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2	Evaluating active genetic options for the control of Sea Lampreys (Petromyzon
3	marinus) in the Laurentian Great Lakes
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6	Ronald E. Thresher <sup>1</sup> , Michael Jones <sup>2</sup> , D. Andrew R. Drake <sup>3</sup>
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12	1. SF Tech, 50 Bramble Street, Ridgeway, Tasmania 7054 Australia
13	2. Fisheries Science Center, Michigan State University, East Lansing, Michigan,
14	USA
15	3. Great Lakes Laboratory for Fisheries and Aquatic Sciences, Fisheries and
16	Oceans Canada, 867 Lakeshore Road, Burlington, Ontario L7S 1A1 Canada
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19	Corresponding author: Ronald Thresher, 50 Bramble Street, Ridgeway, Tasmania
20	7054, Australia; Telephone: 61-3-62391496; e-mail: Ronaldethresher@gmail.com
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29 For more than two decades the Great Lakes Fishery Commission has sought 30 tactics to complement, and potentially replace, the use of barriers and lampricides to 31 control Sea Lampreys in the Great Lakes, but thus far without success. This paper 32 examines the potential of modern genetic technology to suppress these invasive 33 populations. We identified six recombinant options that appeared to be moderately to 34 highly feasible, most of which were judged by an expert panel as extremely low or 35 low risk, and for which R&D was broadly supported by stakeholders. The two 36 options judged to overall best combine high efficacy and low risks were a Mendelian 37 "sex ratio drive" and genetically modifying a prey species as to kill or sterilize Sea 38 Lampreys that fed on it. Core issues regarding use of genetic biocontrol in the Great 39 Lakes include technical problems associated with maintaining a Sea Lamprey brood 40 line, information gaps for most options, the extent of broader public support, and the 41 extent and nature of national and international consultation required in making 42 decisions about control options.

## 43 Introduction

45	The Sea Lamprey (Petromyzon marinus) is one of the world's most
46	destructive environmental species. After invading the upper Laurentian Great Lakes
47	(GL) in the 1930s, the Sea Lamprey destroyed commercial fisheries worth millions of
48	dollars and fundamentally altered the lake ecosystems. Currently, the species is
49	controlled by a joint U.S. and Canadian program, managed by the Great Lakes
50	Fishery Commission (GLFC), that is based principally on trapping, barriers to prevent
51	access to spawning sites, and biocidal treatment of spawning and nursery tributaries.
52	The annual cost exceeds US \$20 million. Alternative approaches, such as
53	chemosterilization/sterile male release programs (Twohey et al., 2003; Bergstedt and
54	Twohey 2007) and pheromone-based attractants and repellents (Li et al., 2003;
55	Johnson et al., 2009b), have yet to demonstrate efficacy as management tools (e.g.,
56	Johnson et al., 2013, Dawson et al., 2016). Over the next several years, the costs of
57	control could increase markedly due to stakeholder pressure to remove barriers that
58	currently prevent Sea Lamprey access to large areas of productive habitat (Lavis et
59	al., 2003) and possibly the development of resistance to lampricides (Dunlop et al.,
60	2017). Further, significant Sea Lamprey production arises from connecting
61	channels, such as the St. Clair River between Lake Huron and Lake St. Clair, that are
62	poorly suited for cost-effective management using existing control tactics. These
63	issues reinforce the need for alternative long-term, cost-effective options for
64	managing Sea Lampreys in the Great Lakes (Dunlop et al., 2017).
65	Modern genetic technology could help satisfy this need. The concept of
66	actively using genetic techniques to suppress invasive pest populations is not new. As
67	early as the 1960s, entomologists speculated that genetics could be used to manage

68	insect pests, based on the observation that meiotic drive could distort sex-ratios in
69	insect populations to the point of extinction (Hamilton, 1967). However, the idea
70	languished in the absence of practical genetic tools to effectively manipulate inherited
71	sex ratios. Since then, modern recombinant genetics has provided a range of options
72	for manipulating phenotype. A recombinant lethal construct - the genetic equivalent
73	of the traditional Sterile Insect Technique (SIT) widely used to control insect pests
74	(Krafsur, 1998) - has been extensively tested in the laboratory and in cage trials
75	(Thomas et al., 2000; Phuc et al., 2007; Klein et al., 2012), and has been used
76	successfully to suppress mosquito populations in Brazil, Malaysia and the Cayman
77	Islands (Harris et al. 2012; Lacroix et al., 2012). Several variants of a female-specific
78	lethal gene, a form of inherited sex ratio distortion, have also been demonstrated in
79	insects (Heinrich and Scott, 2000; Thomas et al. 2000; Fu et al. 2007, 2010;
80	Windbichler et al., 2007; Ant et al. 2012; Galizi et al. 2014) and in fish (Thresher et
81	al., 2014b). In part because of these developments, and in part because most well-
82	established invasive species still cannot be effectively controlled using conventional
83	techniques, interest in "genetic biocontrol" has surged (e.g., Saey, 2015; Hall, 2017;
84	Owens, 2017; Prowse et al., 2017). Over this same period, a considerable amount of
85	work has been done on lamprey genetics, including full sequencing of the Sea
86	Lamprey genome (Smith et al., 2013), analysis of gene functions (e.g., Parker et al.,
87	2014), the development of tools for manipulating gene expression (Heath et al., 2014;
88	Romasek et al., 2015) and transgenic gene insertion (Kusakabe et al., 2003). These
89	developments greatly increase the possibility of identifying and manipulating genes
90	that could potentially affect the survival, fertility, and sexual differentiation of GL Sea
91	Lampreys.

92	In this paper, we review and evaluate genetic approaches that could be
93	used to control Sea Lampreys in the Great Lakes. Specifically, we canvas the
94	range of genetic options that could be used against the species, assess the
95	likelihood that any will suppress their abundance (and therefore, their
96	ecological and social impacts) substantially and cost effectively, review the
97	technical feasibility and the risks involved in the deployment of the more
98	promising options, and in a preliminary way, gauge the level of support (or
99	opposition) to research and development of these options and to beginning a
100	wider consultative process that might ultimately lead to their implementation.
101	Specifically, two overarching questions were tested by our analyses:
102	1. Can GL Sea Lamprey abundance be controlled to target levels with a
103	combination of genetic control tactics and other existing tactics using a
104	program that is cost-effective compared to other known alternatives; and
105	2. Can any genetic control tactic for Sea Lamprey management be implemented
106	at levels of risks of unintended consequences that would be judged acceptable
107	by GL stakeholders.
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111	Methods
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113	Four approaches were taken to inform the analysis.
114	Consultation with experts – Over an 18-month period, the authors
115	individually or as a group met with expert Sea Lamprey geneticists and
116	ecologists to discuss the state of genetic research on Sea Lampreys, aspects of

117 their ecology or behaviour that might be exploited using genetic techniques (the 118 "Achilles heel" approach), conservation and management concerns, and 119 suggested genetic options that might be used against the species. Overall, 22 experts contributed to the dialogue. As well, the authors met on two occasions 120 121 with Sea Lamprey management staff of the GLFC to discuss ideas as they 122 developed, and presented status reports and received feedback at the 2016 123 autumn meeting of the GLFC Sea Lamprey Control Board and the 2017 annual 124 meeting of the GLFC.

125 Population modelling - A model of Sea Lamprey population ecology/genetics 126 was used to explore the potential efficacy of different genetic biocontrol options. 127 Model details are provided in the Supplemental Material. Key features of the model 128 are that it simulates two age-structured Sea Lamprey populations, one for the Great 129 Lakes and one "other" (i.e., non-GL) population, which are connected by a small 130 amount of unidirectional parasitic phase migration from the GL population to the 131 other population. The structure is based on the concept of a semi-constrained GL 132 population weakly and intermittently connected to a downstream population, which 133 was incorporated to explore potential non-target effects of genetic control on Sea 134 Lamprey populations beyond the Great Lakes basin (e.g., Finger Lakes or North 135 Atlantic populations). The model is age-structured and assumes constant survival and 136 metamorphosis rates, a Ricker stock-recruitment relationship to predict age 0 137 recruitment and the ability to track the independent genetic dynamics for wild type 138 and genetically modified lampreys, as well as their interaction following biocontrol. 139 Model parameters were informed by prior empirical work on recruitment dynamics 140 (Dawson and Jones 2009) and by a more detailed operating model of Sea Lamprey 141 management (Jones et al. 2009). The model operates at the spatial scale of an entire

Great Lake and a single, separate downstream population. The Great Lake population was initialized at a low population level where density dependence is very limited (to reflect the current situation with GL Sea Lamprey given the existing control program). The model was originally developed to explore the expected performance of a Mendelian sex ratio drive tactic (aka a "daughterless construct") and was adapted for this project to consider four other genetic biocontrol options.

148 Risk Assessment - On 13 and 14 October 2016, the authors hosted a risk 149 assessment workshop at the Quantitative Fisheries Center, Michigan State University, 150 on possible genetic options to manage Sea Lampreys in the Great Lakes. Eight 151 experts attended, along with authors, representing a wide range of specialist skills. 152 These included GL invasion biology, genetic technology and biocontrol, the human 153 health implications of genetic techniques, risks and processes underlying horizontal 154 and vertical gene transfer, lamprey developmental genetics, physiology, ecology and 155 control, Sea lamprey reproductive biology and population structure, and Sea Lamprey 156 management in the Great Lakes. An agenda and background document on the genetic 157 options was circulated to panellists prior to the workshop. The workshop was 158 structured around a) an overview of qualitative and semi-quantitative risk assessment 159 methodologies, b) discussion about and evaluation of genetic approaches overall (e.g., 160 genetic transformation and delivery methods and potential application to Sea Lamprey 161 control), and then c) detailed analysis of six "focal" options (see below), followed by 162 d) a broader discussion of overarching elements associated with the use of genetics to 163 control GL Sea Lamprey. Each focal option was briefly summarized by the authors, 164 including salient modelling results where appropriate, and discussed by the panel. 165 Participants then individually scored and commented on potential adverse effects on:

- Native (non-Sea Lamprey) lamprey species in the Great Lakes (encompasses
   all native species including those with conservation concern, such as Silver
   Lamprey and Northern Brook Lamprey)
- 169
  2. The natural (and valued) Sea Lamprey population in the North Atlantic (and
  170 possibly those in non-GL freshwater bodies, some of which may be natural)
- 171 3. Other fish and non-fish species in the Great Lakes (and possibly wider) region,

172 4. Human health

- 173 5. Other social and ecological endpoints, e.g., cultural issues, recreational uses,
  174 water use, ecosystem function
- 175 Risk estimates were scored by panellists as 1 through 7, where 1 (extremely low) =

176 probability of occurrence during the control program < 1 in 1,000,000; 2 (very low) =

177 > 1 in 1,000,000 to < 1 in 10,000; 3 (low) = > 1 in 10,000 to < 1 in 100; 4 (moderate)

178 = 1-10%; 5 (high) = 10-50%; 6 (very high) = 50-95% and 7 (extremely high) = >95%

179 probability of occurrence. Following initial scoring for each option, panellists

180 revealed their scores, participants with the highest and lowest scores explained their

181 logic and all panellists were given the option of changing their scores in light of the

182 discussions. Finally, panellists were asked to score each option by their overall

183 assessment of the risk of that option. Given their collective expertise, panellists were

also asked to comment on and score the logistical and technical feasibility of each

185 focal option.

<u>Stakeholder and community consultation</u> – Along with informal feedback at
the meetings of the Sea Lamprey Control Board and Great Lakes Fishery
Commission, fisheries managers, scientists and members of the Lake Superior and
Lake Huron fishing communities from Canada and the U.S. were formally consulted
about the acceptability of using genetics actively to control GL Sea Lampreys. Based

191 on the Committee on Gene Drive Research in Non-Human Organisms (2016), the 192 respondents were categorized as "stakeholders", which included professional state, 193 provincial, federal, regional and tribal biologists, fishery managers, the GLFC 194 Commissioners and staff, academic and government scientists, and Canadian and U.S. 195 members of the GLFC citizen advisory groups, all of whom have some involvement 196 in managing Sea Lamprey, and the "fishing community", which included avid 197 recreational fishers who would be broadly familiar Sea Lamprey impacts, the ecology 198 of the lakes, and on-going efforts to manage the problem, but not directly connected 199 with the GLFC and its management activities. With the exception of a few scientists, 200 none of the survey participants had apparent backgrounds in biotechnology beyond 201 basic information gleaned from the media.

202 Two on-line surveys were conducted (SurveyMonkey Inc., San Mateo, CA, 203 USA, <u>www.surveymonkey.com</u>). The stakeholder survey was conducted in 204 March/April, 2017 with a response rate of 73% (95 returns from 131 individuals 205 contacted). The fishing community survey was conducted in August/Sept. 2017, with 206 a response rate of 51% (49 of out 96 individuals contacted). All responses were 207 anonymous and the same questionnaire was used for both surveys. It consisted of 208 three sections: (1) questions about the respondent's background and their perception 209 of the importance of managing Sea Lamprey, (2) a brief description of each focal 210 option, including a summary of risks as determined by the expert panel, followed by 211 a) questions regarding level of support for research and development (R&D) on that 212 option and for beginning a consultative process that could lead to its' implementation, 213 and b) a checklist of generic objections (e.g., cost, unethical, other options available, 214 human health risks), along with a space to list "other" objections, and (3) a free-form 215 section soliciting comments on any subject relevant to the issue, but in particular

216	reasons why the respondent favoured or opposed the use of biotechnology to manage
217	Sea Lamprey in the Great Lakes.
218	Non-parametric statistics were used to analyse the data, using Staview.
219	Regression tree analysis (De'ath and Fabricius 2000), using the R-package 'tree'
220	(Ripley 2016), was used to identify key concerns that distinguished between those
221	supporting and those opposing the use of each focal option.
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223	Results
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225	The initial scope of the project was deliberately broad (Table 1),
226	spanning concepts suggested in the literature and ideas solicited from experts
227	and the wider community. Two approaches were subsequently excluded from
228	further consideration. One line of genetic biocontrol research focuses on using
229	modified pathogens or parasites to suppress pest populations (Cowan, 1996;
230	Hardy et al., 2006), including disease-vectoring mosquitoes (Hoffmann et al.,
231	2011). Based on initial discussions with stakeholders and the GLFC, and
232	adverse public reaction to this approach elsewhere when applied to vertebrates
233	(Hardy et al., 2006), we did not evaluate it for future Sea Lamprey applications.
234	An alternative approach, broadly referred to as "autocidal" (Gould and
235	Schliekelman, 2004; Thresher, 2008), involves genetically modifying the pest
236	itself such as to reduce its impacts. One such set of autocidal options involves
237	manipulating parental chromosomes (e.g., triploidy and YY males) to reduce
238	population fecundity or distort population sex ratios (Gutierrez and Teem, 2006;
239	Erickson et al., 2017; reviewed by Thresher et al., 2014a). These options require
240	obligate chromosomal sex determination with few, if any, autosomal sex

