- 1 <u>Changes in the distribution of Michigan crayfishes and the influence of invasive rusty</u>
- 2 <u>crayfish (*Faxonius rusticus*) on native crayfish substrate associations.</u>
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- 5 Kelley R. Smith¹
- 6 Brian M. Roth¹ ORCID: 0000-0002-3366-9080
- 7 Michael L. Jones¹
- 8 **Daniel B. Hayes**¹ ORCID: 0000-0002-8132-4749
- 9 <u>Seth J. Herbst²</u>
- 10
- 11 <u>Nicholas Popoff²</u>
- 12

¹Department of Fisheries and Wildlife, Michigan State University, 480 Wilson Road, East Lansing, MI, 48824

15 ²Michigan Department of Natural Resources, 525 W. Allegan St., Lansing, MI 48909

16 <u>Abstract:</u>

Invasive cravfishes have a strong negative effect on multiple trophic levels, including 17 18 other crayfishes. However, documentation of the spread of non-native crayfish species and 19 their impact on native crayfishes could be improved, particularly over large spatial scales 20 in stream ecosystems. We collected cravfish and quantified habitat at 461 stream sites 21 throughout Michigan in 2014-2016 and compared our collections to a historical account of 22 crayfish collections published in 1975. Our objectives were to: 1) Quantify the change in non-native rusty crayfish (Faxonius rusticus) and native species distributions from 1975 to 23 2014-16; 2) Quantify how rusty crayfish affect the habitat associations of native species in 24 25 Michigan streams; and 3) Determine the effectiveness of dipnets, our primary sampling method. We found all species in more watersheds compared to 1975, likely due in part to 26 increased sampling. However, we found rusty cravfish in 22 more HUC-8 watersheds than 27 28 in 1975, a larger increase than all other species. Habitat associations of native species also shift in the presence of rusty crayfish. In instances where native species co-occurred with 29 rusty crayfish, most obligate aquatic native species were found in less-preferred habitat 30 such as sand or macrophytes compared to cobble substrate when the species is in isolation. 31 Our results indicate a broad range expansion by rusty crayfish over the last 40 years, 32 suggesting that surveys of crayfish diversity and habitat occupancy should be more routine 33 to inform management of native crayfish species. 34

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38 Introduction:

Invasions of non-native species represent one of the most important issues facing native 39 40 species biodiversity and ecosystem sustainability. Aquatic ecosystems are particularly vulnerable to effects of invasions, and those systems that are prone to habitat modification or 41 disturbance are likely to be most susceptible to introductions of non-native species (Lozon and 42 43 MacIsaac 1997; Moyle and Light 1996). For the past few decades, scientists have debated the relative roles of non-native species and habitat modification as engines of global change 44 (Didham et al. 2007; Didham et al. 2005). While some consensus has emerged that both play 45 46 important roles, synergies between non-native species introductions and habitat modification represent a potent driver of ecosystem change. Crayfish invasions represent such a nexus. The 47 ability of crayfish to manipulate ecosystems has led them to be labeled 'ecosystem engineers' 48 (Carreira et al. 2014; Hobbs et al. 1989; Lodge and Lorman 1987; Momot 1995; Momot et al. 49 1978). Further, the burrowing and foraging behavior of many crayfish species can alter both the 50 abiotic and biotic habitat available for native species through hydrologic alterations and 51 52 macrophyte habitat destruction, respectively (Faller et al. 2016; Wilson et al. 2004).

Crayfish invasions are often followed by extensive ecosystem disruption at a number of 53 trophic levels, from primary producers such as periphyton and macrophytes, to top predators 54 such as fishes (Carreira et al. 2014; Ilheu et al. 2007; Kershner and Lodge 1995; Lodge and 55 Lorman 1987; Roth et al. 2006; Roth et al. 2007; Wilson et al. 2004). Thus, there is considerable 56 interest in the distribution of crayfishes, particularly related to non-native species. The spread of 57 some crayfish species over broad spatial and temporal scales and their effects in the ecosystem 58 59 are well-documented, particularly for *Procambarus clarkii* in Europe (e.g. Gherardi 2006), but the spread of most species has gone undocumented, at least over longer temporal scales (but see 60 Olden et al. 2006; Taylor and Redmer 1996). 61

Crayfishes can be a dominant component of freshwater ecosystems, but many species are 62 imperiled (Charlebois and Lamberti 1996; Huner and Lundquist 1995; Lodge et al. 1994; 63 Nystrom et al. 2006). In some aquatic ecosystems, crayfishes account for more biomass than all 64 other macroinvertebrates combined (Rabeni 1992). Crayfishes demonstrate a broad spectrum of 65 life history strategies, such as terrestrial burrowing, that allow them to persist in a wide variety of 66 habitats, including lakes, streams, wetlands, caves, and agricultural fields. These unique life 67 histories have allowed species to coexist by occupying distinct ecological niches depending on 68 seasonal water cycles or habitat heterogeneity (Hobbs 1942; Hobbs 1981; Welch and Eversole 69 2006). Nonetheless, crayfishes remain one of the most imperiled taxa in North America. As of 70 the mid-late 1990s, fewer than 50% of crayfish species populations were classified as stable 71 (Taylor et al. 1996; Wilcove et al. 1998). These estimates are likely conservative, as the most 72 recent comprehensive assessment is nearly 20 years old. Further, many species lack adequate 73 information on current distributions, habitat requirements, and threats to conservation that could 74 help prioritize conservation efforts. Substantial attention has been devoted to determining causes 75 of species replacements in crayfish communities related to differential susceptibility to predators 76 (DiDonato and Lodge 1993; Mather and Stein 1993a; Roth and Kitchell 2005), hybridization 77 with native species (Perry et al. 2001a; Perry et al. 2001b), and morphological or behavioral 78 differences among species (Bergman and Moore 2003a; Garvey and Stein 1993; Garvey et al. 79 1994; Gherardi and Daniels 2004; Pintor and Sih 2009), but less attention has been devoted to 80 81 changes in crayfish habitat selection following an invasion. Habitat selection associations of native species often change in response to introductions of non-native species (Kiesecker and 82

Blaustein 1998; Losos et al. 1993; Trammell and Butler 1995), and previous studies document 83

that habitat is an important determinant of predation risk in crayfish (DiDonato and Lodge 1993; 84

Saiki and Tash 1979), and other species in aquatic and terrestrial ecosystems (Creel et al. 2005; 85

Hrabik et al. 2014; Valeix et al. 2009; Werner et al. 1983). Studies of crayfish species 86

distributions teamed with habitat surveys through time could inform our understanding of how 87 invasions affect native species, but many locales lack recent data on crayfish distributions (or in-

88 89 stream habitat) to perform such analyses.

90 A lack of recent crayfish distribution data is a significant concern in many locations. Many states do not have updated crayfish distribution data, despite findings that indicate that 91 invasive cravfishes are one of the most common threats to native cravfishes(Lodge et al. 1998; 92 Lodge et al. 2000). For instance, Wisconsin and Illinois are the only states in the entire Great 93 Lakes Basin that have statewide crayfish surveys in the last 20 years (Olden et al. 2006; Taylor 94 and Redmer 1996). In the state of Michigan, addressing threats to native crayfish posed by 95 invasive species is a management priority. However, the last published comprehensive survey of 96 crayfishes in Michigan was from Creaser (1931), whom provided maps of crayfish collections 97 for individual species. However, these maps lack sufficient resolution to declare which 98 drainages some collections belong. In contrast, Lippson (1975) provides enough detail from their 99 crayfish collections throughout Michigan from the 1960s to provide us with an opportunity to 100 quantify changes in crayfish distributions from a more recent time, albeit from more than 40 101 years ago. Further, research on changes in native species behavior as it pertains to habitat 102 selection is relatively rare, particularly for non-native crayfish impacts on other crayfish species 103 with a few exceptions (Bergman and Moore 2003a; Jansen et al. 2009; Mather and Stein 1993b). 104 Herein, we document changes in the range of one non-native crayfish species (Faxonius rusticus) 105 across the State of Michigan, and quantify how habitat selection of native crayfish species differs 106 in streams where rusty crayfish are present compared to where they are absent. 107

Eight crayfish species are considered native in Michigan. Three are primarily found in 108 permanent open water habitats such as streams and lakes (Cambarus robustus, Faxonius 109 propinguus, and Faxonius virilis). Three use subterranean burrows (Cambarus diogenes, 110 *Cambarus polychromatus*, and *Creaserinus fodiens*) and are rarely observed in open water as 111

adults, except in spring and early summer when they release their young. Two are facultative 112

burrowers, depending on conditions such as precipitation and water levels in adjacent water 113 bodies (Faxonius immunis, and Procambarus acutus) (Hobbs and Jass 1988; Lippson 1975; 114

