry Disease Management Blast-off for 2015**
Dr. George Sundin, Dept. Plant Pathology, MSU
- 4:30 – 5:00** **Scab, Fuzz, and Ooze: Managing apple diseases in 2015**
Dr. George Sundin, Dept. Plant Pathology, MSU
- 5:00 – 5:30** **Introducing: Pheromone Aerosol Emitters for Codling Moth**
Dr. Larry Gut, Dept. Entomology, MSU
- 5:30 – 6:00** **Preparing for Spotted Wing Drosophila Management in 2015**
Dr. Larry Gut, Dept. Entomology, MSU
- 6:00 – 6:15** **Break**
- 6:15 – 7:00** **Pesticide label changes and updates**
Emily Pochubay, MSU Extension
- Fill out sheets for Pesticide Recertification, 1C private and commercial core (3 credits available) and Certified Crop Advisor (3 credits available).
- 7:00** **Changes to CherrCo discussion**
Jim Jenson, CherrCo, Inc. President

SWEET CHERRY PRUNING DEMONSTRATION: April 30, 2015

Join Dr. Greg Lang for a pruning demonstration tour on April 30th. The tour will begin at King Orchards' new high-density sweet cherry block behind their farm market on US 31; they have many new varieties trained to a tall spindle axe (TSA). The group will have lunch at Pearl's New Orleans Kitchen in Elk Rapids. We will travel to Greg Williams' farm in Cedar where we will prune sweet cherries trained to upright fruiting offshoot (UFO) and TSA systems. The tour will end at the Northwest Michigan Hort. Research Center (NWMHRC) where Dr. Lang will introduce us to his new sweet cherry training manual (<http://www.hrt.msu.edu/greg-lang/pg3>). We wrap up the day with a review of these systems in the field. Please join us for a great day of pruning—we are sure to have great weather with Dr. Lang on hand. For additional information, please contact Nikki Rothwell or Emily Pochubay at 231-946-1510.

- 10:30-11:30 King Orchards
Creswell Road (behind King Orchards Farm Market on US 31)
- 11:30-12:15 Lunch at Pearl's New Orleans Kitchen
617 Ames Street, Elk Rapids, MI 49629
- 12:15-1:15 Travel to Greg Williams Orchards
- 1:15-2:15 Greg Williams Orchards
Novak Road, Cedar, MI

2:15-2:45 Travel to NWMHRC

2:45-4:00 Overview of new sweet cherry training manual and in-field systems pruning
6686 S. Center Highway, Traverse City, MI 49684

2015 Tree Fruit IPM Update Series

Emily Pochubay and Nikki Rothwell
Michigan State University Extension

Tree Fruit IPM Updates beginning the first week of May through mid-July (as needed) will highlight management of the season's current potential pest challenges dictated by weather and pest biology. Attendees are encouraged to bring examples of pests and damage found on the farm to these workshops for identification and discussion. Workshops will be held weekly in Leelanau and Grand Traverse counties and bi-weekly in Antrim and Benzie Counties. Tree fruit growers are welcome to attend meetings at any of the locations and times that are most convenient (see below). These workshops are free and do not require registration. Certified crop advisor continued education credits (two per meeting) and pesticide recertification credits (two per meeting) will be available. We are looking forward to seeing you in a few weeks! For more information, please contact Emily Pochubay (pochubay@msu.edu), 231-946-1510.

IPM Update Dates, Times, and Locations

Leelanau County

Location: Jim and Jan Bardenhagen, 7881 Pertner Rd, Suttons Bay
Dates: May: 5, 12, 19, 26; June: 2, 9, 16, 23, 30; July: 14
Time: 12PM – 2PM

Grand Traverse County

Location: Wunsch Farms, Phelps Road Packing Shed, Old Mission
Dates: May: 5, 12, 19, 26; June: 2, 9, 16, 23, 30; July: 14
Time: 3PM – 5PM

Antrim County

Location: Jack White Farms, 10877 US-31, Williamsburg (is not correct in Google Maps) *North of Camelot Inn and South of Elk Rapids on the southeast side of US-31*
May: 6, 20; June: 3, 17; July: 1, 15
Time: 10AM – 12PM

Benzie County

Location: Blaine Christian Church, 7018 Putney Rd, Arcadia, MI 49613
May: 6, 20; June: 3, 17; July: 1, 15
Time: 2PM – 4PM

REQUEST FOR APPLE GROWER COOPERATORS

Emily Pochubay, MSU Extension

The NWMHRC is looking for apple grower cooperators for three projects (see below) in the coming season. Please see additional information on each of the projects below. If you are interested in participating in any of these projects or if you would like more information, please contact Emily Pochubay (pochubay@msu.edu), 231-945-1510. Thank you for your continued support!