241	modifiers. Available information, though not definitive, suggests weak
242	chromosomal sex determination in Sea Lampreys, with an individual's sex
243	determined by environmental cues or growth rates (Docker and Beamish, 1994;
244	Johnson et al., 2017). Consequently, we have also excluded chromosomal
245	approaches from our evaluation.
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247	Evaluation of genetic options
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249	Modelling, plus the initial discussions with lamprey and GL biologists and
250	managers, resulted in the diverse genetic options initially considered being narrowed
251	down to six "focal" options. All appear to be logistically and technically feasible, and
252	all could potentially suppress Sea Lamprey population abundance and associated
253	impacts in the Great Lakes.
254	
255	Heritable (Mendelian) sex ratio drive – Among the numerous genetic
256	biocontrol options being considered globally, one that heritably distorts offspring, and
257	hence population, sex ratios is considered the most effective (Bax and Thresher,
258	2009). Specifically, the approach is based on a genetic construct that is carried by
259	individuals of one sex with no adverse effects but when passed on to offspring
260	sterilizes or kills members of the other sex, or that results in obligate male or female
261	development, irrespective of genotype. Theory and modelling indicate that for most
262	species, a construct that is lethal to females or that causes females to develop as
263	phenotypic males ("daughterless") is more effective than a "sonless" construct, as the
264	number of females typically constrains population fecundity (Hamilton, 1967).
265	Application of this technique requires identification of genes that are either sex-

266 specific or that determine phenotypic sex. Sexual differentiation in lampreys may be 267 polygenic, in which individuals with high numbers of "sex determining genes" 268 become one sex and those with fewer such genes become the other (M. Docker, 269 University of Manitoba, Winnipeg, Manitoba, personal communication, 2016). If so, 270 then a knockout of a mammalian SRY-equivalent (Sekido and Lovell-Badge, 2009) in 271 lampreys may not be possible. However, targeting key molecules further along the 272 differentiation pathway, perhaps equivalent to aromatase in teleosts, may be feasible. 273 Aromatase converts androgens to estrogens in teleosts and, when blocked chemically 274 (Piferrer et al., 1994; Kwon et al., 2000) or genetically (Thresher et al., in prep.), can 275 cause individuals to develop as males irrespective of genetic sex. Aromatase is absent 276 in lampreys (M. Docker, University of Manitoba, Winnipeg, Manitoba, personal 277 communication, 2016; W. Li, Michigan State University, Lansing, Michigan, personal 278 communication, 2016) but there may be analogues. The genes involved in sexual 279 differentiation in Sea Lamprey have not yet been identified, but considerable work has been done on their reproductive physiology (reviewed by Sower, 2015). The 280 281 complex set of steroidogenic pathways in the lamprey reproductive system differs 282 chemically from those of jawed vertebrates and could offer options for manipulating 283 sex ratios in an agnathan-specific fashion. Alternatively, field data suggests that 284 gender in Sea Lamprey is often growth rate dependent (see below), suggesting that 285 offspring sex ratios could be manipulated indirectly by genetically down- or up-286 regulating rates of larval growth using, for example, CRISPR-Cas9 knock-outs or 287 knock-ins (Square et al., 2015; Zu et al., 2016) of growth hormones. 288 The use of a sex-biasing construct inherited via Mendelian processes to 289 suppress invasive species would require high stocking rates of individuals with many

290 independently segregating copies of the blocking construct (Schliekelman et al., 2005;

291 Bax and Thresher, 2009; Thresher et al., 2014a). However, this strategy assumes 292 strong chromosomal sex determination and an equal sex ratio. Neither assumption 293 may hold for Sea Lamprey in the Great Lakes. Laboratory and field data provide 294 evidence for environmental sex determination in lampreys (Beamish, 1993; Docker 295 and Beamish, 1994), possibly driven by population density and consequent variations 296 in larval growth rates. Faster growing, earlier maturing individuals are predominantly female (Johnson et al., 2017). As a likely consequence, long term data from the Great 297 298 Lakes indicate a shift from an initial pronounced male bias among Sea Lamprey 299 populations at the start of control efforts using lampricides to one dominated by 300 females after effective control had been achieved (Purvis, 1979). By the late 1970s, 301 females constituted an average of about 70% of the adult Sea Lamprey population in 302 the upper Great Lakes. More recent work suggests populations may have shifted back 303 to a roughly 50:50 male-to-female sex ratio (Hanson et al., 2016), though the authors 304 note the inherent difficulties of sampling sex ratios accurately. Data on sex ratios of 305 Sea Lamprey populations native to the North Atlantic are sparse. Beamish (1980a) 306 reports males outnumbering females in Canadian Sea Lamprey populations. Recent 307 qualitative estimates suggest roughly equal sex ratios in small samples of Sea 308 Lampreys collected in the north-eastern US (S. Sower, University of New Hampshire, 309 Durham, New Hampshire, personal communication, 2016). In contrast, Beaulaton et 310 al. (2008) reports a female bias in Sea Lamprey populations in French rivers. 311 Our modelling indicates that release of individuals carrying conventionally 312 (Mendelian) inherited constructs that dictate the male phenotype or are lethal to 313 females can effectively suppress GL Sea Lamprey populations, consistent with 314 models based on other taxa (Fig. 1A, Table 2). In contrast with other studies (Bax & 315 Thresher, 2009) we found that a female-lethal construct was somewhat less effective

316 than a male-specifying (daughterless) construct. For a given stocking rate of 317 genetically modified (GM) individuals, we identified two other factors that affect the 318 efficacy of a sex-ratio-distorting program. First, effective suppression depended on 319 Sea Lamprey abundance already being at low levels as a result of other, coincident 320 control actions (e.g., lampricides or barriers). At higher abundances, compensatory 321 processes severely limit the effect of sex ratio distortion on future recruitment. 322 Second, the sex-ratio of the target population had a large impact on suppression (Fig. 323 1B, Table 2). If sex ratios are biased towards females in low density populations, then 324 female numbers, and hence population fecundity, will decline as genetically modified 325 individuals that would otherwise have been females develop as phenotypic males. 326 The smaller the target population, the more the daughterless carriers dominate the 327 male population and the smaller the remaining number of females, resulting in a 328 ratchet that will eventually drive the population to extinction. The more female-329 biased the wild type sex-ratio, the stronger the impact of a daughterless construct. For 330 populations in which sex ratios are roughly equal, the impact of stocking daughterless 331 carriers is far less, and is reversible if stocking is discontinued.

332

333 <u>Trojan Gene</u> – A Trojan Gene is a construct that pleiotropically has a positive 334 effect on one or more fitness components, and negative effects on others. An example 335 would be a gene that increases mating advantage or attractiveness while decreasing 336 the viability of genetically modified offspring. The concept was developed and 337 modelled by Muir and Howard (1999, 2002), who noted that the use of genetic 338 sterility to contain genetically (or otherwise) modified animals could have a severe 339 adverse effect on wild type populations if the construct also increased carrier 340 competitiveness for mates. However, they also noted that a Trojan Gene could a

useful tool for managing invasive pests (Muir and Howard, 2004). Bax and Thresher
(2009) further modelled Trojan Genes as a method of pest control, confirming
predictions by Muir and Howard (1999, 2002), but also noted that there were tight
constraints on the combinations of partial sterility and mate attractiveness that could
result in eradication of a wild type population.

346 The use of technology analogous to a Trojan Gene to control GL Sea Lamprey 347 populations was suggested by Li et al. (2003b), based on genetically increasing the 348 attractiveness of males that had been independently, artificially sterilized using 349 Bisazir (Siefkes et al., 2003; Twohey et al., 2003). Diverse genes can potentially be 350 targeted to induce sterility in Sea Lamprey, though none have been described thus far. 351 In other taxa, including teleosts, disrupting expression of genes associated with 352 spermatogenesis and oogenesis constitute tissue- and stage-specific targets for 353 reducing fertility or causing full sterility (Hardy et al., 2006; Klein et al., 2012). 354 Analogues in Sea Lamprey are highly likely. As well, the 15alpha hydroxylated steroids, apparently unique to lampreys (Sower, 2015), constitute appealing targets for 355 356 inducing lamprey-specific sterility if other effects on sexual development and 357 reproduction are minimal. Increasing mating success in male individuals could be 358 accomplished by genetically up-regulating the production of pheromones that attract 359 females to nest-tending males (Li et al., 2003a). The physiological factors that limit 360 pheromone production are not yet clear, but there are a number of possibilities that 361 could be explored (W. Li, Michigan State University, Lansing, Michigan, personal 362 communication, 2016). Whether pheromone production can be increased to the point 363 of substantially increasing the mating success of individual males in the complex 364 topography of nesting males and migrating females in streams is also not yet clear.

365 We modelled this option assuming Trojan males are sterile. Allowing the 366 males to be partially fertile results in an on-going, non-hatchery production of Trojan 367 males, but the benefits are relatively small (Bax and Thresher, 2009). The effect of 368 releasing sterile males depends on the number released, their mating competitiveness 369 and the duration of the release program (Fig. 1C and D). If there is no increase in mating success, the program is the GM equivalent of the previous sterile male release 370 371 program that relied on chemosterilization (Twohey et al., 2003), although with 372 differing costs and bio-hazard risks. Under such conditions, a 98% reduction in GL 373 Sea Lamprey abundance within 50 years requires an annual release of approximately 374 1 million sterile male larvae (Table 1). Increasing the mating competitiveness of the 375 released males threefold reduces the required stocking rate to 200,000 males/yr (Table 376 2).

377

378 Development of a non-parasitic Sea Lamprey – Lampreys frequently occur as 379 species pairs, one of which is anadromous and parasitic and one of which is non-380 migratory and non-parasitic (Docker, 2009). The taxonomic status of such species 381 pairs, which typically are identical as larvae and differ only as adults (parasitic forms 382 typically larger) is ambiguous, though recent work by Mateus et al. (2013) finds 383 significant differences in allelic frequencies in a European species pair. The 384 differences appear to relate to genes associated with the migratory ability of the 385 parasitic form.

The ubiquity and apparently small genetic differences between parasitic and non-parasitic forms suggest a relatively simple genetic "switch" between the phenotypes, possibly related to age at sexual maturation (M. Docker, University of Manitoba, Winnipeg, Manitoba, personal communication, 2016). Whether or not

390 such a "switch" occurs in Sea Lampreys is not known, but if so, it may be possible to 391 trigger it prior to metamorphosis either in a genetically modified integrated line or 392 through mass transformation. Further, if parasite population sizes are constrained by 393 density-dependent competition at the ammocoete stage, which is not certain, then 394 release of a heritable non-parasitic form could sustainably reduce numbers of 395 metamorphs of the parasitic form. The magnitude of the reduction would depend on 396 the relative competitiveness of the parasitic and non-parasitic forms and the extent to 397 which the latter can build up high densities in individual drainages. Whether the non-398 parasitic form could displace the parasite within the Great Lakes in general would 399 depend on 1) hybridization between the two phenotypes, which might be low given 400 likely size differences between adults of the parasitic and non-parasitic forms and a 401 tendency for size-selective mating in lampreys (Beamish and Neville, 1992); 2) the 402 construct being dominant (haplosufficient); and 3) the construct conferring a fitness 403 gain high enough to offset the likely much higher individual fecundity of the larger 404 parasitic females.

405 We did not model this approach, given the large uncertainties in larval ecology 406 and the effects of competition not only among Sea Lamprey larvae, but potentially 407 with the larvae of sympatric native lampreys and other stream taxa. Dawson and 408 Jones (2009) did not find evidence for negative effects of high competitor density on 409 Sea Lamprey recruitment, and concluded that higher competitor density was 410 indicative of better rearing habitat in the streams being investigated. Nevertheless, if 411 GL Sea Lamprey ammocoetes compete with those of native species, then it might be 412 possible to reduce population abundance without genetic manipulation by augmenting 413 the abundance of native non-parasitic taxa.

414

415 Sustained release of sterilizing or lethal molecules in nursery drainages -416 Molecular techniques hold promise for developing cost-effective slow release, 417 species-specific biocides, sterilizing agents or reproductive disrupters. A range of 418 molecular blockers (zincfinger nucleases, morphelinos, microRNAs, long hairpin 419 RNAs) are well documented to block gene expression in vivo and have been widely 420 used experimentally to determine gene function (e.g., Meng et al., 2008; Bill et al., 2009). However, the most widely applicable and effective molecular means of gene 421 422 knockdown is short interfering RNA (siRNA), which is widely used for therapeutic 423 applications in human health (Kim and Rossi, 2007) and, in agriculture, to suppress 424 pest impacts through endogenous production in plant tissues of siRNAs that are toxic 425 to pest insects (Kim et al., 2015). Similar approaches appear to be applicable to Sea 426 Lamprey. Short interfering RNAs delivered orally in a mixture of a transfection agent 427 and yeast cells (normal food for cultured ammocoetes) not only delivered the siRNAs 428 to feeding Sea Lamprey larvae, but also resulted in significant larval mortality (Heath 429 et al., 2014). The proof-of-concept study suggests that oral delivery of suitable 430 siRNAs could be used for a species-specific knockdown of Sea Lamprey larvae in 431 nursery drainages throughout the Great Lakes.

432 Four factors need to be addressed before siRNAs can be used in the field. 433 First, although widely cited as sequence-specific (and hence species-specific), partial 434 matching of the siRNA nucleotide sequence can result in off-target effects (e.g., Jackson et al., 2003; Lundgren and Duan, 2013). Off-target effects, in the form of full 435 436 or partial suppression of non-targeted genes, could be irrelevant or even useful if the 437 intent of the siRNA treatment is high levels of larval mortality. However, expression 438 due to partial sequence matching (as few as 6 nucleotides) could reduce the degree of 439 species-specificity and hence result in impacts on taxa other than Sea Lamprey. This

440 risk can be minimized in the design of the construct (e.g., Kamola et al., 2015). 441 Second, disrupting genes, such as those involved in sexual differentiation, require 442 organism-wide (systemic) effects of the siRNA molecules. Whether this is readily 443 achievable when the molecules are delivered in the larval diet is not clear. If impacts 444 are localized only in gut tissues (Heath et al., 2014), suitable targets might be limited 445 to gut-specific genes, many of which are likely to be lamprey specific (e.g., Conlon et 446 al., 1993). Third, the persistence of naked siRNA molecules in the aquatic 447 environment is not known, but could be on the order of hours (Dubelman et al., 2014). 448 If so, effective delivery could need to be in the form of microencapsulated particles 449 (Peanparkdee et al., 2016; for a fish example, see Yufera et al., 1999) of the correct 450 size to be ingested by filter-feeding ammocoetes. The particles could be dispersed 451 manually, or slow-released in the form of slowly dissolving blocks placed into 452 drainage headwaters (S. Whyard, University of Manitoba, Winnipeg, Manitoba, 453 personal communication, 2016.). The latter option would result in long-term 454 accumulation of a lethal, sex determining or sterilizing agent in ammocoetes with a 455 large and potentially cost-effective flow-on effect to parasite population numbers. 456 Fourth, such a strategy requires large scale production of siRNAs. Therapeutic 457 applications of siRNAs involve minute amounts of the molecules. Larger amounts of 458 siRNAs can be produced using bacterial and viral "biofactories" (e.g., Aalto et al., 459 2007), possibly at relatively low cost. 460

An alternative approach, which could achieve permanent reduction in GL Sea Lampreys at negligible sustained cost, would involve genetically engineering an alga or bacteria that is naturally fed upon by the ammocoetes (Dawson et al., 2015) to produce the siRNA. The molecule could be delivered either in the bacteria or alga themselves or in the organic detritus derived from such sources. The approach is

465 analogous to the *in vivo* production of siRNAs in crop plants to suppress insect pest 466 damage, noted above and reviewed by Kim et al. (2015). Methods for genetically 467 modifying algae, bacteria and viruses are very well developed (reviewed by Mendoza 468 et al., 2016) and recently one genetically modified alga was successfully field trialled 469 (Szyjka et al., 2017). Species-specificity, as well as efficacy, would need to be 470 rigorously confirmed and, if an algal or bacterial host was used to express and 471 permanently distribute the agent, it would be critical to use a species found only in 472 Great Lakes drainages as a vector.