Thoma et al. 2005). 115

One non-native species of crayfish, the rusty crayfish (F. rusticus), has been reported in 116 Michigan for over 130 years with major range expansion occurring during the 20th Century 117 (Creaser 1931; Faxon 1884; Lippson 1975). The initial F. rusticus range expansion into the Great 118 Lakes basin is attributed to shipping canals connecting the Ohio River and Maumee River 119 watersheds in Ohio, and subsequent spread in the region is believed to be primarily a result of 120 bait bucket release by anglers or intentional release by lake managers seeking to manage 121 macrophyte communities (Creaser 1931; Olden et al. 2006). F. rusticus lives primarily in streams 122 and lakes and is observed to negatively affect populations of native stream and lake dwelling 123 crayfish, including the northern crayfish (F. virilis) and northern clearwater crayfish (F. 124 propinguus) (Garvey et al. 2003). Negative effects of rusty crayfish are numerous: a variety of 125 studies have demonstrated that they outcompete native species for food and shelter, exhibit less 126 susceptibility to native predators, and hybridize with native F. propinguus (Capelli and Munjal 127

1980; Capelli and Munjal 1982; Hill et al. 1993; Perry et al. 2002; Perry et al. 2001b; Roth and 128

129 Kitchell 2005). *F. rusticus* can also affect native fish assemblages through egg predation and by

altering habitat through extensive macrophyte destruction, thereby disrupting native food webs

(Capelli and Munjal 1982; Dorn and Mittelbach 1999; Kreps et al. 2016; Lodge et al. 1998; Marga et al. 2012; Bath et al. 2007)

132 Morse et al. 2013; Roth et al. 2007).

One of the mechanisms by which *F. rusticus* is believed to negatively affect native species is through displacement from preferred habitats. Habitat associations of crayfish communities have received very little attention, except perhaps in northern Wisconsin lakes subject to rusty crayfish invasions (DiDonato and Lodge 1993; Garvey et al. 2003). Crayfish habitat associations in lotic ecosystems, to our knowledge, have yet to be studied although some information does exist at coarse spatial scales (Burskey and Simon 2010).

The quantity of streams and rivers in Michigan provides an opportunity to evaluate 139 crayfish habitat associations both with and without rusty crayfish present. Further, the past 140 distributional data reported by Lippson (1975) offers an opportunity to quantify how crayfish 141 communities in Michigan have changed through time and across space. This study seeks to a) 142 update our current understanding of the status and range of stream-dwelling crayfish within 143 Michigan's Upper and Lower Peninsula, b) identify habitat associations of crayfish species with 144 and without invasive rusty crayfish, c) document changes in the range of crayfish species 145 compared to historical data with an emphasis on non-native rusty crayfish, and d) evaluate the 146 effectiveness of our sampling method. With respect to b), we hypothesize that native species 147 will associate with less-desirable habitat in the presence of rusty crayfish at a given site. For c), 148 we hypothesize that rusty crayfish have expanded their range in Michigan over the last 40 years, 149 but native species have contracted their range owing to negative interactions with rusty crayfish. 150 We also provide an analysis of detectability given the uncertainty regarding our specific capture 151 method (dipnets), and to provide insight into the repeatability of this survey. Information 152 derived from this study will highlight large-scale trends in crayfish communities, and could assist 153 in prioritization of habitats for native crayfish conservation as well as locations to focus 154 prevention efforts for rusty crayfish. This study also adds to the growing body of literature 155 regarding indirect effects of invasions as related to how non-native species affect the habitat 156 selection of native species. 157

158 159 **Methods:**

160 *Crayfish collection*

Although crayfishes occupy two general habitat types – open water habitats and burrows 161 (Hobbs 1989) – this survey was limited to open water habitats of streams. We used the Michigan 162 Department of Natural Resources (MDNR) Stream Status and Trends Program (SSTP) (Seelbach 163 et al. 1997; Wills et al. 2006) to determine stream sampling sites. Stream segments were selected 164 by stratifying the SSTP database by management unit and major watershed. We selected at 165 random 20% of available stream segments for collection from each watershed stratum to evenly 166 distribute sampling effort across watersheds. Stream segments are inter-confluence stream 167 reaches, defined by tributary confluences or dams. The watershed stratum as defined in the 168 SSTP database were individual streams and their tributaries directly connected to a Great Lake 169 (Wills et al. 2006). All data were collected in summer of 2014-2016. We sampled 69 segments 170 in 2014, 277 segments in 2015, and 133 segments in 2016, for a total of 479 segments and 958 171 sites (two sites per segment) over the three year period of the study. 172

Technicians worked in pairs to sample stream segments at each site, and generally
attempted to access streams from a road crossing, with one individual working upstream and the

175 other downstream of the crossing. Technicians worked to catch as many crayfish as possible in a

- 176 20-minute period. We sampled crayfish with dip nets, using standard protocols for crayfish
- 177 collection (Olden et al. 2006). We selected dip nets because it allowed us to implement a
- 178 consistent sampling technique across all streams regardless of substrate type. This included
- 179 netting individuals off substrate, lifting rocks or larger substrate with the foot or hand, and using
- hands and twigs to probe crayfish out of root structures or undercut banks. Collected crayfish
 were temporarily retained for identification and measuring until dip netting was completed at a
- 181 were ten182 site.

Once sampling at a site was complete, we recorded GPS coordinates at the center of each sampling unit. After exiting the stream each crayfish was identified by species. Once crayfish data were recorded, all rusty crayfish were euthanized whereas native crayfish were returned to the stream.

C. diogenes and *C. polychromatus* were combined for all analyses due to their low catch
rates and difficulty in distinguishing young individuals. Because both species were formerly part
of a species complex (Thoma et al. 2005), data for the two were likely combined during
Lippson's survey and will be referred to as the '*diogenes* complex' in this paper (Lippson 1975;

- 191 Thoma et al. 2005).
- 192 193
- 194 *Habitat sampling*

Substrate characteristics were identified using a visual assessment of upstream and downstream sampling areas. Substrate categories were based on a modified Wentworth scale and included clay (<1/256 mm), silt (>1/256 mm, <1/16 mm), sand (>1/16 mm, <4 mm), pebble (>4 mm, <64mm), cobble (>64 mm, <256 mm), boulder (>256 mm), woody material (roots, tree limbs, etc.), detritus, and living macrophytes (Wentworth 1922). Substrate was classified based on amount present in each sampling area using a scale of 0%, 1-24%, 25-49%, 50-74%, and 75-100%.

- 202
- 203 Data and Statistical Analysis
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- 205 <u>Detectability</u>

206 We used occupancy analysis (e.g. Mackenzie et al. 2006) to analyze detect/non-detect data from our survey of sites. These models provide probabilities associated with detecting an 207 individual species in either the same location through time (temporal detectability) or at proximal 208 sites within the same stream segment (spatial detectability). Temporal detectability models were 209 fit to pooled data of both technicians from 22 stream segments that were visited in 2014 and 210 again in 2015. This model tested whether a species would be detected at a location on every 211 occasion that it was sampled. Spatial detectability was modeled by comparing samples from the 212 same stream segment, i.e. one sample from upstream compared to the other from downstream of 213 the road crossing. This model tested whether crayfish assemblages were uniform throughout a 214 stream segment. Samples conducted on the same segment but at different times were treated as 215 their own unique sampling event, allowing a sample size of 479 comparisons for the spatial 216 217 model.

- For this analysis, we made several assumptions consistent with previous applications (e.g. Mackenzie et al. 2006). Foremost, we treated each observer as an independent survey visit, and
- assumed that if the selected crayfish species was present within one observer's section, it was

221 present at the other. Put another way, we assumed that the occupancy status (i.e., present or 222 absent) applied to both sub-sites sampled. This assumption is critical for the occupancy analysis

and implies that if one searcher detects a crayfish species at a site, and the other observer does

not, this is due to a non-detection, thus allowing the analysis to provide estimates of detection

probability. Two other assumptions were that crayfish did not move into or out of a site within the time to survey a site (i.e., the sites are closed during the time of the survey), and that crayfish

227 were identified accurately (i.e., no false positives).

The following occupancy model allowed us to obtain estimates of site occupancy as well as detection probability {MacKenzie, 2002 #2175;Mackenzie, 2006 #2308}:

(1)

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231
$$L(\psi, p) = (\psi^{n} \Pi p_t^{n_t} (1 - p_t)^{n - n_t}) \times (\psi \Pi (1 - p_t) + (1 - \psi))^{N - n}$$

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where *t* is the number of searchers at a site, *N* is the total number of sites surveyed, and *n*. is the number of sites where at least one detection occurred, ψ is the probability of occupancy, *p* is the detection probability for a single searcher, and *n_t* is the number of detections on tth survey. We implemented this occupancy model and obtained estimates via the unmarked package in R (R Core Team 2018).