1. Apple scab spore monitoring

The NWMHRC is hoping to monitor for apple scab spore release during primary infection this spring to assist the Enviro-weather apple scab model and local apple growers with identifying the end of primary infection for apple scab. For this project, we need an apple block <10 miles of the research station (6686 S. Center HWY) that had some degree of apple scab infected tissue last season and can be easily accessed after each rain event to collect spore collecting rods.

2. San Jose scale monitoring

Apple blocks with known San Jose scale populations are needed to monitor for male emergence/flight as well as crawler activity. Apple orchards in Leelanau county, Old Mission/Grand Traverse, and Antrim county would be preferable for this project as we would like to observe the relationship of San Jose scale behavior in sweet cherries and apples; we are currently scouting in sweet cherries in those three aforementioned regions of NW Michigan.

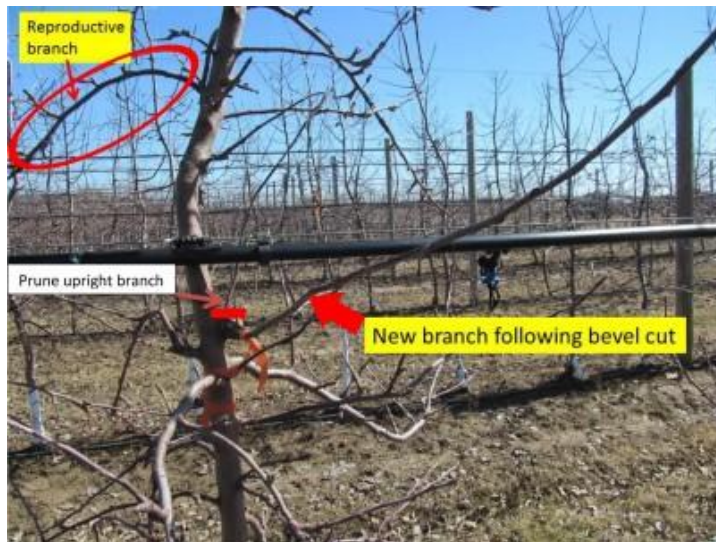
3. Apple maggot monitoring

In 2014, the NWMHRC observed detections of cherry fruit fly using different commercially available sticky traps and found that trap type influences first detections and the number of cherry fruit flies captured. In 2015, we would like to observe this possible effect on apple maggot catches. To test this hypothesis, we are looking for five apple blocks in northwest MI counties (Leelanau, Old Mission/Grand Traverse, Antrim, Benzie, Manistee).

IMPORTANCE OF RECYCLING ON TALL SPINDLE APPLE PLANTINGS

A key to successful canopy development is to retain and encourage productive growth by recycling vigorous branches in a tall spindle apple planting.

Posted on **April 3, 2015, MSUE News**, by [Ron Perry](#), Michigan State University Extension, Department of Horticulture



Tall spindle apple planting showing a reproductive branch, a new branch following a bevel cut, and where to prune an upright branch.

Having ventured into a few tall spindle apple orchards over the last year, I am reminded that I need to remind growers new to these high density systems that a key to canopy development and success is how to retain and encourage productive growth.

One mistake I continue to see are trees in which too many branches are being retained in the canopy, which compete with vertical development of leaders in trees aged 2-4. Just because you purchased trees with a lot of “feathers” does not mean you can retain them all. Too many will stymie leader development. If this happens, canopy volume suffers and so does yield on a per acre basis. Unfortunately, once trees get into this state, it is difficult to reverse the trend.

Once trees get into their fifth and sixth growing season, recycling vigorous branches is very important. Generally, the recommended protocol is to prune or remove those branches where caliper is 50 percent or larger compared to the leader from which it grows. In particular, remove those that have an upright trajectory (see thick dash mark in image). They will usually remain vigorous and vegetative. Instead, retain branches that are weak and possess spurs, meaning they are more “reproductive” (see image).