473 The magnitude of the impacts of a synthetic blocking molecule on parasite 474 populations depends on the efficacy of the blocking and the number of drainages into 475 which it is applied. Modelling suggests that even a sustained 10% increase in rates of 476 ammocoete mortality over those currently achieved using existing biocidal treatments 477 could substantially depress parasite abundance; a 20% increase essentially results in 478 virtual eradication of the GL Sea Lamprey population within 50-75 years (Fig. 2A, 479 Table 2). Direct impacts on non-target populations would be nil if the construct was 480 species-specific and delivery was in the form of manual dispersal or slow-release 481 blocks in nursery area headwaters. Impacts could be more significant if an alga or 482 bacteria was modified to produce and deliver the agent, depending on the distribution 483 of the vector.

484

<u>Vaccinated prey</u> – Parasitism by Sea Lamprey involves post-metamorphic preadults attaching to a host fish, inflicting a non-lethal wound, and consuming blood
and other body fluids. Smaller hosts frequently do not survive the attack due to loss
of host fluids, whereas larger ones frequently survive (Jorgenson and Kitchell, 2005)
and, on the basis of multiple scars, can be attacked again. Fatty acid analysis of

490	lampreys (Happel et al., 2016) and their activity cycles (Bergstedt and Swink, 1995)
491	suggest each individual feeds on multiple hosts, but the number of hosts is not known
492	with certainty. For our analysis, we use ten as a plausible first estimate, based on
493	Table 4 in Bence et al. (2003).
494	The ectoparasitic feeding mode suggests that Sea Lamprey could be
495	vulnerable to molecules in the blood stream of their fish hosts that are lethal to,
496	sterilize or cause morbidity in the parasite. As noted above, a likely candidate set are
497	short interfering RNAs (siRNAs), which have already been demonstrated to affect
498	lampreys when consumed (Heath et al., 2014). Potential targets for siRNAs include
499	lamprey-specific anti-coagulants injected by lampreys into their hosts to facilitate
500	feeding (Ito et al., 2007), and gut-specific genes, many of which are also likely to be
501	lamprey specific given their unusual feeding ecology (e.g., Conlon et al., 1993).
502	Down-regulation of either target could reduce lamprey feeding efficacy and lead to
503	parasite morbidity, increased mortality, or reduced fecundity and/or mating success as
504	adults. If systemic up-take is feasible, mortality may be achievable. Zu et al. (2016)
505	showed that disrupting expression of the gene kctd10 in Sea Lampreys resulted in 55-
506	85% of the injected animals showing severe heart defects, which presumably would
507	lead to mortality in field situations. More broadly, two gonadotropic releasing
508	hormones (GnRH-I and III) in Sea Lampreys not only differ in protein structure from
509	those of gnathostome vertebrates, but when blocked experimentally result in
510	approximately 60% of males being sterile (Sower, 2003). The experiments were not
511	"optimized" in terms of the experimental GnRH antagonists involved, which suggests
512	that inducing higher rates of sterility is possible (S. Sower, University of New
513	Hampshire, Durham, New Hampshire, personal communication, 2016). Agnathan-
514	specific hormones critical to sexual maturation appear to be promising targets for

515 orally sterilizing the parasites with no adverse effects on the construct-carrying hosts 516 and on other GL vertebrates that might consume the hosts, including humans. The 517 DNA sequences of GnRH-I and III have been published (Suzuki et al., 2000; Silver et 518 al., 2004), facilitating design of siRNA blockers. Alternatively, high rates of lamprey 519 sterility could be induced by up-regulating, for example, thiaminase production on 520 one or more host species. Recent observations suggest that a high proportion of 521 female Sea Lamprey collected in Lake Michigan are functionally sterile, producing 522 low quality eggs and larvae (D. Medeiros, University of Colorado, Boulder, Colorado, 523 personal communication, 2016; D. McCauley, University of Oklahoma, Norman, 524 Oklahoma, 2017) reminiscent of the effects of orally consumed thiaminase on Lake 525 Trout reproduction (Jarszewska et al., 2009). The cause of the low fertility is not 526 known, but that Sea Lamprey consume bodily fluids of Lake Trout and other 527 salmonids that can be high in thiaminase is perhaps worth examining in more detail 528 (Dabrowski et al., 2004). The effects of injected molecules on the survival and 529 fertility of Sea Lamprey could be tested experimentally either using an artificial 530 feeding method ("blood bags", originally suggested for lamprey trials in the 1990s, 531 but not subsequently developed) or injected into hosts that are subsequently fed to the 532 parasites.

Methods for genetically modifying some host species (e.g., Rainbow Trout, Carp) are well developed and in routine use (Iyengar et al., 1996), including mass transformation techniques (Powers et al., 1992). It is highly likely these techniques could be readily adapted for use in other host species, e.g., Lake Trout or Lake Whitefish. Host species selection is likely to be highly critical for this technique to be successful, both in terms of maximizing the likelihood of a Sea Lamprey attacking a molecule-carrying target and in terms of public acceptability (see below). An ideal

540 host would be one routinely attacked by Sea Lamprey, but not native parasitic species 541 (e.g., Silver Lamprey) and not consumed by humans, due to perceptions of possible 542 health risks. Although Sea Lamprey attack a range of host species (Happel et al., 543 2016), Lake Trout appear to be a preferred host. Lake Trout are also hatchery 544 produced and stocked in large numbers into most of the Great Lakes, such that the 545 infrastructure required to produce and release large numbers of carriers is mostly 546 already in place. Wounds on the fish suggests that the siscowet Lake Trout of Lake 547 Superior may be a useful possibility, in that the form is relatively sluggish, inhabits 548 deep-water, may be a preferred target for parasitism (Zhuikov, 2016) and, being high 549 in fat, is not preferred by recreational and commercial fishers. The deep-water habitat 550 of the fish overlaps that of Sea Lamprey, but not that of Silver Lamprey, which prefer 551 warmer water and apparently feed inshore (Cochran and Marks, 1995). Siscowet are 552 also morphologically different from other Lake Trout strains (smaller head, deeper 553 body, blunter convex snout, shorter, thicker caudal peduncle) which could facilitate 554 avoidance of the carriers by fishers, if desired, in lakes where the strain is not native. 555 Depending on the stocking strategy (see below), stocked carriers could also be tagged. 556 We modelled the potential effect of vaccinated hosts by adjusting downward 557 Sea Lamprey survival rates at the parasitic life stage to reflect plausible levels of 558 additional mortality due to the genetically modified hosts (Fig. 2B). Assuming 10 559 attacks per parasite, the probability of a parasite attacking a vaccinated host if 4% of 560 hosts were GM type (i.e., 200,000 GM hosts in a population of 5,000,000) would be 561 34%. Assuming a single feed is lethal to or sterilizes a parasite, this would translate 562 into an equivalent reduction in parasitic survival (ignoring possible compensatory 563 effects). Simulating a 34% reduction in parasitic survival, together with continuation 564 of existing control measures, resulted in a 92% population reduction in 50 years

565 (Table 2). If parasites on average attack fewer than 10 hosts, the proportion of GM 566 hosts would need to be higher to achieve the same outcome. A positive aspect of the 567 vaccinated prey option is that if the GM host is a freshwater species there is no risk to 568 native North Atlantic Sea Lamprey populations.

569 The combination of the relatively small number of carriers required to achieve 570 substantial GL Sea Lamprey control, existing hatchery facilities and available 571 expertise suggest it may not be necessary to create a lineage of genetically modified 572 hosts to achieve control objectives. Mass transformation techniques, such as 573 electroporation (Powers et al., 1992) and biolistics (Zelinin et al., 1991) applied to 574 eggs and early stage larvae, can produce very high numbers of carrier phenotypes 575 quickly and at low cost. Intramuscular injection of constructs, though more labor 576 intensive, can also produce salmonids capable of expressing a construct for up to 1.5 577 years post-injection, though this is primarily realized only near the point of injection 578 (Anderson et al., 1996; Seternes et al., 2016) without integration (i.e., germ line 579 transformation). Potentially, hosts could be sterilized using heat or pressure-shock 580 induced triploidy, then transformed en masse using electroporation for subsequent 581 grow out, tagging and release. Given existing hatchery capability and stocking efforts 582 for, e.g., Lake Trout, even if only a proportion of the treated fish proved to be 583 effective carriers, it should be possible to achieve the 6-7% carrier stocking rate 584 required to suppress parasite abundance without genetically contaminating existing 585 host stocks. Such an approach would also ensure that the carrier phenotype 586 disappears at the end of the control program, because they are no longer stocked. 587

588 <u>Gene-driven sex ratio distortion</u> – As noted above, the release of a normally
589 (Mendelian) inherited gene construct that causes male or female-specific lethality,

590 sterility or obligate phenotypic development can result in long-term population 591 decline, irrespective of native population sex ratio. The magnitude and efficiency of 592 the approach depends on the number of independently segregating copies of the 593 construct and the rate at which carriers are stocked into the targeted population 594 (Schliekelman et al., 2005; Bax and Thresher, 2009). Achieving high copy numbers 595 and high stocking rates can be technically and logistically challenging. Burt (2003) 596 suggested an alternative approach, based on "selfish genes". Homing endonucleases 597 are a class of genes that when inherited as a single copy (heterozygousity) duplicate 598 themselves onto the complementary chromosome, thereby becoming homozygous 599 (Gimble and Thorner, 1992). As a result, homing endonucleases are potentially 600 inherited by all of an individual's offspring, rather than only half, and the construct 601 spreads rapidly in the population so long as any fitness cost due to the construct is less 602 than the gain due to producing more than the normal number of carrying offspring 603 (Burt, 2003; Deredec et al., 2008; 2011). Burt (2003) suggested than such "gene 604 drives" could be used to efficiently engineer natural populations to, for example, 605 render than incapable of transmitting disease-causing parasites or, in the case of an 606 invasive species, distort sex ratios to suppress pest populations. In theory, the release 607 of a single individual carrying a gene drive could permanently alter or suppress a 608 targeted population (Saey, 2015; Webber et al., 2015). The logistical advantages 609 gained by use of a gene drive for pest population suppression are substantial, as are, 610 potentially, the risk to non-target populations and species if the construct spreads from 611 the targeted population (Webber et al., 2015; Esvelt and Gemmel, 2017), either 612 through normal vertical transmission or horizontal gene transfer (Kuraku et al., 2012). 613 Whether or not this risk is realized is uncertain at this stage, due to strong selection 614 for resistant individuals, and could depend on the design of the gene-drive construct

and the sequence targeted by it (Hammond et al., 2017). Strong counter-selection
could also dilute and possibly fully negate the gain in efficacy and the ultimate impact
of a gene-driven system on targeted populations

618 Burt and colleagues have been developing the concept for the control of 619 Anopheline mosquitoes, using a modified naturally occurring homing endonuclease 620 (Windbichler et al., 2011; Klein et al., 2012). The need to use naturally occurring 621 drive elements limits the scope of potential targets and also the efficacy of the 622 approach, as the rate of gene duplication (homing) in naturally occurring systems is 623 typically well below 100% (Chan et al., 2011; Windbichler et al., 2011). Recently, 624 however, "synthetic" gene drives have been developed, based on variations of the 625 bacterially-derived CRISPR gene editing system (Esvelt et al., 2014). CRISPR-based 626 gene drives are not only widely applicable, but can achieve homing rates in excess of 90% (Gantz and Bier, 2015; Gantz et al., 2015). The potential of synthetic gene 627 628 drives for cost-effectively engineering wild populations has received considerable 629 attention in the technical and popular media (Champer et al., 2016; Committee on 630 Gene Drive Research in Non-Human Organisms, 2016; Harmon, 2016), including its 631 potential to suppress pest populations currently deemed unmanageable (Hammond et 632 al., 2016; Szymczak, 2016; Hall, 2017; Prowse et al., 2017). However, the threat such 633 technology poses to non-target species has resulted in recommendations for rigorous 634 containment protocols for laboratory studies of the technology (Araki et al., 2014; 635 Oye et al., 2014), and a variety of suggestions to reduce the risks to non-target 636 populations. The latter include incorporating a genetic "off-switch" in the construct, 637 genetically modifying the non-target population(s) to be resistant to the suppression 638 drive (Esvelt et al., 2014), and using "daisy drives", in which the essential 639 components of a series of gene drives are released separately, combine to effect

640 population suppression, but then cease to spread as "lower" elements in the drive 641 chain are lost over time (Noble et al., 2016; Esvelt and Gemmel, 2017). CRISPR 642 gene editing techniques, though not a gene drive version as yet, have already been 643 demonstrated as viable in Sea Lamprey (Square et al., 2015; Zu et al., 2016). 644 We adapted our model to evaluate a gene drive by simulating non-Mendelian 645 inheritance of a sex ratio distorting construct. As expected based on generic modelling 646 (e.g., Beaghton et al., 2016), the simulations indicate that even very modest releases 647 of "gene-drive" larvae (i.e., 100 individuals for 1 year only) results in effective 648 eradication of GL Sea Lamprey within 200 years following their release (Fig. 2C, 649 Table 2). This initial model assumes a 100% homing rate (all offspring of a modified 650 parent carry the construct). The small numbers of stocked carriers needed for control 651 suggest a program based on mass transformation of lamprey eggs rather than the 652 logistically much more demanding need for an integrated line of genetically modified 653 carriers. Virtual eradication could be achieved much more quickly with annual 654 releases of carriers similar to the Mendelian sex ratio drive presented earlier (100,000 655 GM larvae/yr for 10 years, Fig. 2C, Table 2). Despite these positive outcomes, under 656 all scenarios the "Other" (non-target) population of Sea Lamprey is also eventually 657 driven to virtual extinction due to the escape of carriers from the Great Lakes, even at 658 very low emigration rates (Fig. 2D). The extent to which this outcome can be 659 avoided by use of more complex gene-drive-based strategies, such as "daisy drives", 660 is not yet known.

661

<sup>662</sup> Risk Assessment

664 Overall, panellists judged the risks of the six focal options as low to extremely 665 low for all five potentially impacted categories, based on median responses (Fig. 3). 666 The group judged to be at highest risk was "other" Sea Lamprey populations, due to 667 possible spread of the gene construct beyond the Great Lakes. The category judged 668 overall to be at least risk (other than "Other") was Human Health. The latter, 669 however, was predicated on the design of a Sea Lamprey-specific construct, rigorous 670 testing for off-target effects, and dosages of the effector molecules that are very low 671 in consumed fish.