Estimates of detection probability from this model are for a single searcher; we estimated detectability for two searchers using the following equation:

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$$p_d = 1 - (1 - p_s)^2$$
 (2)

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where, p_d is the probability of detection with two searchers and p_s is the probability of detection for a single searcher determined from the occupancy analysis.

246 <u>Crayfish ranges</u>

247 Crayfish presence/absence data were compared to Lippson's 1975 dissertation (Lippson 1975) to determine any changes in the range of crayfishes in Michigan. Lippson (1975) 248 249 presented their data in terms of successful captures for each species found at their sampling locations. We assumed all species captured at a given site were reported, thus all non-reported 250 species were absent. We also compared changes in the co-occurrence of obligate aquatic species 251 252 (C. robustus, F. propinguus, F. virilis), as a result of increased F. rusticus ranges from previous reports. Lippson's (1975) collections are reported at the county, township, range, and section 253 level. We converted the centroid of these locations to GPS coordinates, and then sorted 254 collections by United States Geological Survey (USGS) 8-digit Hydrologic Unit Code (HUC) 255 watersheds and MDNR Fisheries Management Units (FMU) (Michigan Department of Natural 256 Resources 2001; U.S. Department of Agriculture Service Center Agencies - National Geospatial 257 Management Center 2013). In this way, we were able to assign each of Lippson's sampling 258 locations to an 8-digit HUC watershed, for comparison to our survey data. Range maps were 259 260 constructed using shapefiles published by USDA/NRCS - National Geospatial Management Center and the MDNR in ArcGIS version 10.1 (ESRI 2011; Michigan Department of Natural 261 262 Resources 2001; U.S. Department of Agriculture Service Center Agencies - National Geospatial Management Center 2013). 263

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265 <u>Habitat associations</u>

We used model selection to identify significant habitat predictors of presence or absence 266 267 for all crayfish species. We input habitat predictors into a generalized linear model (GLM) using the logit function, and used backward selection using the stepAIC function of the MASS package 268 269 in R version 3.0.2(R Development Core Team 2008), to identify significant predictors. We repeated this process for each species using the substrate classifications from the modified 270 Wentworth scale as covariates. Species were coded as 1 (present) or 0 (absent). Habitat 271 covariates were also coded as 0-4, corresponding to modified Wentworth scale abundance 272 classifications. We used a logit link function to fit the GLM. This allowed us to determine if the 273 presence of a crayfish species was positively or negatively associated with individual substrate 274 classes. Log odds ratio output from GLMs permitted us to identify the direction and magnitude 275 of effect, as any log odds value below 0 indicated lower than a 50% chance of finding a crayfish 276 associated with a given habitat, and values further away from 0 indicating larger effects. Log 277 odds ratios quantify the ratio of collecting a species to not collecting the species. Thus, if the 278 presence of a given habitat increases the odds of collecting the species, the ratio will increase, 279 and the log odds ratio will be greater than 0. To quantify changes in substrate associations based 280 on the presence or absence of F. rusticus, we ran separate analyses for each native species after 281 dividing samples into those where F. rusticus was present and those where F. rusticus was 282 absent. We used a significance level of α =0.05 for all tests. 283

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286 **Results**:

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288 Detectability

The detectability of crayfish was high for obligate stream species both through time and within individual segments. No obligate stream species had less than a 60% probability of detection and most had detectabilities over 80% (Table 1). The primary and secondary burrowing species showed lower detection probabilities, and temporal models for *C. fodiens* and *P. acutus* could not be run due to lack of data, despite having moderate spatial detectability (46% and 67%, respectively).

295

296 Crayfish ranges

297 During May-September of 2014-2016, crayfish species presence and absence was assessed at 461 of the >2,000 unique stream segments in Michigan (Figure 1). Overall, all of 298 Michigan's native crayfish species were detected in more watersheds during this survey than in 299 1975 (Table 2). For F. rusticus, there is evidence for a substantial range expansion. We found F. 300 rusticus in 20% of samples and in 34 of 55 HUC 8 watersheds. By comparison, F. rusticus were 301 documented in only 12 HUC 8 watersheds in 1975 (Lippson 1975) (Table 2; Figure 2). Every 302 watershed that contained F. rusticus in 1975 also contained F. rusticus in this study (Figure 2). 303 Native species ranges appear to have shifted since 1975. Although we sampled more 304 watersheds than Lippson (1975), shifts in the overall range of some native species is apparent, as 305 well as changes in occurrence within some watersheds (Table 2). The most widely distributed 306 native species in our survey was F. propinguus, which we found in 48 of 55 HUC-8 watersheds 307 and 42% of samples. We found F. propinguus in three more watersheds than Lippson (1975), but 308 in a lower percent of samples (42 versus 72%) (Figure 3, top). When we only consider sites 309 where crayfish were collected, we found F. propinguus in 58% of samples, which still represents 310 a 14% decrease from Lippson (1975). The second most common native species found in this 311

survey was F. virilis, found in 43 watersheds and 20% of all samples, and 27% of samples that 312 313 contained crayfish. This represents a substantial increase in occurrence compared to Lippson (1975), who only found F. virilis in 33 watersheds, but an identical percent of samples (27%) 314 (Figure 3, middle). C. robustus was found in 17 watersheds, and 5.8% of all samples and 8% of 315 samples that contained crayfish, making it the least common obligate aquatic crayfish species in 316 Michigan (Figure 3, bottom). We found C. robustus in five more watersheds compared to 317 Lippson (1975), who found this species in 6.4% of samples. We found F. immunis in 18 318 watersheds and 4.9% of all samples, and 6.8% of samples containing crayfish. Lippson (1975) 319 found F. immunis in eight fewer watersheds, but a similar percent of samples (5.5%) (Figure 4, 320 top). P. acutus was found in 3 watersheds and <1% of samples, making it the least common 321 crayfish in Michigan, limited to a few southern watersheds (Figure 4, bottom). Lippson (1975) 322 did not find P. acutus in any samples. The diogenes complex was found in 23 watersheds and 323 5.4% of all samples, and 7.4% of samples containing crayfish. Lippson (1975) only found C. 324 diogenes in 3.4% of samples, and in 15 fewer watersheds than the present study (Figure 5, top). 325 C. fodiens was found in 9 watersheds and 1.3% of all samples, and in 1.8% of samples 326 containing crayfish. C. fodiens was also rare in Lippson (1975), and was only found in 1.5% of 327 samples, although we found this species in six more watersheds (Figure 5, bottom). 328 Although we found all crayfish species in more watersheds compared to Lippson (1975), 329 we could not detect all species in locations where they were found historically. In terms of 330 facultative burrowing crayfish, we were unable to detect the diogenes complex in one watershed 331 that it was reported in 1975, C. fodiens was not detected in one watershed it was detected in 332 1975, and F. immunis was not detected in four watersheds where it was detected in 1975. For the 333 obligate stream species, we did not detect C. robustus in two watersheds that Lippson (1975) did, 334 F. propinguus was not detected in three watersheds it was formerly detected in 1975, and F. 335 virilis was not detected in six watersheds that it was detected in 1975. 336 The co-occurrence between obligate aquatic species and F. rusticus did not substantially 337 change from Lippson (1975), except for the reduced occurrence of F. propinguus in areas 338 occupied by F. rusticus (Table 3). We found F. propinguus co-occurring with F. rusticus in 23% 339 of samples, which represents a substantial decrease from Lippson (1975) who found F. 340 propinguus in 43% of samples that contained F. rusticus. In contrast F. virilis was found in 16% 341 of samples that contained F. rusticus in 1975, and 15% of samples in 2014-2016. C. robustus 342 was found in 8% of samples that contained F. rusticus in 1975 and 10% of samples in 2014-343 2016. We also found similar co-occurrences of C. robustus with other species across studies. F. 344 propinguus was present in 62% of C. robustus samples in both 1975 and 2014-2016. F. rusticus 345 was in 12% of C. robustus samples in 1975, and 16% of samples in 2014-2016. F. virilis was in 346 12% of C. robustus samples in 1975, and 18% of samples in 2014-2016. At sites where F. 347 propinguus was found, it co-occurred with F. rusticus in 6% of samples in 1975 and 9% in 2014-348 2016. F. virilis occurred in 20% of samples in both 1975 and 2014-2016. C. robustus occurred in 349 6% of samples in 1975, and 9% of samples in 2014-2016. 350

- 351
- 352 Habitat Associations

Model selection results for the substrate covariates and their effect on species presence were successfully calculated for all species except for *P. acutus* due to small sample size (Table 4). The best model for the burrowing species *C. fodiens* only contained detritus, but this species was strongly positively associated with this substrate (Log odds ratio = 0.75, z = 3.83, p < 0.0001; Table 4). The *C. diogenes* complex was also positively associated with detritus (Log odds ratio 358 = 0.46, z = 3.56, p = 0.0004), but the best model for this species also contained boulder although 359 this variable was not significant (Log odds ratio = -1.06, z = -1.263, p > 0.1). *F. immunis*, which 360 is known to burrow but is more often found in slow waters with live vegetation, was found to be 361 positively associated with silt (Log odds ratio = 0.33, z = 3.47, p < 0.001) and live vegetation 362 (Log odds ratio = 0.28, z = 2.29, p = 0.02), which agrees with the life history of this species 363 (Lippson 1975; Tack 1939; Taylor et al. 2015).