[Michigan State University Extension](#) recommends growers prune or remove two to three branches per tree. Leave a stub of 3-4 inches. When making the pruning cut, make it so that the cut is oblique with the cut surface facing upward. We call these “bevel” or “Dutch” cuts ([see YouTube video on pruning tall spindle apple branches](#)). Pruning in this manner will discourage adventitious shoots from forcing on the upward side of the branch. During the growing season, follow-up with training adventitious shoots downward (below horizontal) to slow their growth and encourage flower bud induction in mid-June to mid-July.

Spend time during mid-June to mid-July bending branches down using elastic bands and it will reward you many times over in subsequent seasons with productive trees.

For more information, see the YouTube video “[Pruning Tall Spindle Apples: Recycling Strong Branches](#).”

Thanks to Denise Ruwersma, research assistant, for working on the SSCD project for videography and plot canopy management.

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WINTER GRAPE BUD INJURY FOR THE SECOND YEAR IN A ROW

Severe winter cold during the 2014-15 winter has injured grape buds for the second year in a row in Michigan's Grand Traverse region.

Posted on **April 3, 2015, MSUE News**, by [Duke Elsner](#), Michigan State University Extension, and Paolo Sabbatini, MSU Department of Horticulture



The 2014-15 winter injured grape buds in northern Michigan vineyards. Photo credit: Duke Elsner, MSU Extension

A year ago when reports came out on severe winter kill to wine grape buds in northwest Michigan, everyone assumed that such unprecedented cold would not likely happen again. Unfortunately, the assumption was wrong. The 2014-15 winter in the Midwest and Eastern United States was once again impacted by the polar vortex – extreme cold temperatures that normally float around the poles year-round, but “thanks” to climate change these are unfortunately propagated farther, transporting warmer air to the North Pole and polar air into lower latitudes, directly into our vineyards.

Many sites in Michigan's Grand Traverse region were hit with numerous evenings of extremely cold temperatures in February and March. The [Michigan State University Enviro-weather](#) system recorded temperatures at -15 degrees Fahrenheit or lower on six dates from Feb. 15 through March 5 on [Old Mission Peninsula](#) in Grand Traverse County, with -19.9 F as the coldest reading on Feb. 20. [East Leland](#), Michigan in Leelanau County also recorded six nights below -15 F, with four of those nights below -20 F. The [Northwest Michigan Horticultural Research Station](#) (NWMHRS) in Leelanau County only reported -10 F or lower four times, with the lowest temperature at -16 F on Feb. 20.

The minimum low temperatures at East Leland, Michigan and the NWMHRS were even lower than temperatures experienced in early 2014. To make matters worse, there was far less snow cover during this year's cold events, so more of the wood and buds of the vines were fully exposed to the cold.

This year, the strategy for coping with the extensive winter injury should start with delaying pruning as long as possible during the dormant period. The delay should be used to assess the extent of winter injury and then adjusting the pruning strategies (and spring-summer vineyard practices) in relation to bud and vine damage and mortality levels. Therefore, before pruning, grape growers should carefully evaluate each cultivar for bud damage.

Thirty-one varieties of wine grapes in the NWMHRS experimental vineyard were sampled between March 16 and 31 by collecting a minimum of 10 1-year-old canes per variety. Buds and the vascular tissues of each cane were carefully sectioned with a razor blade and examined under magnification to look for indications of cold injury. The results were as expected, with *Vitis vinifera* varieties showing high mortality to buds and many instances of injury to the canes as well. The cane injury was localized, occurring just above where the winter snow cover had protected the wood; the tissues several inches above this zone looked much healthier.

Until further into spring and the beginning of bud swell, it will be difficult to tell if the injured region of the canes will be able to recover enough to support the growth of shoots higher in the vines. Shoots collapsing during the growing season up to veraison are symptomatic of trunks and canes damage.

Looking just at the condition of the buds, Riesling showed a 61 percent primary bud survival rate and Chardonnay at 66 percent. Each had almost 80 percent of their secondary buds looking alive. Pinot Noir and Cabernet Franc both had about 30 percent live primary buds. These numbers may not mean much if the injured vascular tissues in the canes does turn out to be a problem. If so, all of the more distal buds or shoots may eventually die, and the effective bud survival rates would be very low. Reports of more severe levels of bud injury have been received from commercial vineyard sites that recorded much lower temperatures than the research center.