672 The magnitude of perceived risks differed widely among the six focal options 673 (Table 3). Note that risks were assessed separately for deploying an artificial source 674 of a synthetic biocide as opposed to genetically modifying an alga or bacteria to 675 produce it, due to their very different risks of spread beyond the Great Lakes. Across 676 all options, "overall" risk for each correlated highly with the score for the group 677 deemed by panellists to be at highest risk (Spearman rank rho = 0.96, p = 0.002). 678 Risk, however, did not correlate with perceived technical/logistical feasibility (rho = 679 0.11, NS). Panellists considered the release of Trojan males to be the least risky 680 option; they considered gene-driven sex-ratio distortion the most risky. All panellists 681 judged the risks of the Trojan gene approach as Extremely Low to Very Low across 682 all risk categories. In contrast, panellists differed widely in their judgements 683 regarding the gene drive, as well as Mendelian sex-ratio distortion and the release of a 684 self-propagating (algal or bacterially vectored) larval biocide. At least one panellist 685 scored each of these three options as Extremely risky (>95% probability of 686 occurrence) for at least one group potentially at risk. 687 The group perceived as at high risk for the gene-driven and Mendelian sex-688 ratio distortion options was non-target (e.g., Atlantic) Sea Lamprey populations. In

689 theory, even one construct-carrying individual could have a long-term, adverse impact 690 of such populations. The central issue and source of uncertainty among panellists for 691 the gene-drive was the extent to which dispersal of construct carriers out of the Great 692 Lakes could be maintained at essentially zero. Our models assume an escapement 693 each year of 0.01% of the Great Lakes Sea Lamprey population. The estimate is 694 crude, but at least some movement down system by Sea Lampreys is suggested by 695 evidence that Lake Trout, at least, occasionally survive going over Niagara Falls (C. 696 Krueger, Michigan State University, Lansing, Michigan, personal communication, 697 2016). More esoteric dispersal mechanisms (e.g., Najarian, 2015), including 698 intentional and unintentional human-mediated release (Johnson et al. 2009a; Drake et 699 al. 2015), also cannot be ruled out.

700 For the Mendelian approach, the main concern and source of disagreement 701 was the uncertain sex-ratio of non-target populations. Although available data 702 suggest the sex ratios of the Western Atlantic population is roughly 50:50 (and hence 703 would be resistant to the Mendelian approach), data are sparse, Atlantic populations 704 have been heavily impacted by pollution, habitat loss and over-fishing (Hansen et al., 705 2016), all of which could lead to population depression and a consequent shift to a 706 female-biased sex-ratio, and data indicate a female bias among some European 707 populations (Beaulaton et al., 2008). Effects of released construct carriers on 708 European populations might be mitigated by limited genetic exchange between 709 Western and Eastern Atlantic populations (Wright et al., 1985; Rodriguez-Munoz et 710 al., 2004), but probably not reduced to zero. Sex-ratios for other, possibly native 711 fresh-water populations (see discussion in Eshenroder, 2014) are not known, and they 712 could also be vulnerable to the Mendelian sex-ratio option (as well as to the gene 713 drive).

714 Experimental work demonstrates that small landlocked Sea Lamprey parasites, 715 in particular, have difficulty adjusting to seawater (Beamish et al., 1978; Beamish, 716 1980b), suggesting a partial osmoregulatory impediment to GL Sea Lampreys 717 surviving in the marine waters of the Atlantic and breeding with native Sea Lamprey. 718 In order to minimize further these risks, the effector construct in GM Sea Lampreys 719 could be designed such that it is linked to expression of an osmoregulatory gene 720 sequence (Mateus et al., 2013; Covolo-Soto et al., 2015). Several such designs are 721 available (e.g., an IRES or 2A fusion peptide that promoted both salt intolerance and 722 sex-ratio distortion sequences). These construct designs, however, would still be 723 vulnerable to failure due to point mutations in the salinity intolerance coding region. 724 A containment mechanism based on salinity intolerance would also not eliminate 725 risks to native freshwater populations. Other containment options for the gene-drive 726 have been noted above, but are either as yet untested (e.g., a multi-component daisy 727 drive) or were judged by the panel as likely to be unacceptable (e.g., genetically 728 engineering the Atlantic Sea Lamprey population to be resistant to a population 729 suppression drive released into the Great Lakes). 730 The third option rated as potentially high risk by panellists was the 731 deployment of a self-propagating larval biocide. Again, opinions were divergent 732 among panellists, with scores for risks against both native lampreys and non-target

733 Sea Lamprey populations ranging from Extremely low (1) to Extremely high (7). The

central issues were the extent to which a genetically modified alga or bacteria could

be limited to Great Lakes drainages, and the species specificity of an siRNA effector

molecule, along the lines discussed above.

The panel also discussed risk elements that span the six options (Table 4).These elements ranged from the risk of horizontal gene transfer, which has been

739 suggested between lampreys and their hosts (Kuraku et al., 2012; Zhang et al., 2014), 740 to the risk of inadequate rapid response capability by lake managers in the case of re-741 invasion following Sea Lamprey suppression. Overall, the risks of vertical and 742 horizontal gene transfers of the constructs were judged to be extremely low in the 743 time frame (decades) being considered for the control program, the risks of off-target 744 effects of the constructs manageable given modern construct design, the risks of 745 escapement of carriers from the Great Lakes high and possibly difficult to manage, 746 and the risk of inadequate response capacity to Sea Lamprey re-invasion uncertain, 747 but possibly high if, after a successful control program, resources were to be diverted 748 to other issues.

749

750 Stakeholder Consultation

751

752 A large majority of the stakeholders (84.7%) and the fishing community 753 (95.9%) supported research on one or more genetic options for controlling Sea 754 Lamprey in the Great Lakes; for steps towards possible implementation, the figures 755 were similar (86.3% and 95.9%, respectively). Details are provided in Thresher et al., 756 (in press). In both surveys, support was widespread across all respondent groups, all 757 age groups and irrespective of years of professional or fishing experience. Among all 758 respondents, the three main reasons given for opposition to genetic approaches overall 759 were concerns about effects on non-target species and populations, the adequacy of 760 safeguards, and insufficient knowledge by the respondent to be comfortable with the idea. Concerns about risks to human health were not widespread, nor were ethical 761 762 concerns about the use of the technology to control Sea Lamprey.

763 At the level of focal options, support was widespread and positive for the 764 Mendelian sex-ratio distortion, Trojan males, and development of non-parasitic Sea 765 Lamprey, divided for vaccinated prey, and negative for a synthetic larval biocide and 766 the use of a gene drive (Fig. 4). Across all options, support levels by stakeholders and 767 the fishing community correlated very highly ( $R^2 > 0.96$  for both R&D and 768 implementation), with the conspicuous exception of vaccinated prey. Stakeholders 769 were split evenly with regard to researching this option (50% opposed), whereas 770 recreational fishers were broadly opposed to it (69% opposed). Overall, costs of 771 project development, ethical issues, potential availability of non-genetic options and a 772 perception that existing methods of Sea Lamprey control were adequate did not rate 773 highly as concerns for any of the focal options, whereas concerns about impacts on 774 non-target species and populations and about the adequacy of safeguards were 775 widespread. Regression tree analyses to identify the primary co-variates (attitudes) 776 that distinguished between opponents of each approach and those supporting it (analyses do not include strong supporters, which as a group expressed few concerns) 777 778 accounted for a relatively small amount of the variance in predicted level of support 779 for each option (mean 30.1% for R&D and 25.5% for implementation). These results 780 indicated that factors beyond those assessed in the survey should account for the 781 unexplained variation in the level of support for R&D or implementation of a given 782 focal option. As we incorporated a large number of diverse explanatory variables as 783 co-variates, it is unclear what additional information would have led to greater 784 variance explained, especially given that perspectives about the overall importance of 785 Sea Lamprey control and the importance of developing alternative control tactics 786 were included as possible predictor variables. In general, identifying the unsupportive 787 fraction of respondents was usually straightforward. For example, in terms of the

788	level of support for future research and development for the heritable sex ratio drive,
789	strong opponents indicated that the approach was unethical; whereas, despite concerns
790	about safeguards, fishery biologists and managers were likely to be supportive of the
791	approach relative to other respondent types, possibly reflecting long-standing
792	frustration with conventional control tactics (Fig. 5). For the gene drive, the belief that
793	non-target risks were too high, and the belief that existing control tactics were good
794	enough, both signified low overall support for R&D.
795	
796	Discussion
797	
798	Our analyses indicate widespread support among key stakeholders and
799	recreational fishers for research on the use of one or more forms of genetic biocontrol
800	to manage Sea Lamprey in the Great Lakes (Thresher et al., in press). The breadth of
801	support is likely to reflect an appreciation of historical and on-going difficulties in
802	managing the pest, environmental consciousness among respondents regarding the
803	impacts of the species on lake ecosystems, and a hope that biotechnology might allow
804	the problem to be "solved" rather than, as is currently the case, managed. With regard
805	to preferred options, we scaled the six focal options, with the larval biocide split into
806	synthetic versus self-propagating sub-options, by risk, technical/logistical feasibility
807	and stakeholder/fishing community acceptability (Fig. 6). The ideal option is one that
808	is low risk, feasible, effective and acceptable to a broad audience of stakeholders and
809	the fishing community. Mendelian sex ratio distortion currently comes closest to
810	fulfilling all four criteria. Vaccinated prey is assessed to be feasible and low risk, but
811	is marginally acceptable as an option for stakeholders, and opposed by most of the
812	surveyed fishing community. The release of Trojan males was both acceptable and a

813 low risk approach, but panelists doubted whether male mating attractiveness could be 814 enhanced enough to have an impact on the GL Sea Lamprey population. The other 815 three options either scored very low in terms of acceptability (synthetic biocides, the 816 gene-drive) or were not considered likely to be effective (non-parasitic Sea Lamprey). 817 This landscape is dictated in part by information gaps. Human health 818 implications of a particular option, as a prime example, are determined by dosage and the nature of the effecting molecules, neither of which is known at this stage. The lack 819 820 of detail not only makes estimating risk difficult even by informed experts, but also 821 has a large impact on community attitudes towards the different options, i.e., human 822 health concerns largely determined opposition to synthetic biocides and vaccinated 823 prey. However, unique aspects of the physiology of the lamprey reproductive and 824 feeding systems could well be exploited to design effector molecules with potentially 825 no impact on humans or any other non-lamprey target. Similarly, the risks associated 826 with, and the efficacy of Mendelian sex ratio distortion will be informed by better 827 data on the current sex ratios of the Great Lakes and Atlantic Sea Lamprey 828 populations, of a gene-drive by developments in containment technology and the 829 actual rates of downstream gene exchange with the Atlantic population, and of the 830 Trojan gene by the extent to which male attractiveness can be enhanced and its effect 831 on mating success in a non-deterministic stream environment. 832 The choice of focal options is also likely to be a function of how that option is 833 deployed. At this stage, the key considerations in assessing the future viability of a 834 biocontrol program are associated with efficacy, risks, and support by stakeholders

and the fishing community. Consequently, our analyses did not focus on

technological limitations, on the basis that given adequate resourcing, suitable gene

837 targets, construct designs and methods for insertion would be found. A key technical

838 challenge for some options, however, could be the need to create and maintain an 839 integrated brood line of genetically modified Sea Lamprey. This is not an issue with 840 synthetic biocides or vaccinated prey (for which germ line modification and 841 husbandry techniques are likely to be readily available). For Sea Lamprey, methods 842 of germ line modification are well developed (Kusakabe et al., 2003). However, in a 843 typical genetic engineering program, several generations of carriers need to be 844 screened and, often, back-crossed to select and fix optimal gene configurations and 845 phenotype. Doing so with Sea Lamprey could take 10-15 years (even in a best-case 846 scenario without complications) due to the prolonged larval stage (3-7 years) and 847 subsequent two-year parasitic and adult stages. Maintenance of a Sea Lamprey brood 848 line is further complicated by the species' semelparous life cycle. A single pair of 849 genetically modified carp can produce large numbers of carrier offspring for decades, 850 whereas Sea Lamprey adults breed once, die and need to be replaced. Consequently, 851 Sea Lamprey breeding programs would need to be carefully managed as not to lose 852 lineages of valuable brood stock.

853 An alternative strategy is suggested by the relatively small numbers of carriers 854 required to achieve Sea Lamprey suppression in the Great Lakes using the sex ratio 855 distorting options. As noted above, in theory, releasing even a single carrier of a gene-856 driven carrier could permanently alter a target population. The models suggest larger, 857 but still logistically manageable numbers, need to be released for the Mendelian 858 approach. Typical integration rates for fish using micro-injection techniques are 3-5% 859 (e.g., Stuart et al., 1988). Several geneticists advised us that micro-injecting DNA 860 into lamprey eggs is relatively easy, and can be done at a rate of 1000s of eggs per 861 hour by experienced staff. At a 3% integration rate and 1000 eggs/hour, 10,000 single 862 copy carriers could be produced by 10 individuals working full time on the project in

863 less than a week. Sub-optimal integration and off-target effects could reduce the 864 efficiency of such a program, but the basic analysis suggests that mass transfecting 865 eggs annually could be a viable alternative to developing and managing Sea Lamprey 866 brood lines. In practice, bulk techniques, e.g., biolistics and electroporation, if 867 suitable for lamprey eggs, could transform 10s of thousands of eggs per hour, making 868 such a program highly effective. Options are also available that can substantially 869 increase rates of transgene integration (e.g., Soroldoni et al., 2009), though with a cost 870 in insertional stability. The program could also benefit from the single generation 871 contribution of episomal expression in non-integrated carriers.

872 Several additional issues need to be factored in to these evaluations of genetic 873 control. First, except under conditions of very high stocking rates, all genetic options 874 take time to affect a targeted population. In the case of the Great Lakes, at logistically 875 feasible stocking rates most options will take 20-25 years to have a discernible impact 876 on the Sea Lamprey population and strong pest suppression could take in excess of 50 877 years. Whether or not the public and managers would invest in a program with such 878 long delivery times is uncertain. Second, options that require carriers be stocked 879 could result in a short duration (ca. 5 years) but nonetheless controversial increase in 880 Sea Lamprey abundance and associated impacts. This could be compensated for, 881 however, by a short-term increase in conventional control methods, e.g., biocidal 882 treatment of nursery habitats. Third, there needs to be community consultation and 883 agreement about the level of ecological and social risk deemed acceptable. Is any 884 threat to the Atlantic Sea Lamprey population acceptable? Although several of the 885 options canvassed were judged by experts as being of low to extremely low risk 886 across multiple endpoints (native lampreys, human health), none are completely risk 887 free and a decision will need to be made on the trade-off between potentially severe
888 adverse effects and the benefits of a possible long-term suppression in the Great 889 Lakes. Fourth and perhaps most importantly, it is not clear at what level and by 890 whom decisions to implement a program should be made, and the extent to which 891 these decisions need to be supported by additional social license research (including 892 broader canvassing of the public as a whole) in concert with future bio-technology 893 R&D. Decision-making in the U.S. context alone could be complicated, due to the 894 number of jurisdictions that will or could be affected and the as yet ambiguous 895 regulatory pathways for genetically modified animals (Otts, 2014). Managing Sea 896 Lamprey in the Great Lakes is inherently a trans-national issue. The possibility that 897 genetically modified Sea Lampreys could escape to and modify permanently, even if 898 not threaten, the North Atlantic population further confounds the international scope 899 of the issue. A compelling argument can be made that European countries should be 900 involved in deciding acceptable levels of risk for genetic biocontrol options that could 901 spread if seeded into the Atlantic population, and consulted for those that should not. 902 International agreements dictate such a process (Garforth and Miranda, 2014; 903 Committee on Gene Drive Research in Non-Human Organisms, 2016). As such, 904 focal options that are inherently confined to the Great Lakes, notably deployment of a 905 synthetic biocide and vaccinated freshwater prey, might warrant greater examination 906 if effector molecules can be unambiguously and rigorously demonstrated to have no 907 impacts on native species or human health. 908