Substrate associations of obligate stream dwelling species also agreed with literature 364 descriptions of their life history, for the most part. Model selection for C. robustus indicated 365 positive associations for silt, sand, pebble, cobble, boulder, and wood (all p < 0.02 except for 366 boulder which was not significant)(Table 4) F. propinguus demonstrated strong positive 367 associations with cobble, pebble, and sand (all p <0.001), and F. virilis with live vegetation (Log 368 odds ratio = 0.30, z = 4.332, p < 0.001), detritus (Log odds ratio = 0.23, z = 2.23, p = 0.026), and 369 silt (Log odds ratio = 0.13, z = 2.28, p = 0.023), which agrees with descriptions of their life 370 history (Hobbs and Jass 1988; Lippson 1975; Taylor et al. 2007; Taylor et al. 2015). The best 371 model for invasive F. rusticus contained clay, cobble, and boulder (Table 4), although only the 372 latter two were significant (Cobble: Log odds ratio = 0.38, z = 6.67, p <<0.001; Boulder: Log 373 odds ratio = 0.35, z = 2.988, p = 0.002). 374

Evidence of the impact of rusty crayfish on native species was supported by an analysis 375 that separated samples where F. rusticus co-occurred with native species from those where F. 376 rusticus was absent. The analysis indicated shifts in substrate associations for some species when 377 F. rusticus was present (Table 5). The best model for C. robustus presence in the absence of 378 rusty crayfish contained a variety of substrates (silt, sand, pebble, cobble, boulder, and wood), 379 with the first four of these significant predictors (all p < 0.02). However, in the presence of rusty 380 crayfish, C. robustus was only positively associated with cobble and wood, with wood being the 381 sole significant predictor (Log odds ratio = 1.56, z = 2.86, p = 0.004). The best model for F. 382 immunis in the absence of F. rusticus contained positive associations with clay, silt, sand, 383 detritus, and live vegetation, with silt being the only significant predictor of F. immunis presence 384 at a given site (Log odds ratio = 0.49, z = 3.048, p = 0.002; all others p>0.06)(Table 5). 385 However, when F. rusticus was present, vegetation was the only positive association with F. 386 *immunis*, and this variable was not significant (Log odds ratio = 0.44, z = 1.66, p = 0.10). The 387 best model for F. immunis in the presence of F. rusticus also contained negative associations 388 389 with sand, pebble, and cobble, with sand being the only significant variable in the best model (Log odds ratio = -0.54, z = -2.12, p = 0.03; all others p>0.06). F. propinguus shifted from a 390 best model with significant, positive associations with cobble, pebble, and sand (cobble: Log 391 odds ratio = 0.36, z = 5.8, p<0.001; pebble: Log odds ratio = 0.19, z = 2.86, p = 0.004; sand: Log 392 odds ratio = 0.27, z = 5.26, p<0.001) in the absence of F. rusticus to a best model with only one 393 significant positive association, with live vegetation, when F. rusticus were present (Log odds 394 ratio = 0.48, z = 2.35, p = 0.02). Other, non-significant habitat associations of F. propinguus 395 include positive associations with clay, pebble, and detritus, and negative associations with silt 396 and boulder. All these variables were not significant (all p>0.05), although the negative 397 association with silt was marginally significant (Log odds ratio = -0.4, z = -1.9, p = 0.06) (Table 398 5). F. virilis showed relatively little change in the presence of rusty crayfish. Without F. 399 rusticus, the best model for F. virilis contained a negative association with sand (Log odds ratio 400 = -0.21, z = -3.22, p = 0.001) and a positive association with vegetation (Log odds ratio = 0.21, z 401 = 2.75, p = 0.006). The best model with F. rusticus only contained vegetation, although this 402 predictor was not significant (Log odds ratio = 0.31, z = 1.48, p > 0.1). 403

405 **Discussion:**

The introduction and subsequent spread of non-native species, and their consequent 406 407 effects on native fauna, is a central issue in the conservation of biodiversity. Research to inform the management of aquatic invasions needs to combine mechanistic, typically small-scale, 408 studies of invasive species to understand how the invaders cause negative effects on native 409 species, with broad-scale investigations of invasion spread and ecosystem response. The study 410 reported here focuses on this latter component, combining an extensive survey of contemporary 411 crayfish distributions in Michigan with observations of habitat association shifts in native 412 crayfish species in response to the presence of an invader – the rusty crayfish. Our research 413 complements a similarly broad-scale study in crayfish distribution changes in Wisconsin (Olden 414 et al. 2006) and Illinois (Taylor and Redmer 1996) and adds evidence for effects of rusty crayfish 415 on habitat use by native crayfish. Rusty crayfish have been intensively studied, and their impact 416 on native fauna in individual systems has been well-documented, particularly in northern 417 Wisconsin lakes (e.g. Roth et al. 2007; Wilson et al. 2004). There is some prior evidence of the 418 effect of their geographical spread on the distribution of native crayfishes (Olden et al 2006). Our 419 study adds substantially to this record, and also provides evidence for a potential mechanism for 420 these effects - namely displacement of native crayfishes into less preferred habitats. Broad-scale 421 research of this type provides a valuable foundation for both documenting the extent of an 422 invasion's impact on native ecosystems, and suggesting hypotheses about mechanisms giving 423 rise to the observed patterns that can subsequently become the focus of more mechanistic 424 investigations. 425

426

427 Ranges of Michigan crayfishes

Native crayfish species remain broadly distributed across Michigan, but we found 428 429 evidence of an ongoing expansion of F. rusticus from previous surveys (Creaser 1931; Lippson 1975). The increase in F. rusticus range concomitant with a decrease in co-occurrence with F. 430 propinguus suggests that F. rusticus locally displace F. propinguus, consistent with previous 431 literature. F. rusticus are known to hybridize with F. propinguus, which acts to shift the genetic 432 and phenotypic population toward characteristics exhibited by F. rusticus (Capelli and Munjal 433 1980; Perry et al. 2002; Perry et al. 2001b). Antagonistic interactions between native crayfishes 434 and F. rusticus likely exacerbate rusty crayfish invasions (Mather and Stein 1993a, b), and give 435 rise to habitat use shifts, as discussed below. Previous work has shown that F. rusticus 436 outcompete both F. propinguus and F. virilis for habitat, while exhibiting lower susceptibility to 437 native predators (Bergman and Moore 2003a, b; Capelli and Munjal 1982; DiDonato and Lodge 438 1993; Garvey et al. 2003; Hill and Lodge 1994; Roth and Kitchell 2005). Our findings are 439 consistent with these studies given the shift in F. propinguus associations away from preferred 440 cobble and woody debris toward vegetation. The exclusion of F. propinguus from preferred 441 habitat could make them more susceptible to predation, or place them in suboptimal habitat for 442 growth. However, more research must be conducted to quantify predation on Faxonius 443 crayfishes in stream habitats and the role of predation in species displacement, as previous 444 studies were conducted in lakes (e.g. DiDonato and Lodge 1993; Garvey et al. 2003; Roth and 445 Kitchell 2005). 446 Differences in watershed-level species presence-absence between our survey and 447

448 Lippson's earlier survey may have arisen for multiple reasons. First, our sampling effort, at least

in terms of the number of sites where crayfish were captured, was substantially greater than

Lippson's (694 vs 326). Second, either survey could have failed to detect a species in a

- 451 watershed where they were actually present, particularly if the habitat preferred by a species was
- 452 not observed at the sites where samples were collected, but was in fact present in the watershed.
- Third, in-stream biochemical and habitat conditions have likely changed in some watersheds
- over the past 40 years, so that our sampling detected actual changes in species ranges due to
- habitat change. Finally, as noted above the distributions of some native species may have
 changed as a consequence of the increased range of *F. rusticus*. The relative importance of these
- factors in explaining observed changes in native species distributions will require further study.
- 458