The hybrid wine grape varieties fared much better, but there were still significant levels of injury to buds. The best primary bud survival rates were found in St. Croix (93 percent) and Frontenac (92 percent). The worst injury occurred to Aromella (59 percent live primary buds) and Chambourcin (43 percent live primary buds). At this point in time, our estimates of bud injury may not be good indications of the potential yield of the varieties during the growing season ahead; a level of 70 percent primary bud damage does not necessarily correspond to a 70 percent reduction in yield due to the potential for fruit production by secondary buds and the possibility there could be a compensatory increase in the weight of clusters.

The 2014 and 2015 winters are a forceful reminder that cultivar choice and site selection are still the most important tools we have against low winter temperatures. While we need to prune vines to mitigate damage as best as possible for the 2015 growing season, we should also keep in mind the 2016 season and crop, and the effect our choices during pruning and training will have on it. Several viticultural strategies can mitigate the impact of winter cold, such as:

- The use of multiple trunks, sometime defined as "spare-parts viticulture."

- Always having replacement canes (suckers) growing under the vine, which is fundamentally important for grafted *Vinifera* cultivars.
- Covering the graft union with soil during the winter. Covering, and uncovering, vines every year is very labor intensive, but the most efficient technique to guarantee vines and fruiting canes survival for the following season.

In 2012, the USDA released a new [plant hardiness zone map](#) because of the need to be in line with the temperature increase around the United States. We are using the extra heat and the longer growing season to ripen cultivars that were impossible to grow only few decades ago in cool and cold climate viticulture. Unfortunately, this warming trend is also producing extreme winter cold temperatures (polar vortex), and going back to the basic methods of grapevine protection during the winter is our best option to keep a sustainable grape industry in the Midwestern and Eastern United States.

A complete listing of the varieties that were examined and the bud survival percentages will be released in an upcoming [Northern Michigan FruitNet report](#), or by request if you send an email to [Michigan State University Extension](#) small fruit educator [Erwin Elsner](#) at elsner@msu.edu.

The [MSU Enviro-weather](#) system can be used to check the low temperatures at dozens of locations in Michigan. See www.enviroweather.msu.edu for more information.

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DORMANT OIL FOR TREE FRUIT PEST MANAGEMENT IN 2015

Considerations for effective and safe use of dormant oil on tree fruit pests like San Jose scale.

Posted on **April 7, 2015, MSUE News**, by [Amy Irish-Brown](#), and Phil Schwallier, Michigan State University Extension

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[Michigan State University Extension](#) educators and growers around the state reported higher than typical San Jose scale numbers on apple fruit in bins in 2013 and somewhat in 2014. Dormant oil is an effective method to control San Jose scale and other tree fruit pests and should be considered for use in 2015 if the weather and growth stages are right.

The term “dormant” in dormant oil can be a little misleading as the more commonly used timing is delayed dormant, or as close to green tip as possible and up until the pink stage ([view apple growth stages](#)). Keep in mind oil and green tissue are not very compatible. Phytotoxic damage to buds, blossoms and fruitlets is possible, thus care must be taken to avoid weather extremes. Do not use oil sprays 48 hours before and after a frost event. Avoid using oils in very hot (over 85 degrees Fahrenheit) and humid conditions. Do not apply sulfur or Captan within several days of oil applications or phytotoxicity will occur.

Good coverage is important for all spray applications, especially when using oil to target San Jose scale where the treatment target is on and under bark scales where the overwintering scales are. A common way to use dormant oil is 2 gallons per 100 at 100 gallons water per acre at green tip with copper. Some growers will break up their oil applications into two sprays: once at green tip with copper and another at 1 gallon per 100 at tight cluster with Lorsban.

Rates need to be reduced as the oil is applied closer to pink. Spur and bud damage is a high risk at pink and should be down to no more than 1 gallon oil per 100. Late applications will offer the best help to control scale and mites and adding Lorsban or another insecticide will help control rosy aphids.

Different crop protection retailers have different products and they all work similarly. Be sure to work closely with your spray salesperson for the best recommendation and use of their products. Follow all label guidelines. Dormant oil needs to be applied with care and attention to the weather conditions before and after the application.

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TRANSFERRING AND PLACING HONEY BEES THIS SPRING

April and May are the times to transfer new honey bees to their hives and place their hives in an ideal location.