- 909

# 910

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924	
925	References
926	
927	
	Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,
928	Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U., and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for
928 929	<ul><li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li><li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li><li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$6 RNA-dependent RNA polymerase.</li></ul>
928 929 930	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li> <li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li> <li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$\$6 RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> </ul>
928 929 930 931	Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U., and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$6 RNA-dependent RNA polymerase. RNA 13: 422-429.
928 929 930 931 932	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li> <li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li> <li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$6 RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> <li>Anderson, E. D., Mourich, D.V., and Leong JA. C. 1996. Gene expression in</li> </ul>
928 929 930 931 932 933	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li> <li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li> <li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$ RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> <li>Anderson, E. D., Mourich, D.V., and Leong JA. C. 1996. Gene expression in</li> <li>rainbow trout (<i>Oncorhychus mykiss</i>) following intramuscular injection of DNA. Mol.</li> </ul>
<ul> <li>928</li> <li>929</li> <li>930</li> <li>931</li> <li>932</li> <li>933</li> <li>934</li> </ul>	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li> <li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li> <li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$ RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> <li>Anderson, E. D., Mourich, D.V., and Leong JA. C. 1996. Gene expression in</li> <li>rainbow trout (<i>Oncorhychus mykiss</i>) following intramuscular injection of DNA. Mol.</li> <li>Mar. Biol. Biotechnol. 5: 105113.</li> </ul>
<ul> <li>928</li> <li>929</li> <li>930</li> <li>931</li> <li>932</li> <li>933</li> <li>934</li> <li>935</li> </ul>	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U., and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for RNA interference utilizing bacteriophage \$\$\phi6\$ RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> <li>Anderson, E. D., Mourich, D.V., and Leong JA. C. 1996. Gene expression in rainbow trout (<i>Oncorhychus mykiss</i>) following intramuscular injection of DNA. Mol. Mar. Biol. Biotechnol. 5: 105113.</li> </ul>
<ul> <li>928</li> <li>929</li> <li>930</li> <li>931</li> <li>932</li> <li>933</li> <li>934</li> <li>935</li> <li>936</li> </ul>	<ul> <li>Aalto, A. P., Sarin, L. P., Van Dijk, A. A., Saarma, M., Poranen, M. M., Arumae U.,</li> <li>and Bamford, D. H. 2007. Large-scale production of dsRNA and siRNA pools for</li> <li>RNA interference utilizing bacteriophage \$\$\$\$\$\$\$\$\$\$\$\$\$\$\$6 RNA-dependent RNA polymerase.</li> <li>RNA 13: 422-429.</li> <li>Anderson, E. D., Mourich, D.V., and Leong JA. C. 1996. Gene expression in</li> <li>rainbow trout (<i>Oncorhychus mykiss</i>) following intramuscular injection of DNA. Mol.</li> <li>Mar. Biol. Biotechnol. 5: 105113.</li> <li>Ant, T., Koukidou, M., Rempoulakis, P., Gong, HF., Economopoulos, A., Vontas,</li> </ul>

937 J., and Alphey, L. 2012. Control of the olive fruit fly using genetics – enhanced

- sterile insect technique. BMC Biology 10 Number 51:
- 939 www.biomedcentral.com/1741-7007/10/51
- 940
- 941 Araki, M., Nojima, N., and Ishii, T. 2014. Caution required for handling genome
- 942 editing technology. Trends Biotechnol. 32: 234-237.
- 943
- 944 Bax, N. J., and Thresher, R. E. 2009. Ecological, behavioral, and genetic factors
- 945 influencing the recombinant control of invasive pests . Ecol. Applic. 19:873-888.
- 946
- 947 Beaghton, A., Beaghton, P. J. and Burt, A. 2016. Gene drive through a landscape:
- 948 reaction-diffusion models of population suppression and elimination by a sex ratio
- 949 distorter. Theoret. Pop. Biol. 108:51-69.
- 950
- 951 Beamish, F. W. H. 1980a. Biology of the North American anadromous Sea Lamprey,

952 Petromyzon marinus. Can. J. Fish. Aquat. Sci. 37:1924–1943.

- 953
- 954 Beamish, F. W. H. 1980b. Osmoregulation in juvenile and adult lampreys. Can. J.
- 955 Fish. Aquat. Sci. 37:1739-1750.
- 956
- 957 Beamish, F. W. H. 1993. Environmental sex determination in southern brook
- lamprey, Ichthyomyzon greeleyi. Can. J. Fish. Aquat. Sci. 50:1299-1307.
- 959
- 960 Beamish, R., and Neville, C. 1992. The importance of size as an isolating
- 961 mechanism in lampreys. Copeia 1992:191-196.

963	Beamish.	F.	W.	Η.,	Strachan.	Р.	D.	. and T	'homas.	E.	1978.	Osmotic	and	ionic
				,		,		,				• • • • • • • •		

964 performance of the anadromous sea lamprey, *Petromyzon marinus*. Comp. Biochem.
965 Physiol. 60A:435-443.

966

- 967 Beaulaton L., Taverny, C., and Castelnaud, G. 2008. Fishing, abundance and life
- history traits of the anadromous Sea Lamprey (Petromyzon marinus) in Europe. Fish.

969 Res. 92:90–101.

970

- 971 Bence, J.R., Bergstedt, R.A., Christie, G.C., Cochran, P.A., Ebener, M.P., Koonce,
- 972 J.F., Rutter, M.A., and Swink, W.D. 2003. Sea lamprey (*Petromyzon marinus*)
- parasite-host interactions in the Great Lakes. J. Great Lakes Res. 29 (Suppl. 1): 253-

974 282.

975

- 976 Bergstedt, R. A., and Swink, W. D. 1995. Seasonal growth and duration of the
- 977 parasitic life stage of landlocked Sea Lampreys (*Petromyzon marinus*). Can. J. Fish.
- 978 Aquat. Sci. 52:1257–1264.

979

- 980 Bergstedt, R. A., and Twohey, M. B. 2007. Research to support sterile-male-release
- and genetic alteration techniques for sea lamprey control. J. Great Lakes Res.
- 982 33(Special Issue 2): 48-69.
- 983
- 984 Bill, B. R., Petzold, A. M., Clark, K. J., Schimmenti, L.A., and Ekker, S.C. 2009. A
- primer for morpholino use in zebrafish. Zebrafish 6: 69-77.

- 987 Burt, A. 2003. Site-specific selfish genes as tools for the control and genetic
- 988 engineering of natural populations. Proc Biol Soc Lond 270:921–928.
- 989
- 990 Champer, J., Buchman, A., and Akbari, O. S. 2016. Cheating evolution: engineering
- gene drives to manipulate the fate of wild populations. Nature Rev. 17: 146-159.
- 992
- 993 Chan, Y.-S., Naujoks, D. A., Huen, D. S., and Russell, S. 2011. Insect population
- 994 control by homing endonuclease-based gene drive: an evaluation in *Drosophila*
- 995 melanogaster. Genetics 188: 33-44.
- 996
- 997 Cochran, P. A., and Marks, J. E. 1995. Biology of the silver lamprey, *Ichthyomyzon*
- *unicuspis*, in Green Bay and the lower Fox River, with a comparsion to the Sea
- 999 Lamprey, Petromyzon marinus. Copeia 1995: 409-421.
- 1000 Committee on Gene Drive Research in Non-Human Organisms: Recommendations
- 1001 for Responsible Conduct. 2016. Gene Drives on the Horizon: Advancing Science,
- 1002 Navigating Uncertainty, and Aligning Research with Public Values. National
- 1003 Academies Press, Washington, D.C., 214 Pp.
- 1004 Conlon, J. M., Nielsen, P. F., and Youson, J. H. 1993. Primary structure of glucagon
- 1005 and glucagon-like peptide isolated from the intestine of the parasitic phase lamprey
- 1006 Petromyzon marinus. Gen. Comp. Endocrinol. 91: 96-104.
- 1007
- 1008 Covolo-Soto, L., Saura, M., and Moran, P. 2015. Does DNA methylation regulate
- 1009 metamorphosis? The case of the Sea Lamprey (*Petromyzon marinus*) as an example.
- 1010 Comp. Biochem. Physiol. B: Biochem Mol. Miol. 185: 42-46.

- 1012 Cowan, P. E. 1996. Possum biocontrol: prospects for fertility regulation. Reprod.
- 1013 Fertil. Dev. 8:655–660.
- 1014
- 1015 Dabrowski, K., Lee, K.-J., Froscauer, J., Abiado, M. A. G., Wolfe, T., and Ciereszko,
- 1016 A. 2004. Understanding variabilities in lamprey gametes quality in relation to
- 1017 availability of nutrients in host fish. GLFC 2004 Completion Rpt.
- 1018
- 1019 Dawson, H. A., and Jones, M.L. 2009. Factors affecting recruitment dynamics of
- 1020 Great Lakes sea lamprey (*Petromyzon marinus*) populations. J. Great Lakes Res. 35:
- 1021 353-360.
- 1022
- 1023 Dawson, H. A., Jones, M. L., Irwin, B. J., Johnson, N. S., Wagner, C. M., and
- 1024 Szymanski, M. 2016. Management strategy evaluation of pheromone-baited trapping
- 1025 techniques to improve management of invasive sea lamprey. Natural Resource
- 1026 Modeling 29: 448-469.
- 1027
- 1028 Dawson, H. A., Quintella, B. R., Almeida, P. R., Treble, A. J., and Jolley, J. C. 2015.
- 1029 The ecology of larval and metamorphosing lampreys. Pp. 75-136. In: Docker, M.F.
- 1030 (ed.). Lampreys: Biology, Conservation and Control. Vol. 1., Springer, N.Y.
- 1031
- 1032 De'ath, G., and Fabricius, K. E. 2000. Classification and regression trees: a powerful
- 1033 yet simple technique for ecological data analysis. Ecology 8: 3178 3192.
- 1034

1035	Deredec, A., Godfray, H. C., and Burt, A. 2011. Requirements for effective malaria
1036	control with homing endonuclease genes. Proc. Nat. Acad. Sci. (USA) 108, E874-
1037	E880.

1039 Deredec, A., Burt, A., and Godfray, H. C. J. 2008. The population genetics of using

1040 homing endonuclease genes in vector and pest management. Genetics 179: 2013-

1041 2026.

1042

- 1043 Docker, M. F. 2009. A review of the evolution of nonparasitism in lampreys and an
- 1044 update of the paired species concept. In: Brown, L.R., S.D. Chase, M.G. Mesa, R.J.

1045 Beamish and P.B. Moyle (eds). Biology, management and conservation of lampreys

1046 in North America. Amer. Fish. Soc. Symp. 72:71-114.

1047

- 1048 Docker, M. F., and Beamish, F. W. H. 1994. Age, growth, and sex ratio among
- 1049 populations of least brook lamprey, Lampreta aepyptera, larvae: an argument for

1050 environmental sex determination. Env. Biol. Fish. 41: 191-205.

1051

- 1052 Drake, D. A. R., Mercader, R., Dobson, T., and Mandrak, N. E. 2015. Can we predict
- 1053 risky human behaviour involving invasive speices? A case study of the release of
- 1054 fishes to the wild. Biological Invasions 17(1): 309-326.
- 1055
- 1056 Dubelman, S., Fischer, J., Zapata, F., Huizinga, K., Jiang, C., Uffman, J., Levine, S.,
- and D. Carson. 2014. Environmental Fate of Double-Stranded RNA in Agricultural
- 1058 Soils. PLoS ONE9(3): e93155. <u>https://doi.org/10.1371/journal.pone.0093155</u>.

1060	Dunlop, E.S. et al.	2017.	Rapid evolution meet	s invasive s	species control:	the
------	---------------------	-------	----------------------	--------------	------------------	-----

- 1061 potential for pesticide resistance in sea lamprey. Can. J. Fish. Aquat. Sci. 75: 152-
- 1062 168.
- 1063
- 1064 Erickson, R.A., Eager, E. E., Brey, M. K., Hansen, M. J., and Kocovsky, P. M. 2017.
- 1065 An integral projection model with YY-males and application to evaluating grass carp
- 1066 control. Ecol. Model. 361: 14-25.
- 1067
- 1068 Eshenroder, R. L. 2015. The role of the Champlain Canal and Erie Canal as putative
- 1069 corridors for colonization of Lake Champlain and Lake Ontario by Sea Lampreys.
- 1070 Trans. Amer. Fish. Soc. 143: 634-649.
- 1071
- 1072 Esvelt, K. M., Smidler, A. L., Catteruccia, F., and Church, G. M. 2014. Concerning
- 1073 RNA-guided gene drives for the alteration of wild populations. eLife
- 1074 10.7554/eLife.03401.
- 1075
- 1076 Esvelt, K. M. and Gemmell, N. J. 2017. Conservation demands safe gene drive.
- 1077 PLoS Biol. 15(11): e2003850. <u>https://doi.org/10.1371/journal.pbio.2003850</u>.
- 1078
- 1079 Fu, G., Condon, K. C., Epton, M. J., Gong, P., Jin, L., Condon, C. C., Morrison, N. I.,
- 1080 Dafa'alla, T. H., and Alphey, L. 2007. Female-specific insect lethality engineered
- 1081 using alternative splicing. Nature Biotechnol. 25: 353-357
- 1082
- 1083 Fu, G., Lees, R. S., Nimmo, D., Aw, D., Jin, L., Gray, P., Berendonk, T. U., White-
- 1084 Cooper, H., Scaife, S., Kim Phuc, H., Marinotti, O., Jasinakiene, N., James, A. A.,

- 1085 and Alphey, L. 2010. Female-specific flightless phenotype for mosquito control.
- 1086 Proc. Nat. Acad. Sci. (USA) 107: 4550-4554

- 1088 Galizi, R., Doyle, L. A., Menichelli, M., Bernardini, F., Deredec, A., Burt, A.,
- 1089 Stoddard, B. L., Windbichler, N., and A. Crisanti, A. 2014. A synthetic sex ratio
- 1090 distortion system for the control of the human malaria mosquito. Nature Comm.

1091 5:3977.

1092

- 1093 Gantz, V. M., and Bier, E. 2015. Genome editing. The mutagenic chain reaction: a
- 1094 method for converting heterozygous to homozygous mutations. Science 348, 442–
- 1095 444.

1096

- 1097 Gantz, V. M., Jasinskiene, N., Tatarenkova, O., Fazekas, A., Macias, V. M., Bier, E.,
- and A. A. James, A. A. 2015. Highly efficient Cas9-mediated gene drive for
- 1099 population modification of the malaria vector mosquito Anopheles stephensi. Proc.
- 1100 Nat. Acad. Sci. (USA) 112: 6736-6743.

1101

- 1102 Garforth, K., and Miranda, M.. 2014. The Cartagena Protocol on Biosafety and
- 1103 living modified fish. Biol. Inv. 16: 1313-1323.
- 1104
- 1105 Gimble, F. S., and Thorner, J. 1992. Homing of a DNA endonuclease gene by
- 1106 meiotic gene conversion in *Saccharomyces cerevisiae*. Nature 357: 301-306.

- 1108 Gould, F., and Schliekelman, P. 2004. Population genetics of autocidal control and
- 1109 strain replacement. Ann. Rev. Entomol. 49:193–217.