459 *Habitat associations of Michigan crayfishes*

Previous studies indicate that F. virilis adults (in isolation) prefer rocky substrates, and 460 macrophyte beds are important nursery habitat for young (Crocker and Barr 1968; France 1985; 461 Momot and Gowing 1983). Although F. virilis is often considered a habitat generalist, it is 462 vulnerable to exclusion from preferred habitat types as a result of competition, particularly with 463 congeners F. propinguus and F. rusticus (Hobbs and Jass 1988; Lippson 1975; Peck 1985; 464 Taylor et al. 2015). In this study, F. virilis demonstrated an affinity for cobble and a negative 465 association with sand in the absence of both F. propinguus and F. rusticus. However, in areas 466 where F. virilis co-occur with either F. propinguus or F. rusticus we observed that F. virilis was 467 positively associated with vegetation and silt, and was no longer associated with cobble (Table 468 6). Further community change could arise if F. rusticus has a negative effect on macrophyte 469 beds, thus eliminating the remaining refuge for F. virilis and F. propinguus (Lodge and Lorman 470 1987; Roth et al. 2007). Prior to F. rusticus invasion, F. propinguus and F. virilis likely lived in 471 preferred habitat in the absence of the other, with F. virilis persisting in vegetative habitat when 472 the two co-occurred (Garvey et al. 2003; Hill and Lodge 1994; Peck 1985). This still appears to 473 be the case in many locations. However, when F. rusticus excludes F. propinguus from cobble, 474 areas where the three species overlap could result in the eventual removal of F. virilis. 475

C. robustus appeared largely unaffected by F. rusticus despite preferring large coarse 476 substrates. This finding is consistent with Berrill (1978), suggesting that unknown differences in 477 behavior or ecological roles might allow C. robustus to co-occur with F. rusticus and other 478 members of Faxonius spp. This result is echoed by Reid and Nocera (2015), which indicate that 479 C. robustus may occupy a unique niche compared to Faxonius species. However, a report by 480 Daniels (1998) suggests that F. rusticus might be displacing C. robustus in an Ontario watershed. 481 Our data suggests that since 1975 the cohabitation of C. robustus and the other obligate aquatic 482 species, including F. rusticus, is stable. This further suggests that there are determinants that 483 influence the coexistence between C. robustus and Faxonius spp. other than substrate. Berrill et 484 al. (1985) indicate that low pH conditions could be favorable to C. robustus, but the vast 485 majority of rivers in Michigan have pH levels >7.0 (https://waterdata.usgs.gov/mi/), which is 486 above the threshold of 5.5 described in Berrill et al. (1985). 487

We did not observe any negative effects of F. *rusticus* on native burrowing species. Generally, burrowing species tended to prefer silt or detritus substrates, which F. *rusticus* did not prefer in our study. The lack of influence of F. *rusticus* on the presence of these species is likely due in part to their differing life histories. The ability of burrowing species to occupy temporary water bodies and construct semi-terrestrial burrows likely excludes them from much of the shelter competition faced by obligate aquatic species. However, there is a void in literature relating to the relationships between these species and F. *rusticus*.

496 *Detectability*

497 Exclusively sampling streams likely resulted in the under-reporting of Michigan's burrowing crayfish despite their need to enter water during the spring to release young into the 498 499 water (Hobbs and Jass 1988; Lippson 1975). Although we observed burrowing species in more watersheds than Lippson (1975), caution should be used when interpreting this result as a range 500 expansion due to our lack of understanding regarding the specific methods used to capture 501 burrowing crayfish in Lippson (1975). Lippson (1975) indicates that he used a variety of 502 methods including dipnets, seines, burrow excavation, and baited traps, but he was unspecific 503 about which method was used to capture individual specimens or species. The current 504 505 conservation status of burrowing crayfish in Michigan, and in many other locales, is unknown (Taylor et al. 2007). We suggest conducting further surveys aimed at more accurately depicting 506 the range, habitat associations, and status of burrowing species to gain a fuller understanding of 507 burrowing crayfish populations in the state. Surveys could include ephemeral waterbodies, wet 508 meadows, roadside ditches, burrows near streams and ponds, and any other wetlands. Little is 509 known on the status of burrowing species in the state and no extensive work has been done since 510 C. polychromatus was described, separating it as a species apart from C. diogenes (Thoma et al. 511 2005). An evaluation of burrowing crayfish is particularly relevant given the recent detection of 512 red swamp crayfish (P. clarkii) in multiple locations in Michigan (Smith et al in press). All 513 discoveries were in the southern portion of the state in lentic waterbodies (lakes or retention 514 ponds) that were not the focus of the study. However, the discovery of this highly invasive 515 species is concerning not only for the conservation of burrowing crayfish, but for all crayfish 516 species and aquatic food webs as a whole. 517

Overall our methods of dip netting appear to have sufficiently sampled streams for 518 obligate aquatic species of crayfish. Dip netting allowed us to sample all substrate types within 519 flows typical of wading streams. Dip netting also removed the possibility of sample bias related 520 to habitat preferences and sex-specific behavior (Hill and Lodge 1994; Olden et al. 2006; Price 521 and Welch 2009; Smily and Dibble 2000) Passive methods of capture, such as trapping, results 522 in a bias toward males of more aggressive species and might result in different catch rates in 523 different waterbodies based on predator densities (Collins et al. 1983; Dorn et al. 2005). Other 524 studies have stated detection probabilities upwards to 88% for throw traps (Dorn et al. 2005), 525 68% for electroshocking, 38% for trapping, and dip netting as low as 32% (for one half hour) 526 (Price and Welch 2009). Our detectability model showed that dip netting appeared to be an 527 effective method of detecting crayfish in a stream. For obligate aquatic species, spatial or 528 temporal detectability was never below 60%, which is substantially higher than the other studies. 529

530

531 Conclusion

This study presents evidence that rusty crayfish have continued to expand their 532 distribution in the state Michigan since the last comprehensive survey more than 40 years ago, 533 based on a systematic and representative survey of catchments throughout the state. We also 534 found that habitat selection by the two most widespread native species, F. propinguus and F. 535 *virilis*, shifts when the invasive *F. rusticus* is present, suggesting a mechanism for biogeographic 536 effects of this invader on native crayfishes. Our study provides a model for broad-scale 537 investigations of the spread and effects of an aquatic invasive species, and helps to guide more 538 intensive, mechanistic investigations into the causes of invasive species impacts on native 539 540 species, ideally leading to advice on strategies for mitigating the negative effects of invaders. 541

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553	References
554	
555	Bergman D, Moore P (2003a) Field observations of intraspecific agonistic behavior of two
556	crayfish species, Orconectes rusticus and Orconectes virilis, in different habitats. The
557	Biological Bulletin 205:26-35
558	Bergman D, Moore P (2003b) Field observations of intraspecific agonistic behavior of two
559	crayfish species, Orconectes rusticus and Orconectes virilis, in different habitats.
560	Biological Bulletin 205:26-35
561	Berrill M (1978) Distribution and ecology of crayfish in the Kawartha Lakes region of southern
562	Ontario. Can.J.Zool. 56:166-177
563	Berrill M, Hollett L, Margosian A, et al. (1985) Variation in tolerance to low environmental pH
564	by the crayfish Orconectes rusticus, O. propinguus, and Cambarus robustus Canadian
565	Journal of Zoology 63:2586-2589
566	Burskey JL, Simon TP (2010) Reach- and Watershed-scale Associations of Crayfish within an
567	Area of Varying Agricultural Impact in West-central Indiana. Southeast Nat 9:199-216
568	Capelli G, Munjal B (1980) Hybridization between crayfish of the genus <i>Orconectes</i> :
569	morphological evidence. Crustaceana 39:121-132
570	Capelli GM, Munjal BL (1982) Aggressive interactions and resource competition in relation to
571	species displacement among crayfish of the genus <i>Orconectes</i> . Journal of Crustacean
572	Biology 2:486-492
573	Carreira B, Dias M, Rebelo R (2014) How consumption and fragmentation of macrophytes by
574	the invasive crayfish Procambarus clarkii shape the macrophyte communities of
575	temporary ponds. Hydrobiologia 721:89-98
576	Charlebois P, Lamberti G (1996) Invading crayfish in a Michigan stream: direct and indirect
577	effects on periphyton and macroinvertebrates. J N Am Benthol Soc 15:551-563
578	Collins NC, H. H. Harvey, A. J. Tierney, et al. (1983) Influence of predatory fish density on
579	trapability of crayfish in Ontario lakes. Can. J. Fish. Aquat. Sci. 40:1820-1828
580	Creaser EP (1931) The Michigan decapod crustaceans. Papers of the Michigan Academy of
581	Science, Arts and Letters 13:257-276
582	Creel S, Winnie Jr. J, Maxwell B, et al. (2005) Elk alter habitat selection as an antipredator
583	response to wolves. Ecology 86:3387-3397
584	Crocker DW, Barr DW (1968) Handbook of the Crayfishes of Ontario. University of Toronto
585	Press, Toronto, Ontario
586	Daniels R (1998) Changes in the distribution of stream dwelling crayfishes in the Schoharie
587	Creek system, eastern New York State. Northeastern Naturalist 5:231-248
507	Creek System, custem from fork State. Horneustem Futuralist 5.251 210