Posted on **April 9, 2015, MSUE News**, by [Ben Phillips](#), Michigan State University Extension



An apiary utilizing color combinations and unique hive orientations to reduce worker bees from drifting to the wrong colony. It is preferable to raise hives up off of the ground. Photo credit: Susy Morris, Flickr Creative Commons

Spring is the time to transfer your newly arrived package or nucleus bees (nucs) into their bigger home. The process is similar whether you are using top-bar hives, foundationless Warré

philosophies or traditional Langstroth colonies, though most commercial bee suppliers will be selling nucleus colonies with Langstroth frame dimensions. If you built a hive with different dimensions, then buying a package of bees might be a better choice.

Package bees

Packages are small cages full of worker bees from large, active colonies in southern states. It is common that these bees are from different colonies. Further, packages include a freshly mated queen protected in a cage. She is also from a different colony. So, nobody knows each other, and there is no comb or brood or eggs to keep everyone busy.

It is important to transfer this cage of confused bees into a prepared hive as soon as possible. A prepared hive should include frames, with or without foundation, and some food of some sort. Place the queen cage between some frames with the exit hole face down, making sure the screen is exposed so the bees can feed her through it. The workers need to become accustomed to her pheromones and will let her out after a few days.

Frames of honey and pollen, or 1:1 sugar water and pollen substitute will help jump-start these hungry bees. Packages are usually cheaper than nucleus colonies, and are easier to use in top-bar hives or other homemade hives with non-Langstroth dimensions.

Nucleus bees

Nucleus colonies, or nucs, are complete, tiny colonies that are generally available later in the season. They usually consist of four or five frames of honey pollen and brood cells in a small box along with a laying queen, eggs, brood, nurse bees and worker bees that are all related. That is why they are not available as early as package bees. All you need to do with a nucleus colony is put it someplace with good spring forage.

You have more time to transfer these bees to a full size hive because they have everything they need for the short term in the nuc. However, many suppliers sell them when they are very full and ready to expand, so make sure you transfer them before they outgrow their home. To transfer them, choose a home location for the hive and transfer the frames from the nuc into the middle of the home hive, or wherever you want the brood nest if you've built a top-bar hive with Langstroth dimensions. Be careful to get the queen into the new colony. Leave the nuc next to the home hive for a few days so that foraging workers can figure out all their family moved next door. As they build out new comb, you may consider feeding this new colony to give them a jump-start on getting everything ready to roll for the main nectar flow.

Do you want to provide pollination with your bees?

There are many new growers, small growers and community gardens in Michigan that desire pollination, but may not want to keep bees themselves. These types of pollination opportunities aren't usually picked up by the large commercial beekeepers, yet provide a great opportunity for small and mid-size beekeepers with two to 20 hives.

Barb Barton, along with John Hooper of the [Michigan Organic Food and Farm Alliance](#) (MOFFA) and Meghan Milbrath of the [Northern Bee Network](#) and [Michigan State University](#), have put together an excellent resource that aims to match farmers with an organic philosophy and like-minded beekeepers. Check out the [Healthy Food - Healthy Bees Connection](#).

Hive placement

When placing your hives for pollination or honey production, [Michigan State University Extension](#) recommends picking a spot that will be easy to access through the season for supering and mite monitoring. If you plan to overwinter them on site, then you may need to also consider accessibility during the snowy months in Michigan. Avoid placing colonies in low spots. These spots can make packing out heavy honey supers more difficult, and may be in a flood plain. Driving through the country at night with the windows down easily demonstrates the temperature and moisture gradients of low spots. Foggy and cool is not fun for bees. High and dry with a windbreak and access to direct sun from the south and east tends to warm bees more quickly for earlier flying.

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HONEY BEE PEST AND DISEASE DIAGNOSTIC SERVICES COULD BE COMING TO MSU

Michigan State University is asking beekeepers to weigh in on a potential honey bee pest and disease identification service.

Posted on **April 9, 2015, MSUE News**, by [Ben Phillips](#), Michigan State University Extension

Meghan Milbrath of the [Northern Bee Network](#) and [Michigan Pollinator Initiative](#), and [Walter Pett](#) of [Michigan State University Extension](#) are asking for your input on potential diagnostic services for pests and diseases in managed Michigan honey bee hives.

The survey is designed to give campus diagnosticians an idea of the demand and price that beekeepers would be willing to pay for processing samples.