1111	Gutierrez, J. B., and Teem, J. L. 2006. A model describing the effect of sex-reversed
1112	YY fish in an established wild population: the use of a Trojan Y chromosome to cause
1113	extinction of an introduced exotic species. J. Theoret. Biol. 241:333-341.
1114	
1115	Hall, S. S. 2017. Could genetic engineering save the Galapagos. Sci. Amer.
1116	www.scientificamerican.com/article/could-genetic-engineering-save-the-gal-aacuote-
1117	pagos/
1118	
1119	Hamilton, W. D. 1967. Extraordinary sex ratios. Science 156: 477-488.
1120	
1121	Hammond, A., Galizi, R., Kyrou, K., Simoni, A., Siniscalchi, C., Katsanos, D.,
1122	Gribble, M., Baker, D., Marois E., Russell, S., Burt, A., Windbichler, N., Crisanti, A.,
1123	and Nolan, T. 2016. A CRISPR-Cas9 gene drive system targeting female
1124	reproduction in the malaria mosquito vector Anopheles gambiae. Nature Biotechnol.
1125	34, 78–83.
1126	
1127	Hammond, A. M., Kyrou, K., Bruttini, M., North, A., Galizi, R., Karlsson, X., Kranjc,
1128	N., Carpi, F. M., D'Aurizo, R., Crisanti, A., and Nolan, T. 2017. The creation and
1129	selection for mutations resistant to a gene drive over multiple generations in the
1130	malaria mosquito. PLoS Genetics 13(10): e1007039
1131	https://doi.org/10.1371/journal.pgen.1007039
1132	

- 1133 Hansen, M. J., Madenjian, C. P., Slade, J. W., Steeves, T. B., Almeida, P.R., and
- 1134 Quintella, B. R. 2016. Population ecology of the Sea Lamprey (*Petromyzon marinus*)

as an invasive species in the Laurentian Great Lakes and an imperilled species in

1136 Europe. Rev. Fish. Biol. Fish. 26: 509-535.

- 1138 Happel, A., Rinchard, J., and Czesny, S. 2016. Variability in Sea Lamprey fatty acid
- 1139 profiles indicates a range of host species utilization in Lake Michigan. J. Great Lakes
- 1140 Res. 43: 182-188.
- 1141
- 1142 Hardy, C. M., Hinds, L.A., Kerr, P. J., Lloyd, M. L., Redford, A. J., Shellam, G. R.,
- and Strive, T. 2006. Biological control of vertebrate pests using virally vectored
- 1144 immunocontraception. J. Reprod. Immunol. 71:102–111.
- 1145
- Harmon, A. 2016. Gene editing to alter whole species gets limited backing. N.Y.Times, June 8, 2016.
- 1148
- 1149 Harris, A. F., McKenny, A. R., Nimmo, D., et al. 2012. Successful suppression of a
- 1150 field mosquito population by sustained release of engineered male mosquitoes.
- 1151 Nature Biotechnol. 30: 828-830
- 1152
- 1153 Heath, G., Childs, D., Docker, M. F., McCauley, D. W., and Whyard, S. 2014. RNA
- 1154 interference technology to control pest sea lampreys A proof-of-concept. PLoS
- 1155 ONE 9(2): e88387. doi:10.1371/journal.pone.0088387
- 1156
- 1157 Heinrich, J. C., and Scott, M. J. 2000. A repressible female-specific lethal genetic
- 1158 system for making transgenic insect strains suitable for a sterile-release program.
- 1159 Proc. Nat. Acad. Sci. (USA) 97: 8229–8232

- 1161 Hoffmann, A.A., Montgomery, B. L., Popovici, J., Iturbe-Ormaetxe, I., Johnson, P.
- 1162 H., Muzzi, F., Greenfield, M., Durkan, M., Leong, Y.S., Dong, Y., Cook, H., Axford,
- 1163 J., Callahan, A. G., Kenny, N., Omodei, C., McGraw, E. A., Ryan, P. A., Ritchie, S.
- 1164 A., Turelli, M., and O'Neill, S. L. 2011. Successful establishment of Wolbachia in
- 1165 Aedes populations to suppress dengue transmission. Nature 476:454–457.

1166

- 1167 Ito, N., Mita, M., Takahashi, Y., Matsushima, A., Watanabe, Y. G., Hirano, S., and
- 1168 Odani, S. 2007. Novel cysteine-rich secretory protein in the buccal gland secretion
- 1169 of the parasitic lamprey, *Lethenteron japonicum*. Biochem. Biophys. Res. Comm.
- 1170 358: 35-40.
- 1171
- 1172 Iyengar, A., Muller, F., and Maclean, N. 1996. Regulation and expression of

1173 transgenes in fish – a review. Transgenic Res. 5: 147-166.

- 1174
- 1175 Jackson A. L., Bartz, S. R., Schelter, J., Kobayashi, S. V., Burchard, J., Mao, M., et
- al. 2003. Expression profiling reveals off-target gene regulation by RNAi. Nature
- 1177 Biotechnol. 21: 635–637.
- 1178
- 1179 Jaroszewska, M., Lee, B.-J., Dabrowski, K., Czesny, S., Rinchard, J., Trzeciak, P.,
- 1180 and Wilczynska, B. 2009. Effects of vitamin B<sub>1</sub> (Thiamine) deficiency in lake trout
- 1181 (*Savelinus namaycush*) alevins at the hatching stage. Comp. Biochem. Physiol. 154:
- 1182 255-262.
- 1183

- 1184 Johnson, B. M., Arlinghaus, R., and Martinez, P. J. 2009a. Are we doing all we can
- to stem the tide of illegal fish stocking? Fisheries 34: 389-394.
- 1186
- 1187 Johnson, N. S., Yun, S.-S., Thompson, H. T., Chio, J., and Li, W. 2009b. A
- 1188 synthesized pheromone induces an upstream movement in female sea lampreys and
- summons them into traps. Proc. Nat. Acad. Sci. (U.S.) 106:1101-1108.
- 1190
- 1191 Johnson, N. S., Siefkes, M. J., Wagner, C. M., Dawson, H. A., Wang, H., Steeves, T.
- 1192 B., Twohey, M., and Li, W. 2013. A synthesized mating pheromone component
- 1193 increases adult sea lamprey (Petromyzon marinus) trap capture in management
- 1194 scenarios. Can. J. Fish. Aquat. Sci. 70: 1101-1108.
- 1195
- 1196 Johnson, N. S., Swink, W. D., and Brenden, T. O. 2017. Field study suggests that
- 1197 sex determination in Sea Lamprey is directly influenced by growth rate. Proc. R. Soc.
- 1198 B 284: 20170262. http://dx.doi.org/10.1098/rspb.2017.0262.
- 1199
- 1200 Jones, M.L., Irwin, B.J., Hansen, G.J.A., Dawson, H. A., Treble, A. J., Liu, W., Dai,
- 1201 W., and Bence, J.R. 2009. An operating model for the integrated pest management of
- 1202 Great Lakes sea lampreys. Open Fish Sci. J. 2: 59-73.
- 1203
- 1204 Jorgenson, J. C., and Kitchell, J. F. 2005. Growth potential and host mortality of the
- 1205 parasitic phase of the Sea Lamprey (*Petromyzon marinus*) in Lake Superior. Can. J.
- 1206 Fish. Aquat. Sci. 62: 2343-2353.
- 1207

- 1208 Kamola P. J., Nakano, Y., Takahashi, T., Wilson, P. A., and Ui-Tei, K. 2015. The
- 1209 siRNA Non-seed Region and Its Target Sequences Are Auxiliary Determinants of
- 1210 Off-Target Effects. PLoS Comput. Biol. 11(12): e1004656.
- 1211 doi:10.1371/journal.pcbi.1004656
- 1212
- 1213 Kim, D. J., and Rossi., J. R. 2007. Strategies for silencing human disease using RNA
- 1214 interference. Nature Genetics 8: 173-184.

- 1216 Kim, Y. H., Issa, M. S., Cooper, A. M. W., and Zhu, K. Y. 2015. RNA interference:
- 1217 applications and advances in insect toxicology and insect pest management. Pesticide
- 1218 Biochem. Physiol. 120: 109-117.
- 1219
- 1220 Klein, T. A., Windbichler, N., Deredec, A., Burt, A., and Benedict, M. Q. 2012.
- 1221 Infertility resulting from transgenic I-Pol male *Anopheles gambiae* in large cage trials.
- 1222 Pathogens Global Health 106: 20-31.
- 1223
- 1224 Krafsur, E. S. 1998. Sterile insect technique for suppressing and eradicating insect
- 1225 population: 55 years and counting. J. Agricult. Ento. 15: 303–317
- 1226
- 1227 Kuraku, S., Qiu, H., and Meyer, A. 2012. Horizontal transfer of Tc1 elements
- 1228 between teleost fishes and their vertebrate parasites, lampreys. Genome Biol. Evol. 4:

1229 929-936.

- 1231 Kusakabe, R., Tochinai, S., and Kuratani, S. 2003. Expression of foreign genes in
- 1232 lamprey embryos: an approach to study evolutionary changes in gene regulation. J.
- 1233 Exp. Zool. B Mol. Dev. Evol. 296: 87-97.
- 1234
- 1235 Kwon, J. Y., Haghpanah, V., Kogson-Hurtado, L. M., McAndrew, B. J., and Penman,
- 1236 D. J. 2000. Masculinisation of genetic female Nile tilapia (Oreochromis niloticus) by
- 1237 dietary administration of an aromatase inhibitor during sexual differentiation. J. Exp.
- 1238 Zool. 287: 46–53.
- 1239
- 1240 Lacroix, R. et al. 2012. Open field release of genetically engineered sterile male
- 1241 Aedes aegypti in Malaysia. PLoS One 7(8): e42771.
- 1242 Doi:10.1371/journal.pone.0042771
- 1243
- 1244 Lavis, D. S., Hallett, A., Koon, E. M., and McAuley, T.C. 2003. Advances in barriers
- 1245 as an alternative method to suppress sea lampreys in the Great Lakes. J. Great Lakes
- 1246 Res. 29 (Supplement 1): 362-372.
- 1247
- 1248 Li, W., Scott, A. P., Siefkes, M. J., Yun S.-S., and Zielinski, B. 2003a. A male
- 1249 pheromone in the Sea Lamprey (*Petromyzon marinus*): an overview. Comp.
- 1250 Physiol. Biochem., 28, 259–252.
- 1251
- 1252 Li, W., Siefkes, M. J., Scott, A. P., and Teeter, J. H. 2003b. Sex pheromone
- 1253 communication in the Sea Lamprey: implications for integrated management.
- 1254 J. Great Lakes Res., 29(Suppl. 1). 85–94.

- 1256 Lundgren, J. G., and Duan, J. J. 2013. RNAi-based insecticidal crops: potential
- 1257 effects on nontarget species. Biosci. 63: 657-665.
- 1258
- 1259 Mateus, C. S., Stange, M., Berner, D., Roesti, M., Quinella, B. R., Alves, M. J.,
- 1260 Almeida, P. R., and Salzburger, W. 2013. Strong genome-wide divergence between
- 1261 sympatric European river and brook lampreys. Curr. Biol., 23(15): R650.
- 1262
- 1263 Mendoza, S. R., Angulo, C., and Meza, B. 2016. Food-grade organisms as vaccine
- 1264 biofactories and oral delivery. Trends Biotechnol. 34: 124-136.
- 1265
- 1266 Meng, X., Noyes, M. B., Zhu, L. J., Lawson, N. D., and Wolfe, S. A. 2008. Targeted
- 1267 gene inactivation in zebrafish using engineered zincfinger nucleases. Nature
- 1268 Biotechnol. 2008: 26:695–701.
- 1269
- 1270 Muir, W. M., and Howard, R. D. 1999. Possible ecological risks of transgenic
- 1271 organism release when transgenes affect mating success: sexual selection and the

1272 Trojan gene hypothesis. Proc. Nat. Acad. Sci. (USA) 96:13853–13856.

- 1273
- 1274 Muir, W. M., and Howard, R. D. 2002. Assessment of possible ecological risks and
- 1275 hazards of transgenic fish with implications for other sexually reproducing organisms.
- 1276 Transgenic Res. 11:101–114.
- 1277
- 1278 Muir, W. M., and Howard, R. D. 2004. Characterization of environmental risk of
- 1279 genetically engineered (GE) organisms and their potential to control exotic invasive
- 1280 species. Aquat. Sci. 66:414–420.

- 1282 Najarian, M. 2015. Are fish falling from the sky in Fairbanks.
- 1283 http://edition.cnn.com/2015/06/05/us/fairbanks-falling-fish/index.htmlMyCNN
- 1284
- 1285 Noble, C., Min, J., Olejarz, J., Buchthal, J., Chavez, A., Smidler, A. L., DeBendictis,
- 1286 E.A., Church, G.M., Nowak, M.A. and Esvelt, K.M. 2016. Daisy-chain gene drives
- 1287 for the alteration of local populations. bioRxiv. 2016; 057307.
- 1288
- 1289 Otts, S. S. 2014. U.S. regulatory framework for genetic biocontrol of invasive fish.
- 1290 Biol. Inv. 16: 1289-1298.
- 1291
- 1292 Owens, B. 2017. Can New Zealand pull off an audacious plan to get rid of invasive1293 predators by 2050? Nature 541:148-150.
- 1294
- 1295 Oye, K. A., Esvelt, K., Appleton, E., Catteruccia, F., Church, G., Kuiken, T.,
- 1296 Lightfoot, S. B.-Y., McNamara, J., Smidler, A., and Collins, J. P. 2014. Regulating
- 1297 gene drives. Science 345: 626-628.
- 1298
- 1299 Parker, H. J., Sauka-Spengler, T., Bonner, M., and Elgar, G. 2014. A reporter assay
- 1300 in lamprey embryos reveals both functional conservation and elaboration of vertebrate
- 1301 enhancers. PLoS One 9(1): e85492. doi:10.1371/journal.pone.0085492
- 1302
- 1303 Peanparkdee, M., Iwamoto, S., and Yamauchi, R. 2016. Microencapsulation: a
- 1304 review of applications in the food and pharmaceutical industries. Rev. Agric. Sci. 4:
- 1305 56-65.

- Phuc, H. K. et al. 2007. Late-acting dominant lethal genetic systems and mosquitocontrol. BMC Biology 5: 11
- 1309
- 1310 Piferrer, F., Zanuy, S., Carrillo, M., Solar, I. I., Devlin, R. H., and Donaldson, E. M.
- 1311 1994. Brief treatment with an aromatase inhibitor during sex differentiation causes
- chromosomally female salmon to develop as normal, functional males. J. Exp. Zool.270: 255–262.
- 1314
- 1315 Powers, D. A., Hereford, L., Cole, T., Chen, T. T., Lin, C. M., Kight, K., Creech, K.,
- 1316 and Dunham, R. 1992. Electroporation: a method for transferring genes into the
- 1317 gametes of zebrafish (Brachydanio rerio), channel catfish (Ictalurus punctatus) and
- 1318 common carp (*Cyprinus carpio*). Mol. Mar. Biol. Biotechnol. 1: 301-308.
- 1319
- 1320 Prowse, T. A. A., Cassey, P., Ross, J. V., Pfitzner, C., Wittmann, T. A., and Thomas,
- 1321 P. 2017. Dodging silver bullets: good CRISPR gene-drive design is critical for
- 1322 eradicating exotic vertebrates. Proc. Royal Soc. B. 284: 20170799.
- 1323 http://dx.doi.org/10.1098/rspb.2017.0799
- 1324
- 1325 Purvis, H.A. 1979. Variations in growth, age at transformation, and sex ratio of sea
- 1326 lampreys reestablished in chemically treated tributaries of the Upper Great Lakes.
- 1327 Great Lakes Fishery Commission, Technical Report 35. Ann Arbor, MI.
- 1328
- 1329 Ripley, B. 2016. tree: Classification and Regression Trees. R package version 1.0-37.
- 1330 https://CRAN.R-project.org/package=tree.