588	Didham R, Tylianakis J, Gemmell N, et al. (2007) Interactive effects of habitat modification and
589	species invasion on native species decline. Trends in Ecology and Evolution 22:489-496
590	Didham R, Tylianakis J, Hutchison M, et al. (2005) Are invasive species the drivers of
591	ecological change? Trends in Ecology and Evolution 20:470-475
592	DiDonato G, Lodge D (1993) Species replacements among Orconectes crayfishes in Wisconsin
593	lakes: the role of predation by fish. Can. J. Fish. Aquat. Sci. 50:1484-1488
594	Dorn N, Urguelles R, Trexler J (2005) Evaluating active and passive sampling methods to
595	quantify crayfish density in a freshwater wetland. J N Am Benthol Soc 24:346-356
596	Dorn NJ, Mittelbach GG (1999) More than predator and prey: a review of interactions between
597	fish and crayfish. Vie Et Milieu 49:229-237
598	ESRI (2011) ArcGIS Desktop: Release 10. Redlands, CA: Environmental Systems Research
599	Institute.
600	Faller M, Harvey GL, Henshaw AJ, et al. (2016) River bank burrowing by invasive crayfish:
601	Spatial distribution, biophysical controls and biogeomorphic significance. Sci Total
602	Environ 569:1190-1200
603	Faxon W (1884) Descriptions of New Species of Cambarus Proceedings of the American
604	Academy of Arts and Science 20:107-158
605	France RL (1985) Relationship of crayfish (Orconectes virilis) growth to population abundance
606	and system productivity in small oligotrophic lakes in the Experimental Lakes Area,
607	Northwestern Ontario. Can. J. Fish. Aquat. Sci. 42:1096-1102
608	Garvey JE, Rettig JE, Stein RA, et al. (2003) Scale-dependent associations among fish predation,
609	littoral habitat, and distributions of crayfish species. Ecology 84:3339-3348
610	Garvey JE, Stein RA (1993) Evaluating how chela size influences the invasion potential of an
611	introduced crayfish (Orconectes rusticus). Am.Midl.Nat. 129:172-181
612	Garvey JE, Stein RA, Thomas HM (1994) Assessing how fish predation and interspecific prey
613	competition influence a crayfish assemblage. Ecology 75:532-547
614	Gherardi F (2006) Crayfish invading Europe: the case study of Procambarus clarkii. Mar Freshw
615	Behav Phy 39:175-191
616	Gherardi F, Daniels W (2004) Agonism and shelter competition between invasive and
617	indigenous crayfish species. Canadian Journal of Zoology 82:1923-1932
618	Hill A, Lodge D (1994) Diel changes in resource demand: Competition and predation in species
619	replacement among crayfishes. Ecology 75:2118-2126
620	Hill A, Sinars D, Lodge D (1993) Invasion of an occupied niche by the crayfish Orconectes
621	rusticus: potential importance of growth and mortality. Oecologia 94:303-306
622	Hobbs H (1942) The crayfishes of Florida. University of Florida Publications. Biological Science
623	Series 3:1-79
624	Hobbs HH (1981) The crayfishes of Georgia. Smithsonian Contribution to Zoology 318:1-549
625	Hobbs HHI, Jass JP, Huner JV (1989) A review of global crayfish introductions with particular
626	emphasis on two North American species (Decapoda, Cambaridae). Crustaceana 56:299-
627	316
628	Hobbs HI, Jass J (1988) The crayfishes and shrimps of Wisconsin. Wisconsin Public Museum,
629	Milwaukee
630	Hrabik T, Roth B, Ahrenstorff TD (2014) Predation risk and prey fish vertical migration in Lake
631	Superior: Insights from an individual based model of siscowet (Salvelinus namaycush).
632	Journal of Great Lakes Research 40:730-738

633	Huner J, Lundquist O (1995) Phyisological adaptations of freshwater crayfishes that permit
634	successful aquacultural enterprises. American Society of Zoology 35:12-19
635	Ilheu M, Bernardo J, Fernandes S (2007) Predation of invasive crayfish on aquatic vertebrates:
636	the effect of Procambarus clarkii on fish assemblages in Mediterranean temporary
637	streams Biological Invaders of Inland Waters 29:543-558
638	Jansen W, Geard N, Mosindy T, et al. (2009) Relative abundance and habitat association of three
639	crayfish (Orconectes virilis, O. rusticus, and O. immunis) near an invasion front of O.
640	rusticus, and long-term changes in their distribution in Lake of the Woods, Canada.
641	Aquat Invasions 4:627-649
642	Kershner MW, Lodge DM (1995) Effects of littoral habitat and fish predation on the distribution
643	of an exotic crayfish, Orconectes rusticus. J N Am Benthol Soc 14:141-422
644	Kiesecker J, Blaustein A (1998) Effects of introduced bullfrogs and smallmouth basson
645	microhabitat use, growth, and survival of native red-legged frogs (Rana aurora). Conserv
646	Biol 12:776-787
647	Kreps T, Larson E, Lodge D (2016) Do invasive rusty crayfish (Orconectes rusticus) decouple
648	littoral and pelagic energy flows in lake food webs? Freshwater Science 35:104-113
649	Lippson R (1975) The Distribution of the Crayfishes of Michigan with Aspects of Their Life
650	Cycle and Physiology. Michigan State University, pp. 201pp
651	Lodge DM, Kershner MW, Aloi JE, et al. (1994) Effects of an omnivorous crayfish (Orconectes
652	<i>rusticus</i>) on a freshwater littoral food web. Ecology 75:1265-1281
653	Lodge DM, Lorman JG (1987) Reductions in submersed macrophyte biomass and species
654	richness by the crayfish Orconectes rusticus. Can. J. Fish. Aquat. Sci. 44:591-597
655	Lodge DM, Stein RA, Brown KM, et al. (1998) Predicting impact of freshwater exotic species
656	on native biodiversity: challenges in spatial scaling. Australian Journal of Ecology 23:53-
657	67
658	Lodge DM, Taylor CA, Holdich DM, et al. (2000) Nonidigenous crayfishes threaten North
659	American freshwater biodiversity. Fisheries 25:7-19
660	Losos J, Marks J, Schoener T (1993) Habitat use and ecological interactions of an introduced and
661	a native species of Anolis lizard on Grand Cayman, with a review of the outcomes of
662	anole introductions. Oecologia 95:525-532
663	Lozon J, MacIsaac HJ (1997) Biological invasions: are they dependent on disturbance?
664	Environmental Review 5:131-144
665	Mackenzie D, Nichols J, Royle A, et al. (2006) Occupancy estimation and modelling. Academic
666	Press, Burlington, MA
667	Mather ME, Stein RA (1993a) Direct and indirect effects of fish predation on the replacement of
668	a native crayfish by an invading congener. Can. J. Fish. Aquat. Sci. 50:1279-1288
669	Mather ME, Stein RA (1993b) Using growth/mortality trade-offs to explore a crayfish species
670	replacement in stream riffles and pools. Can. J. Fish. Aquat. Sci. 50:88-96
671	Michigan Department of Natural Resources (2001) Fisheries Management Units. Michigan
672	Department of Natural Resources, Fisheries Research Reports, Lansing, MI.,
673	Momot WT (1995) Redefining the role of crayfish in aquatic ecosystems. Rev. Fish. Sci. 3:33-63
674	Momot WT, Gowing H (1983) Some factors regulating cohort production of the crayfish
675	Orconectes virilis. Freshwater Biol. 13:1-12
676	Momot WT, Gowing H, Jones PD (1978) The dynamics of crayfish and their role in ecosystems.
677	Am.Midl.Nat. 99:10-35