Please click on the following link for the survey: [Diagnostics Interest Survey for Beekeepers](#)

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WHERE ELSE DO SPOTTED WING DROSOPHILA LIKE TO FEED?

New online resource provides a list of non-crop host plants of spotted wing Drosophila in North America.

Posted on **April 15, 2015, MSUE News**, by [Rufus Isaacs](#), Michigan State University Extension, Department of

Noncrop Host Plants of Spotted Wing Drosophila in North America

EM 9113 • April 2015

J. Lee, A. Dreves, R. Isaacs, G. Loeb, H. Thistlewood, and L. Brewer

Spotted wing Drosophila (SWD), *Drosophila suzukii*, is an invasive fly that lays eggs in ripening and ripe berries, and stone fruits. The developing larvae can make the fruit unmarketable, so this pest is a concern to producers, packers, processors, and distributors of these crops.

Landscapes surrounding fruit production fields often include hedgerows, adjacent field margins, and woody or riparian areas with ornamentals, unmanaged shrubs, vines, or other plants that also produce fruits. Noncrop habitats can meet the requirements that favor SWD adults and their natural enemies: food, shelter, shade, and humidity. In addition, many noncrop fruits can support developing larvae of SWD. As populations of SWD build in noncrop hosts, these areas can become "hot spots" from which SWD can move into fields as commercial fruits begin to ripen. In some regions, these plants are important for late season population buildup outside crop fields.

From this publication, commercial and backyard fruit growers and field advisors will learn which plants can serve as alternate egg-laying sites for SWD. This list of noncommercial fruits was developed from multiyear sampling to determine likely noncrop hosts for SWD larvae. Regional differences in the importance of each plant host may occur due to differences in environmental conditions. The list is not exhaustive but includes what is known at this time about plants commonly found in British Columbia, Washington, Oregon, California, Michigan, New York, and Florida. We expect this list to expand as more becomes known about noncrop hosts for SWD.



Blue elderberry



Cherry laurel

Thistlewood Ltd, Agriculture and Agri-Food Canada

Amy J. Dreves, © Oregon State University

Jana Lee, USDA-ARS Horticultural Crops Research Unit; Amy J. Dreves, Department of Crop and Soil Science, Oregon State University; Rufus Isaacs, Department of Entomology, Michigan State University; Greg Loeb, Department of Entomology, Cornell University; Howard Thistlewood, Pacific Agri-Food Research Centre, Agriculture and Agri-Food Canada; and Linda Brewer, Department of Horticulture, Oregon State University.



"Noncrop Host Plants of Spotted Wing Drosophila in North America" is available as a **free** PDF on the [MSU Spotted Wing Drosophila website](#).

A new guide to the non-crop host plants used by spotted wing Drosophila (SWD) has just been published by Oregon State University, compiling information from collections made in Michigan, New York, Florida, California, Oregon, Washington and British Columbia. The article titled "[Noncrop Host Plants of Spotted Wing Drosophila in North America](#)" is now posted as a free PDF on the [Factsheets](#) page of the [Michigan State University Spotted Wing Drosophila website](#).

This information combines what was learned from collections of fruit in wild and agricultural habitats since this pest arrived in North America seven years ago. The researchers, including [Steve Van Timmeren](#) from MSU's Department of Entomology and myself, collected fruit and then held it to see what insects emerged from them. This provided definitive information on whether SWD does use a specific host plant.

The information can be used by growers and others to consider which areas may be most likely to provide a reservoir of habitat for this pest, and would therefore pose the greatest risk as a reservoir for pest infestation. There is little information on the effectiveness of removing these non-crop host plants from a landscape, but high densities of these suitable hosts are expected to increase pressure from SWD.

Dr. Isaacs' work is funded in part by [MSU's AgBioResearch](#).

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WEB SITES OF INTEREST:

Insect and disease predictive information is available at:

<http://enviroweather.msu.edu/homeMap.php>

This issue and past issues of the weekly FruitNet report are posted on our website

<http://agbioresearch.msu.edu/nwmihort/faxnet.htm>

60 Hour Forecast

<http://www.agweather.geo.msu.edu/agwx/forecasts/fcst.asp?fileid=fous46ktvc>

Information on cherries is available at the new cherry website:

<http://www.cherries.msu.edu/>

Information on apples:

<http://apples.msu.edu/>

Fruit CAT Alert Reports has moved to MSU News

<http://news.msue.msu.edu>