1332	Rodreiguez-Munoz, R., Waldman, J. R., Grunwald, C., Roy, N. K., and Wirgin, I.
1333	2004. Absence of shared mitochondrial DNA haplotypes between Sea Lamprey from
1334	North American and Spanish rivers. J. Fish Biol. 64: 783-787.
1335	
1336	Romasek, M., Square, T., Jandzik, D., and Medeiros, D. M. 2015. CRISPR/Cas
1337	system in the Sea Lamprey: a tool for understanding ancestral gene functions in
1338	vertebrates. Abstract P2-156, 2015 Annual meeting of the Society for Integrative and
1339	Comparative Biology.
1340	
1341	Saey, T. H. 2015. Gene drives unleashed. Science News, Dec. 2015: 16-22.
1342	
1343	Schliekelman, P., Ellner, S., and Gould, F. 2005. Pest control by genetic manipulation
1344	of sex ratio. J. Econ. Entomol. 98:18-34.
1345	
1346	Sekido, R., and Lovell-Badge, R. 2009. Sex determination and the SRY: down to a
1347	wink and a nudge. Trends Genetics 25: 19-29.
1348	
1349	Seternes, T., Tonheim, T. C., Myhr, A. I., and Dalmo, R. A. 2016. A plant 35S
1350	CaMV promoter induces long-term expression of luciferase in Atlantic salmon.
1351	Nature Sci. Rpts. 6:25096   DOI: 10.1038/srep25096.
1352	
1353	Siefkes, M. J., Bergstedt, R. A., Twohey, M. B., and Li, W. 2003. Chemosterilization
1354	of male Sea Lampreys Petromyzon marinus does not affect sex pheromone release.
1355	Can. J. Fish. Aquat. Sci. 60:23–31.

1357	Silver, M. R., Kawauchi, H., Nozaki, M., and Sower, S. A. 2004. Cloning and
1358	analysis of the GnRH-III cDNA from eight species of lamprey representing the three
1359	families of Petromyzontiformes. Gen. Comp. Endocronol. 139: 85-94.
1360	
1361	Smith, J. J. et al. 2013. Sequencing of the Sea Lamprey (Petromyzon marinus)
1362	genome provides insights into vertebrate evolution. Nature Genetics 45: 415-421.
1363	
1364	Soroldoni, D., Hogan, B. M., and Oates, A. C. 2009. Simple and efficient
1365	transgenesis with meganuclease constructs in zebrafish. Pp. 117-130. In: Lieschke,
1366	G.J. et al. (eds). Zebrafish, Methods in Molecular Biology 546.
1367	
1368	Sower, S. 2003. The endocrinology of reproduction in lampreys and applications for
1369	male lamprey sterilization. J. Great Lakes Res. 29: 50-65.
1370	
1371	Sower, S. 2015. The reproductive hypothalamic-pituitary axis in lampreys. Pp. 305-
1372	374. In: Docker, M. F. (ed.). Lampreys: Biology, Conservation and Control. Vol. 1.,
1373	Springer, N.Y.
1374	

- 1375 Square, T., Romasek, M., Jandzek, D., Cattell, M. V., Klykowsky, M., and Medeiros,
- 1376 D. M. 2015. CRISPR/Cas9-mediated mutagenesis in the Sea Lamprey Petromyzon
- 1377 *marinus*: a powerful tool for understanding ancestral gene functions in vertebrates.
- 1378 Development 142: 4180-4187.
- 1379

- 1380 Stuart, G. W., McMurray, J. V., and Westerfield, M. 1988. Replication, integration
- 1381 and stable germ-line transmission of foreign sequences injected into early zebrafish

1382 embryos. Development 103: 403 – 412.

1383

- 1384 Suzuki, K., Gamble, R. L., and Sower, S. A. 2000. Multiple transcripts encoding
- 1385 lamprey gonadotropin releasing hormone-I precursors. J. Mol. Endocrinol. 24: 365-

1386 376.

1387

- 1388 Szyjka, S. J., Mandal, S., Schoepp, N. G., Tyler, B. M., Yohn, C. B., Poon, Y. S.,
- 1389 Villareal, S., Burkart, M. D., and Shurin, J. S. 2017. Evaluation of phenotype
- 1390 stability and ecological risk of a genetically engineered alga in open pond production.

1391 Algal Res. 24: 378-386.

1392

- 1393 Szymczak, K. 2016. To halt the spread of Lyme, Nantucket residents consider
- 1394 genetically engineered mice. www.statnews.com/2016/06/07/nantucket-lyme-

1395 genetic-engineering/

1396

- 1397 Thomas, D. D., Donnelly, C. A., Wood, R. J., and Alphey, L. S. 2000. Insect
- population control using a dominant, repressible, lethal genetic system. Science287:2474–2476.

1400

1401 Thresher, R. E. 2008. Autocidal technology for the control of invasive fish. Fisheries1402 33:114–121.

1405 towards the use of recombinant technology to manage the impact of an invasive

- 1408 Thresher, R. E., Hayes, K., Bax, N. J., Teem, J., Benfey, T. J., and Gould, F. 2014a.
- 1409 Genetic control of invasive fish: technological options and its role in integrated pest
- 1410 management. Biol. Inv. 16:1201-1216.
- 1411
- 1412 Thresher, R. E., Van de Kamp, J., Campbell, G., Grewe, P., Canning, M., Barney, M.,
- 1413 Bax, N. J., Dunham, R., Su, B., and Fulton, W. 2014b. Sex-ratio-biasing constructs
- 1414 for the control of invasive lower vertebrates. Nature Biotechnol. 32:424-427.
- 1415
- 1416 Twohey, M. B., Heinrich, J. W., Seelye, J. G., et al. 2003. The sterile-male-release
- 1417 technique in Sea Lamprey management. J. Great Lakes Res. 29 (Suppl. 1):410–423.1418
- 1419 Webber, B. L., Raghu, S., and Owains, O. R. 2015. Opinion: is CRISPR-based gene
- 1420 drive a biocontrol silver bullet or global conservation threat. Proc. Nat. Acad. Sci.
- 1421 (USA) 112:10565-10567.
- 1422
- 1423 Windbichler, N., Papathanos, P. A., Catteruccia, F., Ranson, H., Burt, A., and
- 1424 Crisanti, A. 2007. Homing endonuclease mediated gene targeting in Anopheles
- 1425 gambiae cells. Nucleic Acid Res. 35:5922-5933.
- 1426
- 1427 Windbichler, N., Menichelli, M., Papathanos, P. A., Thyme, S. B., Li, H., Ulge, U. Y.,

<sup>1406</sup> species: Sea Lamprey in the North American Great Lakes. Biol. Inv.

1428	Hovde, E	3. T.,	Baker,	D.,	Monnat	Jr,	R. J	J.,	Burt, A.	, and	Crisanti, A	Α.	2011.	А
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- 1429 synthetic homing endonuclease-based gene drive system in the human malaria
- 1430 mosquito. Nature 473:212-217.
- 1431
- 1432 Wright, J., Krueger, C. C., Brussard, P. F., and Hall, M. C. 1985. Sea Lamprey
- 1433 (Petromyzon marinus) populations in Northeastern North America: genetic
- 1434 differentiation and affinities. Can. J. Fish. Aquat. Sci. 42:776-784.

- 1436 Yufera, M., Pascual, E., and Fernadez-Diaz, C. 1999. A highly efficient
- 1437 microencapsulated food for rearing early larvae of marine fish. Aquacult. 177:249-
- 1438 256.
- 1439
- 1440 Zelinin, A. V., Alimov, A. A., Barmintzev, V. A., Beniumov, A. O., Zelenina, I. A.,
- 1441 Krasnov, A. M., and Kolesnikov, V. A. 1991. The delivery of foreign genes into
- 1442 fertilized fish eggs using high-velocity microprojectiles. Federation European
- 1443 Biochem. Soc. Letters 287(1-2):118-120.
- 1444
- 1445 Zhang, H.-H., Feschotte, C., Han, M.-J., and Zhang, Z. 2014. Recurrent horizontal
- 1446 transfers of *Chapaev* transposons in diverse invertebrate and vertebrate animals.
- 1447 Genome Biol. Evol. 6:1375-1386.
- 1448
- 1449 Zhuikov, M. 2016. What happens when a lake trout survives a Sea Lamprey attack?
- 1450 Univ. Wisconsin Sea Grant;
- 1451 seagrant.wisc.edu/Home/AboutUsSection/PressRoom/Details.aspx?PostID=2460

- 1453 Zu, Y., Zhang, X., Ren, J., Dong, X., Zhu, Z., Jia, L., Zhang, Q. and Li, W. 2016.
- 1454 Biallelic editing of a lamprey genome using the CRISPR/Cas9 system. Nature Sci.
- 1455 Rpts. 6:23496 | DOI:

Table 1. Genetic approaches and phenotypic objectives considered as potential geneticbiocontrol options for use against Sea Lamprey in the Great Lakes.

Genetic approach/delivery method	Desired phenotypic effect/objective
Genetically modified pathogen or parasite	Increase mortality
Diet-delivered products	Reduce fertility
Chromosomal modification	Bias sex ratio
Mendelian-inherited constructs	Induce mortality
Super Mendelian (Gene driven)-inherited	Cause non-parasitic maturation
constructs	
	Increase mating competitiveness while
	reducing fertility
	Increase or renew sensitivity to existing
	biocides
	Replace existing biocides

Table 2. Results of 200-year simulations of the effect of 5 genetic control tactics on a Great Lake Sea Lamprey population and a downstream ("other") population that receives emigrants from the Great Lake population at a rate of 0.01% of the latter per year. We did not model the non-parasitic Sea Lamprey option (see text). Each control scenario is summarized below. See the Supplemental Material for modelling details.

Scenario	Stocking regime	Population at year 50	Population at year 200	Final proportion of GM	Final proportion of GM
		relative to initial	relative to initial	individuals in GL	individuals in other
		population	population	population	population
Heritable sex ratio, DD	100,000 larvae each year for 10 years	0.68	0.03	0.23	<0.001
Heritable sex ratio, not DD	100,000 larvae each year for 10 years	0.81	0.78	0.02	< 0.001
Trojan gene, equal mating	1,000,000 larvae each year for 50 years	0.02	0.03	0	0
Trojan gene, 3X mating	200,000 larvae each year for 50 years	0.02	0.03	0	0
GM larval biocide, +20%	NA	< 0.001	0	0	0
Vaccinated prey, 4%	200,000 hosts spanning 1 - 10 years	0.08	0.001	0	0
Gene-drive sex ratio, 1 yr only	100 larvae for 1 year	0.95	0	1	0.08
Gene-drive sex ratio, 10 years	100,000 larvae each year for 10 years	0.02	0	1	0.81

Explanation of scenarios:

- 1. <u>Heritable (Mendelian) sex ratio drive, DD</u>. Sex ratio density dependent with 65% females at low adult densities.
- 2. Heritable (Mendelian) sex ratio drive, Not DD. Sex ratio not density dependent with 50% females.
- 3. <u>Trojan gene, equal mating</u>. Sterile adult GM males are equally competitive with wild type males for mates.
- 4. <u>Trojan gene, 3X mating</u>. Sterile adult GM males are 3 times as successful as wild type males for mates.
- 5. <u>GM larval biocide, + 20%</u>. Simulated 20% increase in larval mortality due to biocide release
- 6. <u>Vaccinated prey, 4%</u>. Based on a host population size of 5,000,000, a 4% host vaccination rate achieved by stocking a total of 200,000 GM hosts over a several year period, 100% sterility or death resulting from a single attack of a vaccinated host, and an average of 10 hosts attacked per parasite. The model is based on the consequent 34% reduction in parasitic Sea Lamprey survival.

- 7. <u>Gene-driven sex ratio distortion, 1 year only</u>. Assumes 100% homing and no counter-selection.
- 8. <u>Gene-driven sex ratio distortion, 10 years</u>. As above, but higher and longer stocking effort

Table 3. Mean (and range among panellists) of the risk scores of adverse outcomes as assessed by a panel of experts for six "focal" genetic control options against five risk categories, and overall risks. Note that the risks for artificially sourced and self-propagating synthetic biocides were assessed separately. Risk estimates were scored by panellists as 1 through 7, where 1 = probability of occurrence during the control program < 1 in 1,000,000; 2 = < 1 in 10,000; 3 = < 1 in 100; 4 = 1-10%; 5 = 10-50%; 6 = 50-95% and 7 = >95% probability of occurrence. Technical and logistical feasibility was scored qualitatively on a scale of 1 to 7.

Option	Native	Other SL	Other	Human	Other	Overall	Feasibility
	lampreys	populations	organisms	health	risks	risk	
Mendelian	1 95 (1-4)	3 09 (2-7)	1 59 (1-3)	1 41 (1-2)	1.33 (1-	3.00 (1-	4 96 (4-6)
sex ratio	100 (1 1)	0.09 (27)	1.57 (1.5)	1 (1 2)	3)	4)	1.50(1.0)
Troian males	1 45 (1-2)	1 55 (1-2)	1.09 (1-2)	1 14 (1-2)	1.13 (1-	1.55 (1-	3 45 (3-4)
i rojun maios	1110 (1 2)	1.55 (1 2)	1.05 (1 2)	1.11 (1 2)	2)	2)	
Non-	2 82 (2 4)	2(12)	1 2 (1 2)	1()	1()	2.36 (2-	2 85 (2 4)
parasitic SL	2.02 (2-4)	2 (1-3)	1.2 (1-2)	1 (-)	1 (-)	3)	2.03 (2-4)
Vaccinated	2.36(1.4)	1 14 (1 2)	18(13)	1 55 (1 2)	1.29 (1-	2.45 (1-	1 80 (2 6)
Prey	2.50 (1-4)	1.14 (1-2)	1.0 (1-5)	1.55 (1-2)	4)	4)	4.00 (2-0)
Gene driven	2.45(1.5)	1 32 (2 7)	1 77 (1 4)	1 45 (1 3)	1.67 (1-	4.11 (2-	1 11 (2 6)
sex ratio	2.45 (1-5)	4.32 (2-7)	1.77 (1-4)	1.45 (1-5)	5)	6)	4.44 (2-0)
Synthetic							
larval	2.23 (1-4)	1.18 (1-2)	1.59 (1-3)	1.27 (1-2)	1 (-)	1.8 (1-3)	4.41 (2-6)
biocide							
Self-							
propagating					1.17 (1-		
larval	2.91 (1-7)	3.55 (1-7)	2.23 (1-7)	1.32 (1-2)	2)	3.8 (2-7)	3.13 (1-4)
biocide							
olociae							

Table 4. Key over-arching risk elements based on the risk assessment workshop and consultations with experts in the field of genetic technology and Sea Lamprey biology. *including rogue human activities (e.g., intentional movement or illegal live sale) facilitate breaking through that gap when measured over decadale scales.* 

#### **Off-target effects of constructs**

Depends critically on effecting molecule. RNAi likely key enabling technology. Cross-reactivity of siRNA with humans or other species potentially high as although nominally it requires 21 bp overlap, it can occur with as little as 6 bp overlap in nucleotide 2-7 position. Hence probability of siRNA match is 1/(4^6) not 1/(4^21); small, but not minute. However, impacts are likely to be dose dependent, so normal, e.g. dietary uptake, would be single or few molecules and hence exceedingly low dose. RNAi is a relatively mature science and methods for sequence design and minimization of off-target effects are well developed and could be applied to SL

#### **Escapement (physical containment) from Great Lakes**

Central issues are whether there is probable movement of SL down system into the Atlantic and if they get there, will they successfully breed with Atlantic SL. Downstream movement over falls highly likely to be successful, while canals could provide alternative pathways, along with ship/boat traffic and hitching rides on parasitized migrating fish. There is a distributional gap of SL in the St Lawrence seaway, but anthropogenic vectors, including rogue human activities (e.g., intentional movement or illegal live sale) facilitate breaking through that gap when measured over decadal scales. facilitate breaking through that gap. There are no obvious genetic, osmoregulatory or developmental barriers to interbreeding, particularly if downstream transport is slow (2 generations suggested) and involves larvae.