- Morse J, Baldridge A, Sargent L (2013) Invasive crayfish *Orconectes rusticus* is a more effective
 predator of substrate nesting fish eggs than native crayfish (*O. virilis*). Crustaceana
 86:387-402
- Moyle PB, Light T (1996) Biological invasions of fresh water: empirical rules and assembly
 theory. Biological Conservation 78:149-161
- Nystrom P, Stenroth P, Holmqvist N, et al. (2006) Crayfish in lakes and streams: individual and
 population responses to predation, productivity and substratum availability. Freshwater
 Biology 51:2096-2112
- Olden J, McCarthy J, Maxted J, et al. (2006) The rapid spread of rusty crayfish (*Orconectes rusticus*) with observations on native crayfish declines in Wisconsin (U.S.A.) over the past 130 years. Biological Invasions 8:1621-1628
- Peck SK (1985) Effects of aggressive interaction on temperature selection by the crayfish,
 Orconectes virilis. The American Midland Naturalist 114:159-167
- Perry W, Feder J, Lodge D (2002) Importance of Hybridization between indigenous and
 nonindigenous freshwater species: an overlooked threat to North American biodiversity.
 Systematic Biology 51:255-275
- Perry WL, Feder JL, Dwyer G, et al. (2001a) Hybrid zone dynamics and species replacement
 between *Orconectes* crayfishes in a northern Wisconsin lake. Evolution 55:1153-1166
- Perry WL, Feder JL, Lodge DM (2001b) Implications of hybridization between introduced and
 resident *Orconectes* crayfishes. Conserv Biol 15:1656-1666
- Pintor L, Sih A (2009) Differences in growth and foraging behavior of native and introduced
 populations of an invasive crayfish. Biological Invasions 11:1895-1902
- Price J, Welch S (2009) Semi-quantitative Methods for Crayfish Sampling: Sex, Size, and
 Habitat Bias. Journal of Crustacean Biology 29:208-216
- R Development Core Team (2008) A language and environment for statistical computing. R
 Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0, URL
 http://www.R-project.org.
- Rabeni CF (1992) Trophic linkage between stream centrarchids and their crayfish prey. Can. J.
 Fish. Aquat. Sci. 49:1714-1721
- Reid S, Nocera J (2015) Composition of native crayfish assemblages in southern Ontario rivers
 affected by rusty crayfish (*Orconectes rusticus* Girard, 1852) invasions implications for
 endangered queensnake recovery. Aquat Invasions 10:189-198
- Roth BM, Hein CL, Zanden MJV (2006) Using bioenergetics and stable isotopes to assess the
 trophic role of rusty crayfish (*Orconectes rusticus*) in lake littoral zones. Canadian
 Journal of Fisheries and Aquatic Sciences 62:335-344
- Roth BM, Kitchell JF (2005) The role of size-selective predation in the displacement of
 Orconectes crayfishes following rusty crayfish invasion. Crustaceana 78:299-312
- Roth BM, Tetzlaff JC, Alexander ML, et al. (2007) Reciprocal relationships between exotic rusty
 crayfish, macrophytes, and Lepomis species in northern wisconsin lakes. Ecosystems
 10:74-85
- Saiki M, Tash J (1979) Use of cover and dispersal by crayfish to reduce predation by largemouth
 bass. In: Johnson D and Stein R (eds) Response of fish to habitat structure in standing
 water. American Fisheries Society, pp. 44-48
- Seelbach P, Whiley M, Kotanchik J, et al. (1997) A Landscape-Based Ecological Classification
 System for River Valley Segments in Lower Michigan (MI-VSEC Version 1.0).
 Michigan Department of Natural Resources Eisberies Report No. 2036
- 723 Michigan Department of Natural Resources Fisheries Report No. 2036.

- Smily PC, Dibble ED (2000) Microhabitat use of an introduced crayfish (*Orconectes rusticus*) in
 Long Lake, Wisconsin. J.Freshw.Ecol. 15:115-123
- Tack P (1939) The life history and ecology of the crayfish *Cambarus immunis* (Hagen).
 Michigan State University Press, East Lansing, MI.
- Taylor C, Schuster G, Cooper J, et al. (2007) A reassessment of the conservation status of
 crayfishes of the United States and Canada after 10+ years of increased awareness.
 Fisheries 32:372-389
- Taylor CA, Redmer M (1996) Dispersal of the crayfish *Orconectes rusticus* in Illinois, with
 notes on species displacement and habitat preference. Journal of Crustacean Biology
 16:547-551
- Taylor CA, Schuster G, Wylie D (2015) Field Guide to Crayfishes of the Midwest. Illinois
 Natural History Survey Press
- Taylor CA, Warren ML, Fitzpatrick JF, et al. (1996) Conservation status of crayfishes of the
 United States and Canada. Fisheries 21:25-38
- Thoma RF, Jezerinac F, Simon TP (2005) *Cambarus (Tubericambarus) polychromatus* (Decapoda: Cambaridae), a new species of crayfish from the United States. Proceedings
 of the Biological Society of Washington 118:326-336
- Trammell M, Butler J (1995) Effects of exotic plants on native ungulate use of habitat. The
 Journal of Wildlife Management 59:808-816
- U.S. Department of Agriculture Service Center Agencies National Geospatial Management
 Center (2013) 8 Digit Watershed Boundary Dataset. USDA/NRCS National Geospatial
 Management Center, Reston, VA.
- Valeix M, Loveridge A, Chamaille-Jammes S, et al. (2009) Behavioral adjustments of African
 herbivores to predation risk by lions: spatiotemporal variations influence habitat use.
 Ecology 90:23-30
- Welch S, Eversole A (2006) The occurrence of primary burrowing crayfish in terrestrial habitat.
 Biological Conservation 130:458-464
- Werner EE, Gilliam JF, Hall DJ, et al. (1983) An experimental test of the effects of predation
 risk on habitat use in fish. Ecology 64:1540-1548
- Wilcove DS, Rothstein D, Dubow J, et al. (1998) Quantifying threats to imperiled species in the
 United States. Bioscience 48:607-615
- Wills T, Zorn T, Nuhfer A (2006) Stream Status and Trends Program sampling protocols. In:
 Schneider J (ed) Manual of fisheries survey methods II: with periodic updates. Michigan
 Department of Natural Resources Fisheries Special Report 25, Ch. 26, Ann Arbor,
 Michigan,
- Wilson KA, Magnuson JJ, Lodge DM, et al. (2004) A long-term rusty crayfish (*Orconectes rusticus*) invasion: dispersal patterns and community change in a north temperate lake.
 Can. J. Fish. Aquat. Sci. 61:2255-2266
- 762



Figure 1. Stream segments sampled from 2014-2016. Each dot represents one paired sample.



Figure 2. Distribution of rusty crayfish in 1975 (L) and 2014-2016 (R). Closed circles indicate where $\underline{F}\Theta$. *rusticus* was found, open circles denote locations where $\underline{F}\Theta$. *rusticus* was not detected.



Figure 3. Distribution maps of obligate aquatic species, <u> $F\Theta$ </u>. propinguus (top), <u> $F\Theta$ </u>. virilis

(middle), and *C. robustus* (bottom) from Lippson (1975) (left) and 2014-2016 (right). Closed

- circles indicate where specimens were found, open circles denote locations where specimens
- 777 were not detected.





783	Figure 4. Distribution map of facultative burrowing species <u>FO</u> . immunis (top) and P. acutus
784	(bottom). Data for <u>OF</u> . immunis is present from Lippson (1975) (left) and 2014-2016 (right),
785	whereas no P. acutus were found in Lippson (1975). Closed circles indicate where specimens
786	were found, open circles denote locations where specimens were not detected.



791 Figure 5. Distribution maps of obligate burrowing species, *C. diogenes* (top) and <u>*CF. fodiens*</u>

792 (bottom) from Lippson (1975) (left) and 2014-2016 (right). Closed circles indicate where

specimens were found, open circles denote locations where specimens were not detected.

ψ being occupant			-	
	Spatial Dete	ectability	Temporal D	etectability
Species	Ψ	Р	Ψ	Р
C. diogenes	0.289	0.222	0.182	0.500
C. robustus	0.132	0.607	0.142	0.800
C. fodiens	0.040	0.462	-	-
F. immunis	0.121	0.533	0.182	0.500
F. propinquus	0.548	0.826	0.683	0.966
F. rusticus	0.283	0.852	0.230	0.889
F. virilis	0.354	0.609	0.371	0.857
P. acutus	0.013	0.667	-	-
	n=350		n=22	

Table 1: Detectability of crayfish species during 2014-2015 stream surveys over time and space.

801 Table 2: Crayfish occurrence by HUC8 watershed (n=55). A comparison of occurrences reported

in a 1975 survey and findings during 2014-2016 field sampling. Specific watersheds

Species	1975	2016
C. diogenes	8	23
C. robustus	12	17
C. fodiens	3	9
F. immunis	10	18
F. propinquus	45	48
F. rusticus	12	34
F. virilis	33	43
P. acutus	0	3

803 occurrences are located in the Appendix.

804

805

- Table 3: Relative co-occurrence (%) of Michigan obligate stream-dwelling crayfish in samples of
- 808 C. robustus (n=56 for this study), <u>FO</u>. propinguus (n=405 for this study), and <u>OF</u>. rusticus
- 809 (n=198 for this study) for years 1975 and 2016 and the amount of change between years. It
- should be noted that the number of samples for 1975 in unknown.
- 811

			Survey	Species		
	C. rob	oustus	F. prop	inquus	F. rus	sticus
Co-occurring Species	1975	2016	1975	2016	1975	2016
C. robustus			6	9	8	10
F. propinquus	62	62			43	23
F. rusticus	12	16	6	9		
F. virilis	12	18	20	20	16	15

813

815 Table 4: Summarized generalized linear model results showing statistically significant

816 relationships for crayfish species presence or absence based on habitat. Potential covariates were

clay, silt, sand, pebble, cobble, boulder, wood, detritus, and live vegetation. Asterisks indicate

s significal	lice at $p < 0.05()$, < 0	.01(), <0.001()	-
	log Odds	log Odds	log Odds	log Odds
	(Std. Error)	(Std. Error)	(Std. Error)	(Std. Error)
	C. diogenes	C. robustus	C. fodiens	F. immunis
(Intercept)	-2.99 (0.17)***	-5.60 (0.77)***	-4.78 (0.36)***	-3.58 (0.25)***
Clay				
Silt		0.59 (0.20)**		0.33 (0.10)***
Sand		0.42 (0.17)*		
Pebble		0.47(0.18)**		
Cobble		0.83 (0.17)***		
Boulder	-1.06 (0.84)	0.48(0.27)		
Wood		0.94 (0.29)**		
Detritus	0.46 (0.13)***		0.75 (0.20)***	
Live Veg.				0.28 (0.12)*

818	significance at p<0.05(*), <0.01(**), <0.001(***)	

	F. propinquus	F. rusticus	F. virilis
(Intercept)	-1.00 (0.13)***	-1.90 (0.12)***	-1.80 (0.12)***
Clay		-0.56 (0.47)	
Silt			0.13 (0.06)*
Sand	0.26 (0.05)***		
Pebble	0.21 (0.16)***		
Cobble	0.18 (0.05)***	0.38 (0.06)***	
Boulder		0.35 (0.11)**	
Wood			
Detritus			0.23 (0.10)*
Live Veg.			0.30 (0.07)***

819

820

822	Table 5. Comparison of generalized linear model results where habitat predictors of crayfish species presence or absence is influence	d
823	by the presence or absence of <u>OF</u> . rusticus.	

	F. rusticus present	F. rusticus absent		F. rusticus present	F. rusticus absent
C. robustus			F. propinqu	ius	
	log Odds (Std. Error)	log Odds (Std. Error)		log Odds (Std. Error)	log Odds (Std. Error)
(Intercept)	-4.33(0.80)***	-5.33(0.80)***	(Intercept)	-1.78(0.33)***	-0.90(0.14)***
Clay			Clay	8.17(441.3)	
Silt		0.56(0.21)**	Silt	-0.40(0.21)	
Sand		0.39(0.18)*	Sand		0.27(0.05)***
Pebble		0.42(0.19)*	Pebble	0.27(0.16)	0.19(0.07)**
Cobble	0.45(0.27)	0.83(0.17)***	Cobble		0.36(0.06)***
Boulder		0.51(0.30)	Boulder	-0.55(0.44)	
Wood	1.56(0.54)**	0.62(0.37)	Wood		
Detritus			Detritus	0.52(0.29)	
Live Veg.			Live Veg.	0.48(0.20)*	
		'	•		•
F. immunis			F. virilis		
(Intercept)	-1.46(0.61)*	-4.52(0.61)***	(Intercept)	-1.96(0.25)***	-1.21(0.14)***
Clay		0.61(0.33)	Clay		
Silt		0.49(0.16)**	Silt		
Sand	-0.54(0.26)*	0.25(0.18)	Sand		-0.21(2.75)**
Pebble	-1.00(0.53)		Pebble		
Cobble	-0.33(0.21)		Cobble		
Boulder			Boulder		
Wood			Wood		
Detritus		0.40(0.22)	Detritus		
Live Veg.	0.44(0.27)	0.30(0.18)	Live Veg.	0.31(0.21)	0.21(0.08)**

- Table 6: GLM output for substrate co-variate effect on ΘF . virilis presence when ΘF .
- 826 propinguus and $\Theta \underline{F}$. rusticus were absent compared to when either $\Theta \underline{F}$. propinguus or $\underline{F}\Theta$.
- *rusticus* were present in samples.

	FR and FP	FR and FP
	absent	present
(Intercept)	-1.46 (0.30)***	-1.89 (0.15)***
Clay		
Silt		0.15 (0.09)*
Sand	-0.29 (0.11)**	
Pebble		
Cobble	0.31 (0.12)**	
Boulder	-0.73 (0.45)	
Wood	0.58 (0.31)	
Detritus	0.22 (0.13)	
Live Veg.	0.22 (0.11)	0.34 (0.10)***

					C. robustus		iens	F. imn		F. propinquus		F. rust		F. virilis		P. acutus	
HUC8	Watershed	1975	2016	1975	2016		2016	1975	2016	1975		1975	2016	1975	2016	1975	2016
	Sum	8	23	12	17	3	9	10	18	45	48	12	34	33	43	0	3
4080101	Au Gres-Rifle		X	X	x		х		х	х	х		x				
4070007	Au Sable			X	x					х	х	X	x	х	x		
4010302	Bad-Montreal										х						
4060104	Betsie-Platte		X							X	х			x	X		
4020201	Betsy-Chocolay		X							X	х				X		
4080104	Birch-Willow									x	х			x	X		
4070005	Black			X						X	X		x		X		
4050002	Black-Macatawa	X	X						Х	X	х		x		X		Х
4020101	Black-Presque Isle		X								х				X		
4060105	Boardman-Charlevoix									X	х	X	x	x	X		
4060107	Brevoort-Millecoquins	X	X							х	х			х	X		
4030106	Brule								Х	х	х	X	X	х			
4070002	Carp-Pine		x							х	х				x		
4080205	Cass		x					х	х	х			x	x			
4030109	Cedar-Ford									х	х		x		x		
4070004	Cheboygan									х	х	x	x	x	x		
4090003	Clinton	x		x	x				х	х	х		x		x		
4020105	Dead-Kelsey										х		x	x	x		
4090004	Detroit				x		х	х		х	х		x		x		
4030110	Escanaba										х				x		
4030112	Fishdam-Sturgeon		x							х	х			x	x		
4080204	Flint			x	x					х	х		x	x	x		
4090005	Huron		x		x					х	х		x		x		
4050003	Kalamazoo	x	x			x	х	х	х	х	х	x	x	x	x		
4080102	Kawkawlin-Pine		x		x				х		х			x			
4020103	Keweenaw Peninsula																
4090002	Lake St. Clair								х		x			x			
4040001	Little Calumet-Galien								х	x	x		x		x		
4070003	Lone Lake-Ocqueoc			x						x	x		x		x		
4050006	Lower Grand		x	x	x				х	x	x			x	x		
4060103	Manistee		x		x				х	x	x	x	x	x	x		
4060106	Manistique	1	1							x	x	1		İ	x		

Table A1. Crayfish occurrence by HUC8 watershed (n=55). A comparison of occurrences reported in a 1975 survey and findings
 during 2014-2016 field sampling.

Table A1 cont.

HUC8	Watershed	L. dio	genes	C. robustus		F. fodiens		F. immunis		F. propinquus		F. rusticus		F. virilis		P. acutus	
4050005	Maple		x		X				X	X	X			X	x		
4030108	Menominee								X	X			Х	X	x		
4030107	Michigamme									х	X			х			
4060102	Muskegon		x				X		х	x	X	x	X	х	x		
4020102	Ontonagon									х	х		х	х	x		
4100001	Ottawa-Stony							Х	х	х			х		x		X
4060101	Pere Marquette-White		x							x	X				x		
4080103	Pigeon-Wiscoggin									x	X		X	X	x		
4080202	Pine									x	X		X		x		
4100002	Raisin	x	x	x	x		X	X	X	x	X			X	x		
4080203	Shiawassee	x	x			x		X	X	x	X		X	X	x		
4090001	St. Clair				x			X	X	x	X		X	X	x		
4050001	St. Joseph 1	x	x		x		X	X		x	X	x	X	X	x		X
4100003	St. Joseph		x	x	x							x	X				
4070001	St. Marys									x	X		Х	X	x		
4020104	Sturgeon													X	x		
4030111	Tacoosh-Whitefish									х	Х		Х	X			
4020202	Tahquamenon									x	X						
4050007	Thornapple		X				x	X		x	X			X	x		
4070006	Thunder Bay			x	x		x			x	X	x	х	X	x		
4100006	Tiffin			x	x							x	х				
4080201	Tittabawassee			x	x					х	X	x	х	X	x		
4050004	Upper Grand	х	х			х	х	Х		х	Х		х	х	х		