### **Vertical Gene Transfer**

<u>Native North American lampreys</u> – No evidence of hybridization between SL and native species. Probability exceedingly low.

<u>Atlantic SL population</u> – No obvious behavioral or other barrier to interbreeding

## Horizontal Gene Transfer (HGT)

- *HGT operates, but on the scales of 10,000 to millions of years. On the time scales we are considering (decades) likelihood of HGT exceedingly low.*
- Transposable and other mobile genetic elements have a higher probability of HGT than less mobile elements, so construct design should avoid e.g., transposon mediated integration
- Despite publications, HGT among vertebrates (or animal metazoans) is still highly debated, and may prove more or less frequent than suggested

### **Risk of re-infestation of the Great Lakes**

- A possible problem for any non-heritable (= "permanent") control option
- *Re-invaders need to be treated logically as any other potential invasive species*
- Prevention of re-infestation and development and maintenance of preventative measures could be a problem due to the diversity of vectors, human and otherwise
- Near or complete eradication of GL SL could lead to a decline in the capacity to mobilize a rapid response containment/eradication program should re-infestation be detected. There needs to be a recognition of the risk, a game plan on how to deal with it, and an on-going capacity for early detection of a re-inoculation event.



Figure 1. Simulated changes in relative abundance of Sea Lamprey as a result of genetic control options: (A) heritable sex ratio drive option with intrinsic sex ratio assumed to be density dependent; (B) heritable sex ratio drive option with intrinsic sex ratio assumed to be fixed at 50% females; (C) Trojan gene option with GM males assumed to be sterile and equal in competitiveness with wild type males for matings, at stocking rates ranging from 250,000 to 1,000,000 males per year; and (D) as in Fig. C, but GM males 3X as effective as non-GM males at attracting mates, and at stocking rates of 50,000 to 200,000 males per year. Note that for the heritable sex ratio drive (A and B), we simulated three duration periods for the stocking of genetically modified larvae after which no additional stocking occurred; for C and D we simulated three different levels of stocking of larvae, which continued for the entire length of the simulation. See Supplemental Material for details of the simulation model.



Figure 2. Simulated changes in relative abundance of Sea Lamprey as a result of genetic control options: (A) addition of a genetically-based larval biocide that reduces larval survival, on average, by 5%, 10%, or 20%; (B) inoculation of 1%, 2%, or 4% of Sea Lamprey hosts with a gene-based vaccine that causes mortality of Sea Lampreys that attack the inoculated hosts: (C and D) introduction of larvae containing a non-Mendelian gene drive using three scenarios: 100 GM larvae for 1 year; 100 GM larvae for 10 years, and 100,000 GM larvae for 10 years. The left panel (C) shows the abundance of Sea Lamprey in the Great lakes as a result of the interventions; the right panel (D) shows changes in the prevalence of the genetically modified larvae in a non-target Sea Lamprey population connected to the Great Lakes population by a 0.01% per year emigration rate. See the Supplemental Material for details of the simulation model.



Figure 3. Level of risk as assessed by an expert panel for each identified risk category averaged (mean) across six focal options and eleven panel members.



Level of support (1 = strongly oppose, 7 = strongly support)

Figure 4. Distribution of level of support or opposition to focal options by the fishing community and stakeholders. Numbers are the percent of respondents that supported (score  $\geq 5$ ) R&D or Implementation for each option. Dashed lines indicate neutral response. The self-propagating and artificially dispersed nursery area synthetic biocides have been pooled for this analysis.


Figure 5. Representative pruned regression trees for a widely and a weakly supported option (heritable [Mendelian] sex ratio drive and a gene drive, respectively) among stakeholders and fishers, pooled, identifying factors predictive of support/opposition for R&D within the 'unconvinced fraction' of respondents. Numeric values are mean levels of support (where Strongly opposed = 1, Opposed = 2, Moderately opposed = 3, Neutral = 4, Moderately supportive = 5, Supportive = 6) and node labels indicate survey responses. The model explained 42% and 32% of the variance in the level of support for R&D, respectively.

Regression tree analyses for Implementation of the two options are essentially identical, though capturing less variance (33% and 24%, respectively).



Figure 6. Scatterplot of median feasibility versus median highest risk scores for each of the seven focal options as assessed by an expert panel. Diameter of each point indicates the amount of survey respondent support for R&D for each option. Support levels were similar for both surveyed groups except for the vaccinated prey option, which was supported by 50% of engaged stakeholders, but 30.6% of the fishing community.

## Thresher, Jones and Drake

## Evaluating active genetic options for the control of Sea Lamprey (Petromyzon marinus) in the Laurentian

Great Lakes

Supplementary Material

## Sea Lamprey population model

<u>Model description</u> – Effects of five of the six genetic biocontrol options considered in this paper were examined using an age-structured model of Sea Lamprey population dynamics. The non-parasitic Sea Lamprey option was not modeled. Here we provide a detailed description of the model originally developed to examine the first option – the heritable (Mendelian) sex ratio drive. We refer to this option below as "daughterless" to reflect the idea that all offspring carrying the construct will be phenotypically male. Following this we explain how the model was adapted to simulate the other four options.

The model included two populations: a single population in the Great Lakes presumed to be under active control using lampricides and barriers, as per current practice; and, a second, uncontrolled, downstream population. The model simulated the entire life cycle of both populations, and the populations were assumed to be connected by a small amount (nominal value: 1% per year) of unidirectional movement from the Great Lakes population to the downstream population. Although bidirectional movement is plausible, our focus was on risks associated with emigration of genetically modified Sea Lamprey from the target Great Lakes population to the non-target downstream population. For simplicity and because little is known of the population dynamics of Sea Lamprey outside of the Great Lakes, we assumed that the two populations had identical demographics.

We used a stage/age structured approach to model both populations of Sea Lamprey, with larvae ages 0-6, transformers, parasites, and adults. Within each stage/age there were males and females. The males were also subdivided by the number of copies of the construct (i.e., the number of loci where the effect is expressed) they had. We allowed for up to eight copies of the construct to be present in a genetically modified male. We assumed that any Sea Lamprey containing at least one copy of the construct would be a phenotypic male; therefore, by definition all females had a copy number of zero and all offspring with a copy number > 0 were classed as males. To simulate the effect of strategic distribution

of genetically modified larvae to enhance their survival, we also needed to separately track these individuals.

The number of Sea Lamprey larvae of sex k, copy number g, age a, in year t was given by:

$$N_{t+1,a+1}^{k,g} = N_{t,a}^{k,g} \cdot s_N \cdot (1 - pm_a) \cdot (1 - m_{cl})$$

$$G_{t+1,a+1}^{k,g} = G_{t,a}^{k,g} \cdot s_G \cdot (1 - pm_a) \cdot (1 - m_{cG})$$
(S1)

All model parameters and state variables are defined and assumed value listed in Table S1. The number of transformers (recently metamorphosed larvae), parasites, and adults of sex k, copy number g, in year t were calculated from:

$$T_{t+1}^{k,g} = \sum_{a=2}^{6} N_{t,a}^{k,g} \cdot pm_a \cdot (1 - m_{cT})$$

$$GT_{t+1}^{k,g} = \sum_{a=2}^{6} G_{t,a}^{k,g} \cdot pm_a \cdot (1 - m_{cGT})$$
(S2)

$$P_{t+1}^{k,g} = (T_t^{k,g} + GT_t^{k,g}) \cdot s_T$$
(S3)

$$A_t^{k,g} = P_t^{k,g} \cdot s_P \tag{S4}$$

We modeled reproduction using a stochastic Ricker stock-recruitment function with log-normal process errors on recruitment:

$$N_{t,0}^* = \alpha \cdot A_t^{1,0} \cdot e^{-\beta \cdot A_t^{1,0} + \varepsilon_t}$$
(S5)

Dawson and Jones (2009) examined recruitment dynamics at the spatial scale of individual streams, whereas the spatial scale of our model was an entire lake. To re-scale the parameters of the Ricker model to an entire lake, we used forecasted lake-scale recruitment dynamics aggregated from a more detailed model of Sea Lamprey population dynamics (Jones et al 2009) for which recruitment operated at the scale of individual spawning streams and that used Ricker parameter estimates from Dawson and Jones (2009). We fit these forecasted dynamics to a Ricker model to estimate the structural parameters and the variance term (Table S1).

To allocate age zero larvae to sexes and copy numbers we needed to assign copy numbers to gametes. For the daughterless construct, all individuals with at least one copy of the construct are effectively males, so female gametes will necessarily have a copy number of 0, and the proportion of female gametes with copy number 0 ( $p^{1,0}$ ) equals 1. To calculate the overall proportion of male gametes

with copy numbers from 0 to 8 we used the binomial probability distribution function. We compute the probability of a male gamete with h copies of the construct deriving from a male adult lamprey with g copies and then sum that proportion, weighted by the relative abundance of male adult lamprey with g copies, over all cases where  $g \ge h$ :

$$p^{0,h} = \sum_{g=h}^{8} \left[ \frac{g!}{h!(g-h)!} \cdot 0.5^{g} \cdot 0.5^{(g-h)} \cdot \frac{A_{t}^{0,g}}{\sum_{g=0}^{8} A_{t}^{0,g}} \right]$$
(S6)

This calculation assumes that the loci containing the construct are not linked (i.e., they disassociate independently during meiosis) and that the construct is never homozygous at a locus. Finally, by assuming random assortative mating, and noting that all female gametes have copy number 0, the proportion of offspring with copy number g will be

$$O^{g} = p^{0,g} \cdot p^{1,0} = p^{0,g}$$
(S7)

The number of age 0 larvae by sex and copy number is then:

$$N_{t,0}^{1,g} = N_{t,0}^{*} \cdot O^{0} \cdot \pi_{1} \qquad g = 0$$

$$N_{t,0}^{1,g} = 0 \qquad g > 0$$

$$N_{t,0}^{0,g} = N_{t,0}^{*} \cdot O^{0} \cdot \pi_{0} \qquad g = 0$$

$$N_{t,0}^{0,g} = N_{t,0}^{*} \cdot O^{g} \qquad g > 0$$
(S8)

where  $\pi_1, \pi_0$  are the expected proportions of females and males with copy number 0 in the population  $(\pi_1 + \pi_0 = 1)$ .

The sex ratios of adult Sea Lamprey in the Great Lakes changed substantially from the precontrol (1950s) to the post-control (1980s) period (see Jones et al. 2003), with a preponderance of males prior to control and a preponderance of females more recently. We assumed that this change in sex ratio has been a compensatory response to changes in Sea Lamprey densities, and model the effect by allowing the proportion of males to decrease linearly from 65% at an adult abundance of 1,500,000 to 35% at an adult abundance of 150,000. This approximates pre- and post-control abundances in one of the upper Great Lakes. Below 150,000 and above 1,500,000 the proportions were fixed at 35% and 65% respectively. We also simulated a scenario where the proportion of males was not assumed to be density dependent but remained at 0.5 regardless of population size. We calibrated the model to produce forecasts of Sea Lamprey abundance, *A*, in the Great Lakes population consistent with levels observed recently in the upper Great Lakes, in the absence of any genetic biocontrol option being simulated. The downstream population was initialized at roughly four times the upper Great Lakes adult abundance (Table S1). Calibration was accomplished by adjusting larval survival rates.

Implementation of Options – The heritable (Mendelian) sex ratio drive option was simulated by introducing a fixed number of genetically modified larvae into the Great Lake population over a specified period of years. We assumed that when genetically modified larvae were introduced, they would be stocked in streams not subject to lampricide control ( $m_{cG}$ ,  $m_{cGT} = 0$ ). The model was designed to allow copy numbers for the construct from 1 to 8 - for this analysis we only simulated scenarios with a copy number of 1. The Trojan gene option was also simulated by introducing a fixed number of genetically modified larvae over a specified period of years, but in this case surviving individuals with the construct would develop into sterile adult males. Sterile males would compete with wild, fertile males for mates, reducing the number of effective female spawners just as would be the case for classic sterile male release tactics (e.g., Twohey et al. 2003). We also simulated a scenario where the genetically modified males were 3x as effective as wild males at successfully mating, resulting in a substantially greater effect on reducing the number of effective female spawners. The non-Mendelian gene drive sex ratio distortion option was modeled similarly to the Mendelian option except that all offspring of individuals carrying the construct were assumed to carry the construct as well. We included a parameter for "leakage" of the construct (i.e., loss of the construct in some fraction of offspring) but assumed leakage was zero for the scenarios presented in this paper.

The vaccinated prey option was modeled by reducing the survival of parasitic Sea Lamprey. We modeled three levels of inoculation for host fishes: 1%, 2%, and 4%, and assumed that any Sea Lamprey attacking an inoculated host would die. Assuming that Sea Lamprey attack, on average, ten hosts during their parasitic life stage, this would translate into 8.5%, 17%, and 34% reductions in parasitic survival, respectively. The GM larval biocide option was simulated by decreasing average larval survival rates. We examined three levels of reduced survival: 5%, 10%, 20%.

Parameter symbol	Definition	Value(s)
k	Sex index	0 – male, 1 - female

Table S1. Model parameters and their assumed values.

g	Construct copy number	0-8
t	Year of simulation	1-200
a	Age	0-6
N	Numbers of wild larvae	Dynamic state variable
G	Numbers of stocked larvae	Scenario specific
S <sub>N</sub> , S <sub>G</sub>	Annual larval survival (wild = N,	wild: 0.395; stocked: 0.395
	stocked = G)	
$pm_a$	Age-specific proportion of larvae	age 0-6: 0,0,0,0.2,0.5,0.75, 1
	entering metamorphosis	
$m_{cl}, m_{cG}$	Larval mortality due to chemical control	wild: 0.2; stocked: 0
	(wild = l, stocked = G)	
$m_{cT}, m_{cGT}$	Transformer mortality due to chemical	wild: 0.46; stocked: 0
	control (wild = T, stocked = GT)	
Т	Numbers of wild transformers	Dynamic state variable
GT	Numbers of stocked transformers	Dynamic state variable
Р	Number of parasitic Sea Lamprey	Dynamic state variable
A	Number of adult Sea Lamprey	Dynamic state variable: initialized
		at 110,000 adults for Great Lake
		population; 450,000 adults for
		downstream population
S <sub>T</sub>	Annual transformer survival	0.75
SP	Annual parasitic survival	0.75
α,β	Ricker structural parameters	469, 0.0000016
$\sigma_r^2$	Process error variance for recruitment	0.19
$p^{k,g}$	Proportion of gametes of sex k with	$p^{l,0} = l; p^{1,g} (g>0) = 0$
	construct copy number g	$p^{0,g}$ = calculated from eqn. S6
$O^{g}$	Proportion of offspring with construct	calculated from eqn. S7
	copy number g	
$\pi_k$	Expected proportions by sex k in the	Scenario specific, see text
	wild population	